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AAR60224
     AAR60224 standard; protein; 145 AA.
ID
ХX
AC
     AAR60224;
XX
DT
     25-MAR-2003
                  (revised)
     30-MAR-1995 (first entry)
DT
XX
     Immunogenic fragment of influenza haemagglutinin.
DΕ
XX
     Antigen; immunogen; vaccine; influenza; fusion protein; immunity;
KW
KW
     haemagglutinin; neuraminidase; flu.
XX
OS
     Influenza virus.
XX
ΡN
     WO9417826-A1.
XX
PD
     18-AUG-1994.
XX
     01-FEB-1994;
                    94WO-US001149.
PF
XX
                    93US-00013415.
PR
     01-FEB-1993;
     18-AUG-1993;
                    93US-00108914.
PR
     05-NOV-1993;
                    93US-00149150.
PR
XX
     (SMIK ) SMITHKLINE BEECHAM CORP.
PA
XX
ΡI
     Shatzman A, Kane J, Scott M, Dillon S;
XX
     WPI; 1994-279392/34.
DR
XX
     Vaccines against multi strain influenza virus infection - protect against
PT
     influenza A and B.
PT
XX
     Disclosure; Page 117-118; 151pp; English.
PS
XX
CC
     A vaccine comprising an immunogenic fragment of the HA2 subunit of the
     influenza haemagglutinin (HA) protein from type A subtype IV and type B
CC
     IV may be used for stimulating protection in animals against injection
CC
     with influenza virus. The vaccine confers multi-strain immunity against
CC
CC
     strains IV A and IV B. The vaccines may be recombinantly produced,
     optionally as fusion proteins. This sequence is a fragment of the HA2
CC
CC
     subunit of haemagglutinin, corresponding to the amino acids at
CC
     approximately 77-221 of the HA2 subunit sequence. This sequence is
CC
     derived from the influenza H3 subtype. (Updated on 25-MAR-2003 to correct
CC
     PN field.)
XX
SO
     Sequence 145 AA;
                          100.0%; Score 221; DB 2; Length 145; 100.0%; Pred. No. 3.4e-20;
  Query Match
  Best Local Similarity
                                                   0;
           43; Conservative
                                 0; Mismatches
                                                      Indels
                                                                 0; Gaps
            1 IQDLEKYVEDTKIDLWSYNAELLVALENQHTIDLTDSEMNKLF 43
Qу
              1 IQDLEKYVEDTKIDLWSYNAELLVALENQHTIDLTDSEMNKLF 43
Db
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# **PCT**

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#### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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#### (57) Abstract

This invention provides vaccine compositions capable of conferring multi-strain immunity against influenza A and influenza B. This invention also provides methods of increasing expression and improving homogeneity of H3HA2 protein, fragments thereof, and fusion proteins containing same, as well as novel nucleotide sequences encoding these proteins and fragments.

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GA	Gabon				

#### VACCINAL POLYPEPTIDES

#### Field of the Invention

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The present invention relates generally to polypeptides useful in vaccine compositions and more specifically to vaccine compositions useful in providing immunity against influenza A and influenza B in an animal. The present invention also relates generally to a method of enhancing expression of polypeptides and, more specifically, to a method of enhancing influenza protein expression and homogeneity in *E. coli*.

#### Background of the Invention

Influenza virus infection causes acute respiratory disease in man, horses, swine and fowl, sometimes of pandemic proportions. Influenza viruses are orthomyxoviruses and, as such, have envelope virions of 80 to 120 nanometers in diameter, with two different glycoprotein spikes. Three types, A, B and C, infect humans. Type A viruses have been responsible for the majority of human epidemics in modern history, although there are also sporadic outbreaks of Type B infections. Known swine, equine, and avian viruses have mostly been Type A, although Type C viruses have also been isolated from swine.

The Type A viruses are divided into subtypes based on the antigenic properties of the hemagglutinin (HA) and neuraminidase (NA) surface glycoproteins. Within Type A, subtypes H1 ("swine flu"), H2 ("asian flu"), and H3 ("Hong Kong flu") are predominant in human infections. In swine, the predominant influenza A subtypes are H1 and H3; in horses, H3 and H7; and in avians, H5 and H7. Presently only one Type B virus has been identified, with no subtypes.

Genetic "drift" or "shift", i.e., rapid and unpredictable change in the antigen, occurs at approximately yearly intervals, and affects antigenic determinants in the HA and NA proteins. Therefore, it has not been possible to prepare a "universal" influenza virus vaccine using conventional killed or attenuated viruses, that is, a vaccine which is non-strain specific. Recently, attempts have been made to prepare such universal, or semi-universal, vaccines from reassortant viruses prepared by crossing different strains. More recently, such attempts have involved recombinant DNA techniques focusing primarily on the HA protein.

There remains a need in the art for vaccine formulations and compositions capable of inducing protective responses in animals against influenza viruses.

The expression of recombinant proteins in bacterial systems, particularly *E. coli*, is highly desirable because it can be used to produce large amounts of the desired proteins relatively inexpensively. However, high level expression of several eukaryotic proteins in *E. coli* has not been achieved for reasons including, among others, unfavorable codon usage and toxicity of the gene product [U. Brinkmann *et al.*, Gene, 85:109-114 (1989)]. Methods of overcoming these impediments to high-level expression in bacteria have been described, but are not universally applicable.

For example, Brinkmann et al., cited above, described low-level expression of certain genes, such as human tissue-type plasminogen activator or gp41 of human immunodeficiency virus, which the authors attributed to the presence of the rare triplets AGA and AGG which encode arginine (Arg) in unexpectedly high amounts in the gene (3.2%). However, other eukaryotic genes, such as the NS1 gene of influenza virus, contain greater than 3% of such triplets yet express at high levels in E. coli [Young et al, Proc. Natl. Acad. Sci., 80:6105-6109 (1983)].

Another group, Spanjaard et al., Nucl. Acids Res., 18(17):5031-5036 (1990) describe a translation shift in about 50% of ribosomes after tandem (double) AGA and AGG codons in cloned tRNA genes, but observed no frame shifts following single AGG or AGA codons. The authors attribute this frame shift to tRNA depletion. There also remains a need in the art for improved methods of producing vaccinal polypeptides capable of inducing protective responses in animals against influenza viruses.

#### 25 Summary of the Invention

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The present invention provides compositions containing, and methods for use of a protein which is capable of inducing protection in animals and avians against challenge with more than one strain of influenza Type A and influenza Type B.

Thus, one aspect of the invention provides a DNA sequence encoding a modified purified recombinant protein. The DNA sequence of the invention encodes a modified protein sequence derived from the HA2 subunit of a selected hemagglutinin (HA) protein. In one embodiment, the sequence is derived from an H3N2 subtype influenza virus. These H3N2 fusion proteins are capable of inducing T cell responses in the absence of neutralizing antibodies. In another embodiment, a DNA sequence of this invention encodes a modified protein sequence derived from the HA2 subunit from a Type B influenza virus. Still further embodiments include

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DNA sequences obtained as described for the two above viruses, where the sequences are derived from other Type A influenza strains infecting animals as well as humans. Such viruses include, without limitation, Type A subtypes of H1, H2, H3, H4, H5, H6 and H7.

In another aspect, the invention provides a DNA sequence encoding a recombinant fusion protein, in which the desired Type A subtype HA2 subunit sequence or a portion thereof, is fused in frame to another protein or protein fragment capable of enhancing expression of the fusion protein. One embodiment includes the H3N2 subtype HA2 subunit sequence described above fused in frame to another protein or fragment capable of enhancing expression thereof. Another embodiment of such a fusion protein comprises a Type B HA2 sequence, described above, or a portion thereof, fused in frame to another protein or protein fragment capable of enhancing expression of the fusion protein. Additionally, other Type A subtype HA2 sequences can be similarly used. It is desirable that this fusion partner protein be an influenza protein sequence or fragment thereof.

In still another aspect, a protein encoded by a DNA sequence of the invention is provided. The protein may be a protein sequence derived from the HA2 subunit of an HA protein from a selected Type A subtype virus. Desirably the subtype virus is an H3N2. In another embodiment, the protein may be derived from the HA subunit of a Type B influenza virus. Other embodiments include H5 or H7 subtypes. Additionally, preferred embodiments include fusion proteins comprising a protein sequence derived from the HA2 subunit of an HA protein from a Type A virus, e.g., an H3N2 subtype, or from a Type B virus fused in frame to a selected influenza sequence. The proteins of this invention are particularly useful in inducing protection in mammals, especially humans, against challenge by Type B or an H3N2 subtype of influenza A. The proteins employing other Type A subtypes, e.g., H5 and H7, are useful in inducing protection in animals against influenza viruses.

In another aspect, the invention provides a method of recombinantly producing the fusion proteins of the invention, and a method of purifying the same.

In a further aspect, the invention provides a vaccine composition containing a purified protein of the invention, as described above. Such a vaccine composition may include a fusion protein of the invention. In other embodiments of the invention, the vaccine compositions contain an H3HA2 protein of the invention and other influenza antigens; a Type B HA2 protein of the invention and other influenza antigens; or both an H3HA2 protein, a BHA2 protein and other influenza antigens. In a preferred embodiment for human use, a combination vaccine of the invention will contain an H3HA2 and a BHA2 protein of the invention in

combination with influenza antigens derived from the other Type A influenza virus subtypes, H1 and H2. An embodiment for use in animals may contain an H5HA2 or H7HA2 protein, among others.

A further aspect of this invention is a method for inducing in an animal protection against influenza Type A, influenza Type B, influenza Type C, or combinations thereof, which comprises internally administering to the animal an effective immunogenic amount of a vaccine composition of the present invention.

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Still a further aspect of this invention is a method for inducing in an animal protection against multiple strains of influenza Types A and B which comprises internally administering to the animal an effective immunogenic amount of a vaccine composition of the present invention.

In another aspect, the present invention provides a method of enhancing in *E. coli* the expression of influenza vaccinal proteins characterized by a naturally-occurring amino acid pattern comprising Arg-Arg-Xaa-Xaa-Arg [SEQ ID NO:8]. In this pattern, Arg is arginine, Xaa is any amino acid, and at least one of the arginines in the naturally-occurring sequence is encoded by the rare nucleic acid triplets AGG or AGA.

In one embodiment, the method of the invention involves mutating one or more of these AGG or AGA codons to a preferred arginine codon and expressing the mutated sequence in *E. coli*. Surprisingly, it has been found that this modification, which does not result in a change in the encoded amino acid sequence, can increase the expression and homogeneity of an influenza protein in *E. coli* significantly.

In another embodiment, the method of this invention involves increasing the expression of the above-identified proteins by inserting into the host cell tRNA molecules capable of translating the native rare arginine codons. Thus, the *E. coli* host cells are modified such that they are capable of efficiently translating the rare, native arginine codons.

In another aspect, the present invention provides novel nucleic acid sequences of influenza proteins which contain the nucleotide sequence CGn-CGn-Xaa-Xaa-CGn, where n represents a nucleotide selected from the group consisting of T, C, A or G [SEQ ID NO:9], in place of the native nucleotide sequence AGr-AGr-Xaa-Xaa-AGr, where r represents the nucleotides A or G [SEQ ID NO:10]. When expressed in E. coli, these sequences result in increased expression of the encoded protein as compared to the native sequence.

In still another aspect, the invention provides the novel modified nucleic acid sequences described above fused in the same reading frame to another

DNA sequence encoding a polypeptide or protein, i.e., a fusion partner, which may further enhance the expression of, or immunogenicity of, the encoded influenza protein. It is desirable that the fusion partner be an influenza protein sequence or fragment thereof.

Other aspects and advantages of the present invention are described further in the following detailed description of the preferred embodiments thereof.

#### Brief Description of the Drawings

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Fig. 1 illustrates the nucleic acid sequences of the HA2 portions of (a) A/Udorn [SEQ ID NO: 1], (b) A/Victoria [SEQ ID NO: 3], (c) A/PR/8/34 [SEQ ID NO: 5], and (d) a consensus sequence [SEQ ID NO: 7]. Dashes indicate the same nucleotide as the consensus sequence. Different nucleotides from that of the consensus sequence are reported in lower case letters. Dots indicate no corresponding nucleotide when compared to the consensus sequence.

Fig. 2 illustrates the nucleic acid and amino acid sequences of H3C13, NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> fusion protein [SEQ ID NO: 9 & 10], with the mutant nucleic acid sequences of H3C13mut5855 [SEQ ID NO: 58] illustrated above the sequence of the unmodified H3HA2 portion.

Fig. 3 illustrates the nucleic acid and amino acid sequences of the NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> fusion protein [SEQ ID NO: 11 & 12].

Fig. 4 illustrates the nucleic acid and amino acid sequences of the Type B fusion protein, NS1<sub>(1-42)</sub>HA2<sub>(41-223)</sub>. [SEQ ID NO: 13 & 14].

Fig. 5 illustrates the pOTS208NS1BLmut2 vector nucleic acid sequences [SEQ ID NO: 54] encoding the amino acid sequences [SEQ ID NO: 55] of the mutant NS(1-81)BLHA2(1-223)(met-leu) fusion protein, with the nucleic acid sequences of the coding region NS(1-81)BLHA2(1-223) [SEQ ID NO: 56] and native amino acid sequences [SEQ ID NO: 57], which include a Met in amino acid position 98, illustrated above the modified BLHA2 sequences.

Fig. 6 illustrates the nucleic acid [SEQ ID NO:17] and amino acid [SEQ ID NO:18] sequences of the H1N1 fusion protein, NS1<sub>(1-81)</sub>HA2<sub>(65-222)</sub>, also known as flu D.

Fig. 7 illustrates the naturally-occurring nucleic acid sequence [SEQ ID NO:1] and corresponding amino acid sequence [SEQ ID NO:2] of the HA2 portion of the H3N2 virus, A/Udorn.

Fig. 8 illustrates the naturally-occurring nucleic acid sequence [SEQ ID NO:3] and corresponding amino acid sequence [SEQ ID NO:60] of the HA2 portion of the H3N2 virus, A/Victoria.

#### Detailed Description of the Invention

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The present invention provides novel proteins, DNA sequences, pharmaceutical vaccine compositions, and methods of use thereof for conferring protection in vaccinated mammals against one strain, or desirably multiple strains, of influenza viruses. The proteins and vaccine compositions of the present invention demonstrate the ability to stimulate or produce a protective immune response which is capable of recognizing an influenza virus or influenza virus-infected cells and protecting the vaccinated mammal against disease caused thereby. This protective response is desirably a T cell response, produced in the substantial absence of vaccine-induced neutralizing antibody.

While the proteins and DNA sequences specifically described herein are directed to the H3HA2 and BHA2 sequences originating from viral strains to which humans are susceptible, it is expected that similar sequences and molecules can be prepared for veterinary applications. For example, selected HA2 sequences obtained from Type A viral strains, e.g., H5HA2, H7HA2 and other strains of interest may be obtained following the teachings described herein for the exemplified H3HA2 and BHA2 sequences. One of skill in the art should understand that this invention is not limited to the exemplified protein and DNA sequences, even though the following disclosure is limited to the two latter sequences for simplicity. Such additional viral HA2 subunits are expected to share the biological characteristics of the exemplified sequences.

Thus, this invention provides a protein or fragment thereof characterized by an amino acid sequence derived from the HA2 subunit of an HA protein, e.g., from a H3N2 subtype virus. As used herein, a "fragment" of the HA2 subunit is an amino acid sequence derived from the HA2 subunit which is characterized by having an immunogenic determinant of the HA2 subunit. Such a fragment is desirably at least about 8 amino acids in length.

The H3 proteins of the invention are capable of inducing T helper cells, particularly cytotoxic T lymphocytes, in the absence of neutralizing antibodies. Among H3N2 subtype strains of influenza A include A/Udorn and A/Victoria viruses. Other H3N2 virus strains of influenza A may also produce HA proteins for use in vaccine compositions according to this invention. Fig. 1 compares the nucleic acid sequences of the HA2 portions of the A/Udorn [SEQ ID NO: 1] and A/Victoria [SEQ ID NO: 3] strains with the nucleic acid sequence of an H1N1 subtype virus, A/PR/8/34 [SEQ ID NO: 5]. A consensus sequence [SEQ ID NO: 7] was computer generated, and may likewise be useful in producing proteins

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according to this invention. This consensus sequence [SEQ ID NO: 7] can be constructed by a commercially available computerized sequence analysis program, such as Genetics Computers Group [University of Wisconsin].

Proteins according to this invention may include unfused HA2 subunits of the influenza A viruses, particularly H3N2 subtype. For example, in one embodiment, a protein of the invention contains amino acids 1-221 of a selected H3HA2 subunit. In another embodiment, a protein of the invention contains amino acids 77-221 of the H3HA2 subunit. Other fragments of this HA2 amino acid sequence characterized by the ability to stimulate similar immunological activity in an immunized animal are also encompassed by this invention.

Proteins of this invention also include fusion proteins comprising a protein sequence derived from the HA2 subunit of an HA protein from a Type A virus, e.g., an H3N2 subtype virus, fused in frame to another protein or protein fragment capable of enhancing expression of the fusion protein. It is desirable that this fusion "partner" protein be an influenza protein sequence or fragment thereof derived from the same or another strain of influenza virus as the HA protein or protein fragment. Preferably, this fusion partner protein is all or a portion of the influenza virus NS1 protein or an HA2 subunit protein.

In the embodiments exemplified herein, the NS1 portion of the fusion protein is derived from an H1N1 subtype virus, A/PR/8/34. For example, in one embodiment, the NS1 portion may comprise amino acid residues 1 to 42 of H1NS1. In another embodiment the NS1 portion may comprise amino acid residues 1 to 81 of the selected virus. The HA2 fragment may alternatively be fused to a portion of the NS1 peptide derived from a selected Type A virus, e.g., an H3 subtype virus (H3HA2), or a Type B (BHA2) virus.

However, other non-influenza fusion proteins may also produce desirable fusion proteins with the H3N2, or other Type A, or Type B protein or portion thereof. Thus, in still another alternative embodiment, as discussed below, the HA2 fragment may be fused to any peptide capable of enhancing its expression in the host cell selected. One of skill in the art may readily select a fusion "partner" protein or fragment taking into account the desired host cell and utilizing the teachings herein. The fusion proteins of the present invention are not limited by the selection of the "partner" protein or fragment to which the HA2 fragment is fused.

In yet another embodiment, the present invention provides a modified protein containing a portion of the HA2 subunit of a Type B influenza virus. Currently, the preferred human virus strain is B/Lee/40. However, the vaccinal proteins of this invention are not limited to this Type B strain, and other

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strains infecting other species, or other as yet unidentified Type B virus strains, may be used to produce the HA2 protein. These Type B HA2 proteins may be fused to a fusion "partner" protein or protein fragment, as described above for the H3HA2 proteins of this invention, or remain unfused.

In the construction of a fusion protein according to this invention, a linker sequence may optionally be inserted between the two fused sequences, i.e., between the NS1 portion and the HA2 portion. This optional linker may provide space between the two linked sequences. Alternatively, this linker sequence may encode, if desired, a polypeptide which is selectively cleavable or digestible by conventional chemical or enzymatic methods. For example, the selected cleavage site may be an enzymatic cleavage site, including sites for cleavage by a proteolytic enzyme, such as enterokinase, factor Xa, trypsin, collagenase, and thrombin. Alternatively, the cleavage site in the linker may be a site capable of being cleaved upon exposure to a selected chemical, e.g., cyanogen bromide or hydroxylamine. The cleavage site, if inserted into a linker useful in the fusion sequences of this invention, does not limit this invention. Any desired cleavage site, of which many are known in the art, may be used for this purpose.

A presently preferred example of an H3 fusion protein of this invention is NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 10], which comprises the first 81 amino acids of NS1 fused to amino acids 1 to 221 of the H3HA2 subunit (amino acids 1-221). (Fig. 2) Another exemplary fusion protein, NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> [SEQ ID NO: 12], comprises the first 81 amino acids of NS1 fused to amino acids 77 to 221 of the truncated H3HA2 subunit. (Fig. 3)

A present preferred example of a Type B fusion protein of this
invention is NS1<sub>(1-42)</sub>BHA2<sub>(41-223)</sub> [SEQ ID NO: 14], which comprises the first
42 amino acids of NS1 fused to amino acids 41 to 223 of the truncated BHA2
subunit. (Fig. 4) Another fusion protein of this invention is NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub> [SEQ ID NO: 57], which contains the first 81 amino acids of NS1 fused to
amino acids 1 to 223 of the BHA2 subunit. (Fig. 5) Another preferred fusion
protein of the invention is NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub>(met-leu) SEQ ID NO: 55, which
contains the same amino acid sequence as NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub>, with the
exception that the internal methionine residue at position 98 of the fusion protein
has been changed to a leucine. (Fig. 5)

These proteins, fusion proteins, and similar proteins encoded by the below-described DNA sequences are referred to collectively herein as H3HA2 proteins.

The NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> protein [SEQ ID NO: 10] of the invention has a three-dimensional structure which is substantially similar to that of the NS1<sub>(1-81)</sub>HA2<sub>(1-222)</sub> protein [SEQ ID NO: 16] derived from the H1N1 subtype virus (C13). However, the amino acid sequence of the NS1(1-5 81)H3HA2(1-221) protein [SEQ ID NO: 10] has only approximately 50% homology with the amino acid sequence of C13 protein [SEQ ID NO: 16]. Additionally, as illustrated in Fig. 1, the nucleic acid sequence of the H3HA21-221 protein derived from A/Udorn (nucleotides 25-560 from that virus) [SEQ ID NO: 1] has only approximately 60% homology with the nucleic acid sequence of the H1HA2<sub>1-222</sub> protein derived from strain A/PR/8/34 (nucleotides 1872-2407 from 10 A/PR/8/34) [SEQ ID NO: 5]. However, the nucleic acid sequence of H3HA21-221 from A/Udorn (nucleotides 1-499 of A/Udorn) [SEQ ID NO: 1] has approximately 99% homology with the nucleic acid sequence of H3HA2<sub>1-221</sub> from A/Victoria/H3/75 (nucleotides 1226-1725 of A/Victoria) [SEQ ID NO: 3] [Fiers et al, Cell, 19:683-696 (1980)]. 15

Analogs of the HA2 peptides from a Type A virus, e.g., an H3, or Type B viruses, included within the definition of this invention, include truncated polypeptides (including fragments) and HA2 polypeptides, e.g. mutants that retain the epitopes and thus the biological activity of HA2. It is anticipated that, because the NS1 portion of the fusion peptide provides a means of expressing the protein at high levels and does not appear to play as significant a role in the immunological responses to the HA2 fusion proteins as does the HA2 portion, any number of analogs of this fusion partner can be made.

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Typically, the analogs of the HA2 peptides and/or the fusion partner differ by only 1 to about 4 codon changes. Other examples of analogs include polypeptides with minor amino acid variations from the natural amino acid sequence of HA2; in particular, conservative amino acid replacements. Conservative replacements are those that take place within a family of amino acids that are related in their side chains. Genetically encoded amino acids are generally divided into four families: (1) acidic = aspartate, glutamate; (2) basic = lysine, arginine, histidine; (3) non-polar = alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polar = glycine, asparagine, glutamine, cysteine, serine, threonine, tyrosine. Phenylalanine, tryptophan, and tyrosine are sometimes classified jointly as aromatic amino acids. For example, it is reasonable to expect that an isolated replacement of a leucine with an isoleucine or valine, an aspartate with a glutamate, a threonine with a serine, or a similar conservative replacement of an amino acid with a structurally related amino acid will not have a

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significant effect on its activity, especially if the replacement does not involve an amino acid at an epitope of the HA2 polypeptide. The construction of such analogs, given the description herein and conventional methods of protein modification known to one of skill in the art, are believed to be encompassed by this invention.

Currently, it is theorized that the HA2 portion of the fusion peptide (e.g., H3HA21-221, H3HA277-221 and BHA241-223) confers the majority of the necessary epitopes for antibody binding or T cell (particularly CTL) targeting. Once these epitope sequences are precisely identified, portions of the HA2 sequence which are not part of these epitopes may be altered without significantly affecting the bioactivity of the fusion protein.

The present invention also encompasses DNA sequences of this

invention encoding the above-described proteins and fusion proteins, the sequences characterized by having an immunogenic determinant of a modified HA2 subunit of an HA protein, derived from a Type A virus, e.g., an H3 subtype, or Type B virus. Other DNA sequences of this invention encode such HA2 subunits, optionally fused to a DNA sequence encoding a protein or peptide which is capable of enhancing expression of the protein in a selected host cell. For example, the consensus sequence illustrated in Fig. 1(d) may provide a source of HA2 DNA. The currently preferred embodiment provides a DNA sequence encoding a Type A virus, e.g., an H3 or Type B HA2 protein or fragment thereof fused in frame to a DNA sequence encoding a portion of the nonstructural influenza protein 1 (NS1).

Coding sequences for the HA2, NS1, and other viral proteins of influenza virus can be prepared synthetically or can be derived from viral RNA or from available cDNA-containing plasmids by known techniques. For example, in addition to the above-cited references, a DNA coding sequence for HA from the A/Japan/305/57 strain was cloned, sequenced and reported by Gething et al, Nature, 287:301-306 (1980). An HA coding sequence for strain A/NT/60/68 was cloned as reported by Sleigh et al, and by Both et al, in Developments in Cell Biology, Elsevier Science Publishing Co., pages 69-79 and 81-89, respectively, (1980). An HA coding sequence for strain A/WSN/33 was cloned as reported by Davis et al, Gene, 10:205-218 (1980); and by Hiti et al, Virology, 111:113-124 (1981). An HA coding sequence for fowl plague virus was cloned as reported by Porter et al and by Emtage et al, both in Developments in Cell Biology, cited above, at pages 39-49 and 157-168. Also, influenza viruses, including other strains, subtypes, and types are available from clinical specimens and from public depositories, such as the American Type Culture Collection (ATCC), Rockville, Maryland, U.S.A.

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Allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) of DNA sequences encoding the H3HA2 or BHA2 protein sequences are also included in the present invention, as well as analogs or derivatives thereof. Similarly, DNA sequences which code for H3 or other Type A or Type B HA2 proteins of the invention but which differ in codon sequence due to the degeneracies of the genetic code or variations in the DNA sequence encoding H3HA2, other Type A or BHA2 proteins which are caused by point mutations or by induced modifications to enhance the activity, half-life or production of the peptide encoded thereby are also encompassed in the invention. Suitably, this invention provides certain silent mutations to the coding sequences for NS1(1-81)H3HA2(1-221), which have been found to increase expression yields. See Fig. 2. Further, the NS1<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub>(1-81)</sub>BHA2<sub></sub> 223)(met-leu)-encoding sequence, BC13mut2, in addition to modifying the codon encoding amino acid position 98 of the fusion protein (position 17 of the HA2 portion), contains a number of silent modifications designed to increase protein expression. See Fig. 5.

Also covered by this invention are DNA sequences which hybridize under stringent conditions with the DNA sequences encoding the HA2 subunit proteins, e.g., H3HA2 or BHA2 proteins, of this invention. DNA sequences which hybridize under non-stringent conditions with the disclosed sequences, but which encode proteins or fragments retaining the biological activities of the H3HA2 or BHA2 proteins, are also included in this invention. Typical conditions for stringent or non-stringent hybridization are known to those of skill in the art. [See, e.g., Sambrook et al, Molecular Cloning. A Laboratory Manual, 2nd edition, Cold Spring Harbor Laboratory, NY (1989)].

The fusion proteins of the invention may be prepared by conventional genetic engineering and recombinant techniques known to those of skill in the art. Similarly, the proteins may be purified from expression in host cell or vector systems by conventional means.

Preferably, however, the recombinantly-produced fusion proteins of the invention are purified as described herein. Generally, method of purification involves (step 1) the isolation of the proteins, (step 2) enzymatic digestion and extraction, (step 3) urea extraction, (step 4) solubilization, reduction, and DEAE chromatography, (step 5) reverse phase chromatography, (step 6) precipitation, and (step 7) desalting and preparation of the final product. More specifically, the host cells containing the fusion proteins are disrupted, either chemically or by mechanical means. Preferably the cells are lysed by osmotic shock. Following

centrifugation, the resulting pellet (P1) is subjected to nuclease digestion extraction and centrifuged to yield pellet 2 (P2). A second extraction step is then performed using urea (pH 6) and the mixture centrifuged to yield pellet 3 (P3). P3 is then solubilized and reduced. Preferably, solubilization is performed using urea at pH 12.5 and reduction is via DTT DEAE chromatography followed by SDS elution. The resulting DEAE product is further reduced, preferably using DTT, and subjected to reverse phase chromatography. The reverse phase product is then precipitated by adjusting to pH 6 and centrifuged. The precipitated product is resolubilized, preferably with urea at pH 12.5, and subjected to G25 chromatography. The resulting G25 product is then filtered (e.g. with a 0.2 micron filter) to yield the final product. Further details of this method are provided in Example 17 below.

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Systems for cloning and expression of the vaccinal polypeptide of this invention in various microorganisms and cells, including, for example,  $\underline{E.\ coli}$ ,  $\underline{Bacillus}$ , Streptomyces, Saccharomyces, mammalian and insect cells, are known and available from private and public laboratories and depositories and from commercial vendors. The preferred host is  $\underline{E.\ coli}$  because it can be used to produce large amounts of desired proteins safely and cheaply. To circumvent the requirement of ampicillin for plasmid selection in production fermentations, a desirable method of production employs an alternative expression system in which the  $\beta$ -lactamase coding sequence is wholly or partially replaced by a coding sequence for an alternative selectable marker such as, for example, kanamycin or chloramphenicol.

Thus, the polypeptide employed in the presently preferred embodiment is preferably expressed in <u>E. coli</u>. A suitable strain, LW14, has the following genotype: galE::Tn10\(\lambda\)CI857 bio- uvrB-; phenotypically, strain LW14 requires biotin for growth, is sensitive to UV light and DNA damaging agents, and cannot use galactose as a carbon source. Construction of this strain is described in the examples below.

To aid in expression of the H3 or other Type A subunit or Type B HA2 peptides or fusion protein described above, these protein sequences or fragments thereof may also be fused to a polypeptide capable of enhancing expression of these fragments in the selected host system. Ordinarily, such a peptide would contain a leader sequence fragment that provides for secretion of the Type A subunit fragment, e.g., the H3HA2 fragment, or Type B HA2 fragment in the host cell. The leader sequence fragment typically encodes a signal peptide comprised of hydrophobic amino acids which direct the secretion of the protein from the cell. There may be processing sites encoded between the leader sequence

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and the Type A subtype or Type B HA2 fragment that can be cleaved either in vivo or in vitro. Alternatively, a promoter sequence may be linked directly with the DNA molecule encoding the HA2 fragment. Such polypeptides, promoter and leader sequences are known to those of skill in the art and may be readily selected for expression in the selected host.

Construction of expression systems, including expression vectors and transformed host cells are thus within the art. See, generally, methods described in standard texts, such as Sambrook et al, Molecular Cloning A Laboratory Manual, 2d edit., Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1989). The present invention is therefore not limited to any particular expression system or vector, nor to any particular purification process from cell lysates or cell medium.

The proteins and fusion proteins of this invention may be employed in vaccine compositions. Pharmaceutical vaccine compositions of this invention, therefore, contain an effective immunogenic amount of a selected HA2 protein, e.g., H3HA2 or BHA2 protein, of the invention in admixture with a suitable adjuvant in a nontoxic and sterile pharmaceutically acceptable carrier.

Suitable carriers for vaccine use are well known to those of skill in the art. However, exemplary carriers include sterile saline, lactose, sucrose, calcium phosphate, gelatin, dextrin, agar, pectin, peanut oil, olive oil, sesame oil, squalene, and water. Additionally, the carrier or diluent may include a time delay material, such as glyceryl monostearate or glyceryl distearate alone or with a wax. Optionally, suitable chemical stabilizers may be used to improve the stability of the pharmaceutical preparation. Suitable chemical stabilizers are well known to those of skill in the art and include, for example, citric acid and other agents to adjust pH, chelating or sequestering agents, and antioxidants.

While any aluminum adjuvant may be used in the vaccine compositions of this invention, two desirable adjuvants are available commercially, i.e., REHSORPTAR<sup>TM</sup> adjuvant [Armour Pharmaceuticals, Kankakee, IL] and REHYDRAGEL<sup>TM</sup> adjuvant [Reheis Chemical Co., Berkeley Heights, NJ]. These products are aluminum hydroxide gels which contain approximately 2% w/v Al<sub>2</sub>O<sub>3</sub>, which is equivalent to approximately 10.6 mg/ml Al<sup>+3</sup>.

Vaccine compositions of this invention may employ an immunogenic amount of a purified recombinant protein as described above. A preferred embodiment of the vaccine of the invention is composed of an aqueous suspension or solution containing the recombinant HA2 protein molecule, e.g., H3HA2 or BHA2, together with an adjuvant, preferably an aluminum, most preferably aluminum hydroxide, buffered at physiological pH, in a form ready for injection. A

preferred protein for use in these vaccine compositions includes a protein comprising amino acid residues 1 to 81 from NS1 fused to C-terminal amino acid residues 1-221 from the hemagglutinin subunit 2 (HA2) from influenza A, subtype H3N2. Another preferred vaccine composition of this invention employs a purified recombinant protein made up of amino acid residues 1 to 81 from NS1 fused to amino acid residues 77-221 of the HA2 from influenza A, subtype H3N2. Still another preferred vaccine composition of this invention employs a purified recombinant protein made up of amino acid residues 1 to 42 fused to amino acid residues 41-223 of the HA2 from influenza B.

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Vaccine compositions of the invention may also employ an immunogenic amount of a recombinant protein of the invention in combination with other influenza antigens. Suitable influenza antigens for combination in a vaccine composition with the proteins of this invention may be derived from Type A, H1 subtype viruses and may include the recombinant fusion proteins described in detail in copending U. S. Patent Application Ser. No. 07/387,200, filed July 28, 1989 and its corresponding European Patent Application No. 366, 238, published May 2, 1990; and in co-pending U. S. Patent Application Ser. No. 07/387,558, filed July 28, 1989 and its corresponding European Patent Application No. 366,239, published May 2, 1990. The C13 protein (NS1(1-81)HA2(1-222)) [SEQ ID NO: 15 & 16], D protein (NS1(1-81)HA2(65-222)) [SEQ ID NO: 17 & 18] and other fusion proteins derived from the H1N1 influenza virus subtype and the recombinant expression and purification thereof are disclosed in detail in these applications, and in the parent applications identified in this application, all of which are incorporated by reference herein.

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More specifically, suitable H1 subtype immunogenic proteins include C13 (NS1<sub>(1-81)</sub>-D-L-S-R-HA2<sub>(1-222)</sub>) [SEQ ID NO: 15 & 16], D (NS1<sub>(1-81)</sub>-Q-I-P-HA2<sub>(65-222)</sub>) [SEQ ID NO: 17 & 18], C13 short (NS1<sub>(1-42)</sub>-M-D-L-S-R-HA2<sub>(1-222)</sub>) [SEQ ID NO: 19 & 20], D short (NS1<sub>(1-42)</sub>-M-D-H-M-L-T-S-T-R-S-HA2<sub>(66-222)</sub>) [SEQ ID NO: 21 & 22], A (NS1<sub>(1-81)</sub>-Q-I-P-HA2<sub>(69-222)</sub>) [SEQ ID NO: 23 & 24], C (NS1<sub>(1-81)</sub>-Q-I-P-HA2<sub>(81-222)</sub>) [SEQ ID NO: 25 & 26], ΔD (NS1<sub>(1-81)</sub>+HA2<sub>(150-222)</sub>) [SEQ ID NO: 27], Δ13 (NS1<sub>(1-81)</sub>-D-L-S-R-HA2<sub>(1-70)</sub>-S-C-L-T-A-Y-H-R) [SEQ ID NO: 28], M (NS1<sub>(1-81)</sub>-Q-I-P-HA2<sub>(65-196)</sub>-G-G-S-Y-S-M-E-H-F-R-W-G-K-P-V) [SEQ ID NO: 29], ΔM (NS1<sub>(1-81)</sub>-Q-I-P-HA2<sub>(65-200)</sub>-L-V-L-L) [SEQ ID NO: 31 & 32]. These H1N1 fusion proteins are described in published European Patent Application 366,238 and in copending U.S. Patent Application Ser. No. 07/751,896. Other suitable H1 proteins consist of

unfused polypeptides, such as H1HA2<sub>66-222</sub> [SEQ ID NO: 33 & 34] which is disclosed in co-pending U. S. Patent Application Ser. No. 07/751,898, incorporated herein by reference. Thus, one desirable combination vaccine to provide protection against Type A influenza contains NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> protein [SEQ ID NO: 9 & 10] of the invention, one or more proteins derived from subtype H1N1 as described above, and an aluminum adjuvant.

Preferably, a combination vaccine of the invention will contain an immunogenic amount of the H3 fusion protein of the invention in combination with immunogenic amounts of influenza antigens derived from the other Type A influenza virus subtypes, including among others, H1, H2, H3, H4, H5, H6; and H7, as well as a Type B fusion protein of the invention.

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A currently preferred combination vaccine of the invention contains the H3 subtype fusion protein NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 10], the B subtype fusion protein NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub>(met-leu) [SEQ ID NO: 55], and the H1 subtype fusion protein NS1<sub>(1-81)</sub>HA2<sub>(65-222)</sub> [SEQ ID NO: 18]. Studies have shown that such a combination vaccine is protective against challenge with H1, H3 and Type B influenza viruses in mice.

Other preferred combination vaccines would include the NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> protein [SEQ ID NO: 12] or the NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub> [SEQ ID NO: 57] in combination with one or more additional influenza antigens derived from the type or subtype influenza viruses described above. These combination vaccines will protect against influenza infections caused by both Type A and Type B influenza viruses. Still other combination vaccine compositions will employ other proteins described herein.

The compositions of the present invention are advantageously made up in a dose unit form adapted for the desired mode of administration. Each unit will contain, at a minimum, a predetermined quantity of the selected HA2 subunit protein, e.g., H3HA2 protein and/or BHA2 protein, and adjuvant calculated to produce the desired therapeutic effect in optional association with a pharmaceutical diluent, carrier or vehicle.

Dosage protocol can be optimized in accordance with standard vaccination practices. Typically, the vaccine will be administered intramuscularly, although other routes of administration may be used, such as intradermal. It is expected that an effective immunogenic amount of a protein, fusion protein or combination of proteins of this invention for average adult humans is in the range of 1 to 1000 micrograms. Another desirable immunogenic amount ranges between 50

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to 500 micrograms. Most preferably, the proteins of the invention are in admixture with the same amount or more adjuvant to form a vaccine composition.

While the proteins described herein have been particularly developed for use in humans (e.g., the H3HA2 and BHA2 sequences), it is expected that due to species cross-reactivity, these vaccines will be useful in other animals, particularly swine. Additionally, similar molecules can be prepared for equine and avian veterinary applications utilizing the HA2 proteins from other strains to which animals are susceptible. Combination vaccines for use in swine would preferably include protections against both H1 and H3 viruses. Combination vaccines for use in equine would preferably include protection against H3 and H7 viruses. Combination vaccines for use in avian species would preferably confer protection against H5 and H7 viruses. Appropriate dosages can be determined by one skilled in veterinary medicine.

It will be understood, however, that the specific effective immunogenic amount for any particular patient will depend upon a variety of factors including the age, general health, sex, and diet of the vaccinee; the species of the vaccinee; the time of administration; the route of administration; interactions with any other drugs being administered; and the degree of protection being sought.

The vaccine can be administered initially in late summer or early fall and can be readministered two to six weeks later, if desirable, or periodically as immunity wanes, for example, every two to five years. Of course, as stated above, the administration can be repeated at suitable intervals if necessary or desirable.

The present invention provides methods for producing enhanced expression and improved homogeneity of influenza viral proteins and polypeptides in *E. coli*. Also provided are novel modified nucleotide sequences which encode these influenza proteins and are useful in the methods of production.

Preferably, the influenza proteins or polypeptides produced according to the invention include the complete HA2 protein of the hemagglutinin antigen (HA) of a selected H3N2 influenza virus, a complete HA protein of an H3HA2 virus, fragments thereof, and fusion proteins containing the complete H3HA2 protein or desired fragments thereof fused in the same reading frame with a selected fusion partner polypeptide or protein. These proteins are characterized by having the native amino acid sequence pattern described above.

By the term "fragment" is meant a subunit of HA, or a span of contiguous amino acids from the complete protein capable of stimulating an antigenic or protective immunogenic response in an animal. A fragment may contain at least about 8 amino acids from the selected influenza protein, and can

contain up to the number of amino acids which make up the entire protein. When the term 'fragment' is used herein to modify a nucleotide sequence, it refers to nucleotide sequences which encode the above-defined amino acid fragments.

Native (or naturally-occurring) nucleotide sequences which encode certain influenza proteins are characterized by a nucleotide sequence pattern encoding the fragment Arg-Arg-Xaa-Xaa-Arg [SEQ ID NO:61]. Arg represents arginine and Xaa represents any amino acid in this formula. Hereafter, this five amino acid sequence is referred to as Formula I.

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Formula I sequences are typically encoded by native nucleotide sequences of the formula of codons AGr-AGr-Xaa-Xaa-AGr, where r represents the nucleotides A or G and Xaa represent any codon [SEQ ID NO:63]. Hereafter, this five codon nucleotide sequence is referred to as Formula II. Specifically, the native nucleic acid sequence encoding a subtype H3N2 influenza virus protein, fusion protein, or a fragment or subunit thereof, specifically the HA2 portions of H3N2 virus strains, is characterized by a Formula II sequence.

Among H3N2 subtype strains of influenza A characterized by this nucleotide fragment Formula II include the A/Udorn and A/Victoria viruses. Figs. 7 and 8 provide the native nucleic acid sequences of the HA2 portions of the A/Udorn [SEQ ID NO: 1] and A/Victoria [SEQ ID NO: 3] strains. Other H3N2 virus strains of influenza A may also provide native nucleotide sequences containing Formula II, which sequences are susceptible to the modifications described herein.

Additional examples of native nucleotide sequences encoding proteins whose expression may be enhanced according to this invention are those native sequences which encode certain fragments of influenza proteins including the fragment spanning amino acids 1 to about amino acids 221 of H3HA2 [Fig. 7 SEQ ID NO:2 and Fig. 8 SEQ ID NO:3]; the fragment spanning from about amino acid 77 to about amino acid 221 [Fig. 7 SEQ ID NO:69 and Fig. 8 SEQ ID NO:70], or other desirable fragments. Other desirable fragments of this H3HA2 amino acid sequence include those characterized by the ability to stimulate immunological activity in an immunized animal similar to that stimulated by use of the entire 221 amino acid sequence of H3HA2.

Nucleotide sequences encoding fusion proteins which contain fragments of the native nucleotide sequences encoding these influenza proteins or subunits, e.g., the fusion protein NS1(1-81)H3HA2(1-221) [SEQ ID NO:10], can also be characterized by the Formula II nucleotide sequence. Thus these fusion proteins are also desirable for enhanced expression according to the method of this invention.

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The inventors have discovered that when native nucleotide sequences of influenza proteins, which sequences comprise Formula II, are expressed in E. coli, a frame shift of one nucleotide after the third triplet in Formula II in the native sequence occurs, resulting in the increased translation of truncated proteins. It has been surprisingly found that by application of a method of the present invention, the expression and homogeneity of the influenza protein is increased significantly.

The methods of this invention involve enhancing the expression of proteins characterized by the amino acid pattern of Formula I, which proteins have a native nucleotide sequence of Formula II. According to one embodiment of the method of this invention, a native nucleotide sequence encoding a selected influenza protein or fragment, which sequence comprises Formula II, is modified by mutating one or more of the rare AGG or AGA arginine codons of Formula II to a preferred Arg codon. A preferred arginine codon for use in replacing a native AGA or AGG codon according to this invention is defined herein by the codons CGT, CGG, CGA and CGC. Of these codons, CGT and CGC are currently the most preferred. The modified influenza protein-encoding nucleotide sequence is then expressed in an E. coli expression system, resulting in enhanced expression in comparison to that obtained by expression of the native nucleotide sequence encoding the same protein in the same expression system.

The enhanced protein expression occurs even though the mutation does not result in a change in the encoded amino acid sequence of the protein. By the terms 'enhanced expression' or 'enhanced protein expression' is meant an expression level of at least 40% higher than the expression level of the protein encoded by the native, non-mutated nucleotide sequence comprising Formula II, when expressed in *E. coli*.

While not wishing to be bound by theory, the inventors believe that the enhanced expression levels are obtained because the silent mutation of the AGA or AGG to a preferred arginine codon in Formula II eliminates the frame shift mutation found in the unmutated nucleotides encoding these proteins, thus substantially reducing the production of truncated messages (proteins). It is believed that the resulting influenza proteins are more homogeneous when expressed in an *E. coli* expression system according to this invention.

In a second embodiment of the method of the invention, the expression of the proteins containing arginines encoded by the rare codons AGG and AGA (i.e. proteins encoded by amino acid and nucleotide sequences characterized by Formulae I and  $\Pi$ ) can be increased by inserting into the host in which expression is desired one or more genes for tRNA molecules which are

capable of properly translating the AGG and AGG arginine codons. Preferably the host cells are E. coli.

This method can be accomplished as follows. A gene for a tRNA molecule described above can be selected from among known gene sequences. The genes and tRNA molecules which can translate the rare Arg codons identified above are known and readily available to one of skill in the art. See, e.g., [P. Saxena and J. Walker, J. Bacteriol., 174(6):1956-1964 (Mar. 1992)].

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According to conventional techniques, these genes may be placed on a plasmid which will increase the copy number of these genes and therefore the tRNA molecules encoded by these genes. Alternatively, these sequences can be genetically engineered and placed on the host cell chromosome behind an appropriate promoter element in such a manner that the effective concentration of these tRNA molecules is increased inside the cell. Conventional texts describe the techniques useful in this method [See, e.g., Sambrook et al., Molecular Cloning. A Laboratory Manual. 2d edition, Cold Spring Harbor, New York (1989)].

The insertion of the tRNA genes into the host cell expressing the protein increases the concentration of these tRNA molecules inside the host cells which are naturally deficient for these tRNA molecules. This allows the host cells to translate these rare arginine codons in an efficient manner, eliminating the production of the truncated or lower molecular weight species of the fusion protein observed in the unmodified host cell. Thus, this method may be used to increase expression of a protein in host cells lacking sufficient amounts of the appropriate tRNA to permit efficient expression of the protein. Use of this method obviates the need to modify the sequences encoding the selected protein, and thus provides an alternative method to the first embodiment described above.

As another aspect of this invention novel modified nucleotide sequences are provided, which in *E. coli* expression systems, can be employed to produce the encoded influenza proteins, subunits, fragments and fusion proteins described above according to the first embodiment of the method of this invention. The proteins encoded by these nucleotides are produced at levels of expression enhanced over that of the native sequences, by about forty percent or more. The novel nucleotide sequences of the invention are characterized by comprising the nucleotide sequence CGn-CGn-Xaa-Xaa-CGn, where n represents a nucleotide selected from the group consisting of T, C, A or G [SEQ ID NO:62], in place of the Formula II fragment in the native nucleotide sequence encoding the selected influenza protein or fragment. The nucleotide fragment identified by the formula above is referred to herein for simplicity as Formula III.

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For example, a modified DNA sequence of the invention comprises the Formula III nucleotide sequence and may encode the amino acid sequences identified specifically above, e.g., Fig. 7 [SEQ ID NO:2], Fig. 8 [SEQ ID NO:3]; Fig. 7 [SEO ID NO:69] and Fig. 8 [SEQ ID NO:70], or other fragments.

In one example of the present invention, the nucleic acid sequence encoding the HA2 subunit protein which contains the native sequence of Formula II has been provided with three silent mutations, which have changed each of the three native arginine-encoding AGG codons each to a preferred arginine codon CGT. These codons encode amino acid numbers 123, 124 and 127 of the H3HA2 subunit protein of the A/Udorn strain identified in Fig. 7. The same codons (and amino acid numbers) are altered in the A/Victoria strain identified in Fig. 8 to provide another example of a modified nucleotide sequence according to this invention.

Thus, with reference to each of Figs. 7 and 8, the native nucleotide sequences encoding the HA2 subunit proteins of the aforementioned viruses [SEQ ID NO:1 and 60], are modified according to this invention at nucleotides 367, 370, and 379. At each of these nucleotide sites, the native A (adenine) is changed to a C (cytosine) and the native nucleotides at sites 369, 372 and 381 in each sequence are changed from a G (guanine) to a T (thymine), resulting in preferred Arg codons.

Other nucleotide sequences encoding the influenza vaccinal polypeptides described herein, or other such influenza proteins or subunits characterized by Formula II may be mutated into novel nucleotide sequences of this invention, i.e., by mutating Formula II into Formula III within those sequences using the first embodiment of the methods of this invention. The silent mutations described herein may be inserted at analogous regions in each nucleotide sequence.

The novel modified H3HA2 nucleotide sequences, whether alone or in association with a nucleotide sequence encoding a fusion partner of a fusion protein of the invention are useful in *E. coli* expression systems. The novel nucleotide sequences of the invention will also encode analogs of the H3HA2 peptides, such as truncated polypeptides (including fragments) and H3HA2 polypeptides, e.g. mutants that retain the epitopes and thus the biological activity of H3HA2. Where the nucleotide sequence encodes a fusion protein, it is anticipated that, because the non-HA2 fusion partner, e.g., NS1 as described below, the fusion peptide provides a means of expressing the protein at high levels and does not appear to play as significant a role in the immunological responses to the HA2 fusion proteins as does the HA2 portion, any number of analogs of this fusion partner can be made.

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Typically, the analogs of the nucleotide sequences encoding the HA2 peptides and/or the fusion partner may differ by only 1 to about 4 codon changes, in addition to the nucleotide mutations to the above-identified fragment. Other sequences of this invention include modified nucleotide sequences which encode polypeptides with minor amino acid variations from the natural amino acid sequence of HA2. For example, conservative amino acid replacements may be introduced by altering, deleting or replacing codons of the native sequence, in addition to altering those codons in Formula II according to one embodiment of this method.

Conservative replacements are those that take place within a family of amino acids that are related in their side chains and are well known in the art. For example, it is reasonable to expect that an isolated replacement of a selected amino acid with a conservative replacement of an amino acid with a structurally related amino acid will not have a significant effect on the activity of the protein, especially if the replacement does not involve an amino acid at an epitope of the HA2 polypeptide.

The construction of modified nucleotide sequences and proteins or fusion proteins, given the description herein and conventional methods of protein modification known to one of skill in the art, are believed to be encompassed by this invention.

The novel modified nucleotide sequences of this invention are further characterized by encoding an immunogenic determinant of a modified HA2 subunit of an HA protein, derived from an H3N2 subtype. The encoded protein may contain all or a portion of the H3N2 HA2 sequence, including the Formula I amino acid sequence. The currently preferred embodiment provides a novel DNA sequence encoding an H3HA2 protein or fragment thereof fused in frame to a DNA sequence encoding a portion of the nonstructural influenza protein 1 (NS1). One modified fusion protein-encoding nucleotide sequence is obtained by making mutations according to this invention in the nucleotide sequence encoding the fusion protein NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO:10]. Upon mutation, the nucleotide sequence [SEQ ID NO:58] for this fusion protein [SEQ ID NO:10] is referred to herein as pOTS208NS1H3mut5585.

The modified coding sequences for the HA2 proteins, as well as the coding sequences for NS1 and other viral proteins of influenza virus can be prepared synthetically or can be derived from viral RNA or from available cDNA-containing plasmids by known techniques. For example, see references known to the art which disclose the nucleotide coding sequences for HA from the A/Japan/305/57 strain [Gething et al., Nature, 287:301-306 (1980)]; strain A/NT/60/68 [Sleigh et al., and

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Both et al., in <u>Developments in Cell Biology</u>, Elsevier Science Publishing Co., pages 69-79 and 81-89, respectively, (1980)]; strain A/WSN/33 [Davis et al., <u>Gene</u>, 10:205-218 (1980); Hiti et al., <u>Virology</u>, 111:113-124 (1981)]; and fowl plague virus [Porter et al. and by Emtage et al., both in <u>Developments in Cell Biology</u>, cited above, at pages 39-49 and 157-168]. Also, influenza viruses, including other strains, subtypes and types, are available from clinical specimens and from public depositories, such as the American Type Culture Collection (ATCC), Rockville, Maryland, U.S.A.

Novel modified nucleotide sequences of this invention may also include allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) of DNA sequences encoding the H3HA2 protein sequences, and the Formula III fragment [SEQ ID NO:62]. Similarly, DNA sequences having the Formula III fragment, which sequences encode other H3N2 HA2 proteins of the invention include sequences which differ in codon sequence outside of Formula II due to degeneracies of the genetic code or variations in the DNA sequence encoding H3HA2 proteins. Such codon differences may be caused by point mutations or by induced modifications to enhance the activity, half-life or production of the peptide encoded thereby. Also covered by this invention are DNA sequences characterized by the above modification of Formula II into Formula III, which hybridize under stringent conditions with the DNA sequences encoding the HA2 subunit proteins, e.g., H3HA2 proteins, of this invention. DNA sequences which hybridize under non-stringent conditions with the disclosed sequences, but which encode proteins or fragments retaining the biological activities of the H3HA2 proteins, are also included in this invention. Typical conditions for stringent or non-stringent hybridization are known to those of skill in the art [See, e.g., Sambrook et al., cited above].

The actual techniques for producing the mutations described herein are now conventional to the art of genetic engineering, and are readily known and available to one of skill in the art. See, e.g., Sambrook et al, cited above. Such conventional techniques include, for example, site directed mutagenesis, which is available in commercial kits from, e.g. Clonetech and Promega Corporation. Other suitable techniques include, e.g., total gene synthesis and removing the fragment and replacing it with a synthetically generated, mutated fragment. It is anticipated that similar modifications to any H3HA2 sequence having an analogous codon pattern will result in the enhanced expression in *E. coli*, exemplified by the modified H3HA2 sequence.

The mutations described herein are preferentially developed for increased expression of the influenza protein or fusion protein in  $E.\ coli$ , which is the preferred host because it can be used to produce the desired proteins safely and cheaply. To circumvent the requirement of ampicillin for plasmid selection in production fermentations, a preferred method of production which uses the modified nucleotide sequences of this invention employs an alternative expression system in which the  $\beta$ -lactamase coding sequence is wholly or partially replaced by a coding sequence for an alternative selectable marker, such as, kanamycin or chloramphenicol.

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To aid in expression of the H3HA2 peptides or fusion proteins, these protein sequences or fragments thereof may also be fused to a polypeptide capable of further enhancing expression of these fragments in the selected host system. Ordinarily, such a peptide would contain a leader sequence fragment that provides for secretion of the H3HA2 subunit fragment, in the host cell. The leader sequence fragment typically encodes a signal peptide comprised of hydrophobic amino acids which direct the secretion of the protein from the cell. There may be processing sites encoded between the leader sequence and the H3HA2 fragment that can be cleaved either *in vivo* or *in vitro*. Alternatively, a promoter sequence may be linked directly with the DNA molecule encoding the H3HA2 fragment. Such polypeptides, promoter and leader sequences are known to those of skill in the art and may be readily selected for expression in the selected host.

Construction of bacterial expression systems, preferably *E. coli* expression systems, including expression vectors and transformed host cells are also within the skill of the art. See, generally, methods described in standard texts, such as Sambrook *et al.*, cited above. The present invention is therefore not limited to any particular vector, nor to any particular purification process from cell lysates or cell medium.

Influenza proteins encoded by the modified nucleotide sequence may be expressed in enhanced manner according to the first embodiment of the method of this invention, or the influenza proteins may be expressed in an enhanced manner by translation from their native sequences by the second embodiment of the method. Additionally, the methods of this invention may be used to enhance the expression of a fusion protein which comprises a pretein sequence encoded by the modified nucleotide sequence containing Formula III in place of Formula II in the native nucleotide sequence encoding an HA2 subunit of an HA protein from an H3N2 subtype virus, fused in frame to another protein or protein fragment (a "fusion partner") capable of enhancing expression of the fusion protein.

One of skill in the art may readily select a fusion partner protein or fragment taking into account the desired host cell, i.e., *E. coli*, and utilizing the teachings herein. For the purposes of this invention, the H3HA2 fragment or sequence encoded by a modified nucleotide sequence as described above or the native sequence used in the second embodiment of this method may be fused to any peptide capable of further enhancing its expression in the host cell selected or of increasing its immunogenicity. The method of the present invention does not limit the nature of the "partner" protein or fragment to which the H3HA2 fragment is fused to provide the enhanced expression of the resulting fusion protein.

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For example, the influenza protein or fragment bearing the amino acid sequence of Formula I may be fused to a number of conventionally known and used "partner" proteins [See, general texts on expression such as Current Protocols in Molecular Biology, Vol. 2, suppl. 10, publ. John Wiley and Sons, New York, NY, pp. 16.4.1-16.8.1 (1990); Smith et al, Gene, 67:31-40 (1988); U. S. Patent No. 4,801,536, among others]. However, it may be desirable that this fusion "partner" protein be an influenza protein sequence or fragment thereof derived from the same or another strain of influenza virus as the HA protein or protein fragment. Preferably, this fusion partner protein is all or a portion of the influenza virus NS1 gene or an HA2 subunit.

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In such a fusion protein, a linker sequence may be inserted optionally between the two sequences, i.e., between the sequence encoding the fusion partner and the HA2 protein encoded by the modified nucleotide sequence of this invention or the native sequence for expression according to the second embodiment of the method. This optional linker may provide space between the two protein sequences; and may encode a polypeptide or contain a cleavage site, which is selectively cleavable or digestible by conventional chemical or enzymatic methods. An example of a fusion protein whose expression can be enhanced by a method of this invention is NS1(1-81)H3HA2(1-221) illustrated in Fig. 2 [SEO ID NO: 10], which comprises the first 81 amino acids of NS1 (derived from an H1N1 subtype virus, A/PR/8/34) fused to the sequences spanning amino acid 1 to 221 of the H3HA2 subunit (amino acids 1-221) via an optional four amino acid linker sequence. Another exemplary fusion protein, NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> SEQ ID NO:72, comprises the first 81 amino acids of NS1 fused to the sequences spanning amino acid 77 to 221 of the truncated H3HA2 subunit. In other embodiments, the NS1 portion may comprise the sequence spanning amino acid residues 1 to amino acids 42 of H1N1. The HA2 fragment may alternatively be fused to a portion of the NS1 peptide derived from a selected Type A virus, e.g., an H3 subtype virus (H3N2).

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These proteins, their native nucleotide sequences, and their uses, are described in co-pending U.S. application 07/837,773, filed February 18, 1992, which is incorporated by reference.

As described below in the examples, the host cells used to express these fusion proteins may be modified by the second embodiment of the method of this invention to contain tRNA molecules capable of translating the rare arginine codons of Formula II. See, e.g., Example 25. Alternatively, the nucleic acid sequence encoding these and other suitable H3HA2 proteins or H3HA2-containing proteins, i.e. those comprising a native Formula II sequence [SEQ ID NO:9], may be modified by the first embodiment of the method of this invention to replace Formula II with the Formula III sequence to increase the expression of the encoded protein in *E. coli* according to the method of this invention.

The proteins and fusion proteins whose expression is enhanced by the methods of this invention may be employed in vaccine compositions. Several of the specific influenza proteins or fusion proteins described herein, which are produced according to the methods of this invention, have demonstrated the ability to stimulate or produce a protective immune response capable of recognizing an influenza virus or influenza virus-infected cells and protecting the vaccinated mammal against disease caused thereby. This protective response is desirably a T cell response, produced in the substantial absence of vaccine-induced neutralizing antibody. Such H3HA2 proteins and fusion proteins are capable of inducing T helper cells, particularly cytotoxic T lymphocytes, in the absence of neutralizing antibodies.

Pharmaceutical vaccine compositions can contain an effective immunogenic amount of a selected H3HA2 protein produced according to this invention or encoded by a modified nucleotide sequence of this invention in admixture with a suitable adjuvant in a nontoxic and sterile pharmaceutically acceptable carrier. Suitable carriers for vaccine use, as well as other vaccine formulation additives and adjuvants, are well known to those of skill in the art. See, e.g., European Patent Application No. 366, 238, published May 2, 1990; and European Patent Application No. 366,239, published May 2, 1990. Such compositions may be effectively administered to human and animal patients to induce the appropriate immune response. The details of dosage and treatment using such compositions are also described in the above-cited published patent applications.

The following examples illustrate methods for preparing H3HA2 and BHA2 fusion proteins of the invention and demonstrate the subtype specific

protection against heterologous virus induced upon vaccination with the H3HA2 proteins. The following examples also illustrate methods for preparing the modified DNA sequences of the invention. All of these examples are illustrative only and do not limit the scope of the invention.

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#### EXAMPLE 1 - PLASMID DMS3H3HA

Plasmid pFV88 contains the entire 221 amino acid length HA from A/Udorn, an H3 subtype virus [C. J. Lai et al, <u>Proc. Natl. Acad. Sci. USA</u>, <u>77</u>:210-214 (1980)], which HA nucleic acid sequence is illustrated in Fig. 1 [SEQ ID NO: 1]. This plasmid was cut with Pst I. The resulting 1900 bp fragment, which contains the entire HA (HA1 and HA2) fragment and some GC tailing, was then inserted into pUC18 [Bethesda Research Laboratories]. The resulting plasmid is termed pMS3 or pMS3H3HA.

#### 15 EXAMPLE 2 - pMG1

Plasmid pAPR801 is a pBR322-derived cloning vector which carries the NS1 coding region (A/PR/8/34). It is described by Young et al, in <u>The Origin of Pandemic Influenza Viruses</u>, ed. by W. G. Laver, Elsevier Science Publishing Co. (1983).

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Plasmid pAS1 is a pBR322-derived expression vector which contains the PL promoter, an N utilization site (to relieve transcriptional polarity effects in the presence of N protein), and the cII ribosome binding site including the cII translation initiation codon followed immediately by a BamHI site. It is described by Rosenberg et al, in Methods Enzymol., 101:123-138 (1983).

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Plasmid pAS1ΔEH was prepared by deleting a non-essential EcoRI-HindIII region of pBR322 origin from pAS1. A 1236 base pair BamHI fragment of pAPR801, containing the NS1 coding region in 861 base pairs of viral origin and 375 base pairs of pBR322 origin, was inserted into the BamHI site of pAS1ΔEH. The resulting plasmid, pAS1ΔEH/801, expresses authentic NS1 (230 amino acids). The plasmid has an NcoI site between the codons for amino acids 81 and 82 and an NruI site 3' to the NS sequences. The BamHI site between amino acids 1 and 2 is

retained.

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Plasmid pMG27N, a pAS1 derivative [Mol. Cell. Biol., 5:1015-1024 (1985)], was cut with BamHI and SacI and ligated to a BamHI/NcoI fragment encoding the first 81 amino acids of NS1 from pAS1ΔEH801 and a synthetic DNA NcoI/SacI fragment of the following sequence:

**SEQ ID NO: 35:** 

**SEQ ID NO: 36:** 

3'- CTAGTATACAATTGTCTATAGTTCCGGACTGACTGACTC -5'.

The resulting plasmid, pMG1, allows the insertion of DNA fragments after the first 81 amino acids of NS1 in any of the three reading frames within the synthetic linker fragment followed by termination codons in all three reading frames.

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#### EXAMPLE 3 - pMG1H3HA

Plasmid pMG1, described above in Example 2, was digested with NcoI and XbaI, releasing a 54 bp fragment, which was discarded. Plasmid pMS3H3HA, described in Example 1 above, was digested with HhaI and XbaI, and a 701 bp fragment containing the coding sequence for the HA2 subunit of influenza strain A/Udorn (H3N2) was isolated, as illustrated in Fig. 1 [SEO ID NO: 1].

Synthetic oligonucleotides were annealed to generate an NcoI 5' overhang sequence (at the 5' end) and a HhaI 3' overhang sequence (at the 3' end). The sequence of these oligonucleotides is as follows:

20 SEQ ID NO: 37: 5'-CATGGGCGCCCATATGGGCATATTCGGCG-3' SEQ ID NO: 38: 3'- CCGCGGGTATACCCGTATAAGCC-5'.

The annealing reaction was performed as follows. The annealing mixture was made up of  $2.5\mu L$  each of 5' oligo ( $1.3~\mu g/\mu L$ ), the 3' oligo ( $1.2~\mu g/\mu L$ ), and added water ( $15~\mu L$ ) to a final volume of  $20~\mu L$ . The reaction tubes were then placed in 4 mL culture tubes containing water which had been heated to  $65^{\circ}C$  for 10 minutes and allowed to cool down slowly. The tubes were then put on ice and used immediately for ligation.

This three part ligation generates pMG1H3HA2<sub>(1-221)</sub> [SEQ ID NO: 9] which codes for the first 81 amino acids of NS1 fused to four amino acids donated from the linker and amino acids 1-221 of the HA2 subunit. This sequence is illustrated in Fig. 2 [SEQ ID NO: 9 & 10]. This molecule is also designated NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 9 & 10] or H3C13.

#### EXAMPLE 4 - NS1(1-81)H3HA2(77-221) [SEO ID NO: 11 & 12]

pMS3H3HA, described in Example 1 above, was digested with EcoRI and end-filled (Klenow). Subsequently, the vector was digested with XbaI. A 487 bp fragment, which contains the coding sequence for amino acids 77-221 of

the HA2 subunit, was isolated and ligated to the HpaI and XbaI sites of pMG1. The resulting vector codes for a fusion polypeptide containing amino acids 1-81 of NS1 fused to amino acids 77-221 of the HA2 subunit. This molecule has been termed NS1(1-81)H3HA2(77-221) and is illustrated in Fig. 3 [SEQ ID NO: 11 & 12].

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## EXAMPLE 5 - pMG<sub>42</sub>BLHA2

To derive a vector similar to pMG1 (described in Example 2), which contains the coding region for the first 42 amino acids of NS1 rather than the first 81 amino acids of NS1, pMG1 was digested with BamHI and NcoI and ligated to the BamHI/NcoI fragment encoding amino acids 2 to 42 of NS1 from pNS142TGF α. pNS142TGFα is derived when pAS1ΔEH801 is cut with NcoI and SalI and ligated to a synthetic DNA encoding human TGFα as an NcoI/SalI fragment. pNS142TGFα encodes a protein comprised of the first 42 amino acids of NS1 and the mature TGFα sequence. The NS1 portion of pNS142TGFα contains an amino acid change from Cys to Ser at amino acid 13.

The resulting plasmid, termed pMG<sub>42</sub>A, was then modified to contain an alternative synthetic linker after the NS1<sub>42</sub> sequence with a different set of restriction enzyme sites within which to insert foreign DNA fragments into the three reading frames after the NS1<sub>42</sub>. This linker has the following sequence:

20 **SEQ ID NO: 39:** 

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CATGGATCATATGTTAACAAGTACTCGATATCAATGAGTGACTGAAGCT-3'

**SEQ ID NO: 40:** 

3'- CTAGTATACAATTGTTCATGAGCTATAGTTACTCACTGACT -5'.
The resulting plasmid is called pMG42B. This vector is needed to contain the neomycin phosphotransferase-1 (NPT-1) gene which confers kanamycin resistance.

As described in Shatzman and Rosenberg, Met. Enzymol., 152:661-673 (1987), pOTS207 is a pAS derived cloning vector which carries the kanamycin resistance gene from Tn903 [Berg et al, Microbiology, ed. D. Schlessinger, pp. 13-15, American Society for Microbiology (Washington, DC 1978); Nomura et al, The Single-Stranded DNA Phages, ed. D. Denhardt et al, pp.467-472, Cold Spring Harbor Laboratory (New York 1978); Castellazzi et al, Molecul. Gen. Genet., 117:211-218 (1982)]. It was constructed by digesting plasmid pUC8 [Yanisch-

Perron et al, <u>Gene</u>, <u>33</u>:103-119 (1985)], with BamHI and ligated to a BcII fragment containing the kanamycin gene from Tn903. The resulting plasmid, pUC8-Kan, was digested with EcoRI and PstI, and the fragment containing the kanamycin gene

was inserted between the EcoRI and PstI sites of pOTSV [Shatzman and Rosenberg, cited above]. The resulting plasmid is pOTS207.

The pOTS207 was digested with EcoRI and PstI, and the 1467 bp fragment containing the kanamycin resistance gene was isolated. Synthetic oligonucleotides:

SEQ ID NO: 41:

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5' AATTCGTACCTA 3'

**SEQ ID NO: 42:** 

3' GCATGGATCTAG 5'

were made to link the NPT-1 gene to pMG42B vector. pMG42B was digested with BgIII and PstI. The EcoRI/PstI NPT-1 gene fragment and the synthetic oligo linker were ligated to the digested pMG42B. The resulting plasmid, pMG42Kn allows fusions, in three different reading frames, to the NS1-42 gene, while allowing antibiotic selection with kanamycin.

Plasmid pBHA is a pBR322-derived vector, containing the complete nucleotide sequence of the HA gene of a Type B influenza virus (B/Lee/40). It is described by Krystal et al, Proc. Natl. Acad. Sci. USA, 79:4900-4804 (1982). pBHA was digested with RsaI and a 813 bp fragment containing the HA subunit was isolated. This fragment was ligated into plasmid pMG42Kn (described above) that had been digested with ScaI. During the cloning, a nucleotide base (T) was deleted from the ScaI recognition site shifting the gene out of the reading frame.

The vector was digested with NcoI, and filled-in using Klenow, putting the gene back into the reading frame.

The resulting construct, pMG<sub>42</sub>BLHA2 [SEQ ID NO: 14], expresses a fusion polypeptide containing amino acids 1-42 of NS1 and 41-233 of the HA2 subunit. This construct contains the Cys to Ser change at amino acid 13 of the NS1 portion of the fusion peptide.

In preliminary studies with this construct, vaccinated laboratory mice demonstrated protection from challenge with Type B influenza in the absence of neutralizing antibody for the virus.

# 30 EXAMPLE 6 - PREPARING SEED VIRUS AND RAISING ANTISERA

The seed virus, A/Udorn, was prepared according to the procedures described in P. Palese and J. Schulman, <u>Virol.</u>, <u>57</u>:227-237 (1974). Briefly, this technique is as follows.

Influenza virus strain A/Udorn was inoculated in 10-day old embryonated hen's eggs into the allantoic cavity. The eggs were incubated for 24-48 hours at 35°C then chilled at 4°C overnight. A portion of the eggshell over the airsac was removed and the allantoic fluid was aseptically removed using a

10-ml syringe. The fluid was centrifuged at low speed (3,000 x g) to remove particulates. This clarified supernatant was centrifuged at high speed using an SW28 Beckman rotor at 27,000 rpm (4°C for 90 minutes), resulting in the virus pellet. The virus was resuspended in 10 mM Tris (pH 7.5) containing 100 mM NaCl, 1 mM EDTA and repelleted as before. The virus was layered on 30-60% sucrose gradient in 1 mM EDTA (NTE) and spun for 3-5 hours at 25,000 rpm. The band in the middle of the tube was withdrawn, diluted in NTE and centrifuged at 27,000 rpm for 90 minutes. The pellet was suspended in phosphate-buffered saline (PBS). These viral particles were used as immunogens for preparation of antisera.

Antisera was prepared as follows. 100-200 micrograms of purified virus in complete Freund's adjuvant was injected into the subscapula of a New Zealand White rabbit. A second injection in incomplete Freund's adjuvant was done 4 weeks later, and the animals were bled and antisera collected 7-10 days later.

#### 15 EXAMPLE 7 - EXPRESSION OF H3HA2 FUSION PROTEINS

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### A. <u>NS1(1-81)H3HA2(1-221) [SEO ID NO: 9 & 10]</u>

The plasmid pMG1H3HA2<sub>(1-221)</sub> [SEQ ID NO: 9] was transfected into E. coli strain AR58 [SmithKline Beecham Pharmaceuticals]. Cultures were grown at 32°C to mid-log phase at which time cultures were shifted to 39.5°C for 2 hours. The E. coli cell pellets containing the recombinant polypeptide were then stored at -70°C until used.

Production of the NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> protein [SEQ ID NO: 10] was confirmed by Western blot analysis [Towbin et al, <u>Proc. Natl. Acad. Sci. U.S.A.</u>, <u>76</u>:4350 (1979)] using antisera prepared against A/Udorn virus, as described in Example 5. A major immunoreactive species was found at a molecular weight of 35,050 daltons.

# B. NS1(1-81)H3HA2(77-221) ISEO ID NO: 11 & 121 The plasmid encoding the NS1(1-81)H3HA2(77-221) peptide [SEQ

ID NO: 12] was expressed as described in part A above. Production of this peptide was confirmed by Western blot analysis, as described above. A major immunoreactive species was found at a molecular weight of 26,697 daltons.

#### EXAMPLE 8 - PARTIAL PURIFICATION OF H3HA2 FUSION PROTEINS

<u>E. coli</u> cell pellets containing the recombinant polypeptides, prepared as described in Example 6, were stored at -70°C until used. <u>E. coli</u> cells were thawed and resuspended in lysis buffer A (50 mM Tris-HCl, 5% glycerol, 2 mM EDTA and 0.1 mM DTT, pH 8.0) at 10 mL/gram. The stirred suspension was then

treated with lysozyme (0.2 mg/mL) for 45 minutes at room temperature and sonicated 2x for 2-3 minutes each time by a Sonicator. The resultant suspension was treated with 0.1% DOC for 60 minutes at 4°C, then centrifuged at 25,000 x g. The pellet was resuspended by sonication in 50 mM glycine pH 10.0, 5% glycerol, 2 mM EDTA and then the suspension was treated with 1% Triton X-100 [J.T. Baker Chemicals Co.] at 4°C for 60 minutes and centrifuged as above.

The resulting pellet was solubilized in 50 mM Tris, 8 M urea, pH 8.0 and centrifuged to remove any insoluble material. This solubilized material is dialyzed against 10 mM Tris, 1 mM EDTA, pH 8.0 followed,

again, by centrifugation of insoluble material. The solubilized material is designated as "crude" material and is used in in vitro and in vivo mouse assays. At this point, the material is approximately 40 - 50% pure.

The "crude" material was electrophoresed through an SDS-PAGE and the appropriate H3HA2 protein bands were visualized by KCl staining according to D. Hager et al, <u>Anal. Biochem</u>, <u>109</u>:76-86 (1980). The band was cut-out and eluted electrophoretically by the "S&S Elutrap Electro-Separation System" [Schleicher & Schuell]. The electro-eluting buffer was the Tris-glycine. A concentrated and eluted sample was obtained and exhaustively dialyzed against 0.01 M NH4HCO<sub>3</sub> and 0.02% SDS [M. Hunkapiller et al, <u>Method. Enzymol.</u>, <u>91</u>:227-236 (1983)]. This sample was frozen quickly by dry ice and lyophilized to complete dryness.

This sample was frozen quickly by dry ice and lyophilized to complete dryness. The lyophilized material was brought back into solution using 50 mM Tris pH 8.0 and used for in vitro and in vivo mouse assays.

Following this gel elution step, the protein is usually greater than 75% pure.

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#### EXAMPLE 9 - CONSTRUCTION OF POTS208 VECTORS

pOTSV is described in Devara et al, Cell, 36: 43-49 (1984). Briefly, this vector is a pAS1 derivative with t-oop inserted at the NruI site and a synthetic oligonucleotide encoding SacI, XhoI and XbaI restriction sites inserted at the SaII site (which is destroyed).

#### A. pOTS208

pOTS208 was prepared by digesting pOTSV with EcoRI and ScaI, followed by fill in reaction using Klenow. Tn5 Plasmid DNA [described in R. Jorgensen et al., Mol. Gen. Genet., 177:65-72 (1979)] was digested with HindIII and SmaI, followed by a fill in reaction using Klenow yielding a 1323 bp fragment encoding for neomycin phosphotransferase-2 gene (NPT-2). This fragment is described in detail in Rothstein et al., Cell, 19:795-805 (1980) and Jorgensen, cited

above. This fragment and the above digested vector were ligated together to create pOTS208, which is kanamycin resistant.

### B. <u>pOTS208H3C13</u>

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pMG1H3HA2(1-221) (Example 3) was digested with BamHI and XbaI, releasing two fragments: an 806 bp BamHI fragment and a 160 bp BamHI/XbaI fragment. These fragments together code for NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub>. A three part ligation between the two fragments and BamHI/XbaI digested pOTS208 (part A) yield pOTS208H3C13 which utilizes NPT-2 for kanamycin resistance.

# C. pOTS208NS181Nco

pOTS208H3C13 (part B) was digested with BgIII. pSelect [Promega] was cut with BamHI and ligated with the BgIII fragment, resulting in pSelectNPTII102. Transformation into E. coli JM101 [ATCC E. coli 33876] was followed by selection on kanamycin and tetracycline plates. KanR was conferred by the NPT2 region from pOTS208H3C13. Some lambda sequence was also on the BgIII fragment. Oligo 4852, SEQ ID NO: 49 GCATCGCCATGAGTCACGACG, was used to mutate the NcoI site to CCATGA in pSelectNPTII102, resulting in pSelectNPTII102-8. This vector was cut with BstEII and BssHII. pOTS208H3C13 was cut with BstEII, BssHII and SphI, and fragment exchange generated pOTS208NS181H3HA2-26. This clone has the NcoI site of NPT2 mutated. pOTS208NS181NS181H3HA2-26 was cut with NcoI and SalI, filled in and ligated with Linker 1041 [New England Biolabs] to insert a KpnI site and regenerate the NcoI site. This step also deletes the H3C13 region. The unique XbaI site of the parent pOTS208 vector is downstream of the deletion. The resulting vector is pOTS208NS181Nco.

# EXAMPLE 10 - MODIFICATION OF GENE ENCODING H3HA2 FUSION PROTEIN

In order to increase yield of the H3HA2 protein, silent mutations to certain rare arginine codons were made to the coding sequence of the H3HA2 protein. These nucleotide changes resulted in no change in the protein sequence.

A mutant H3C13 protein was prepared by mutating the nucleotide sequences of the fusion protein prepared according to Example 3 above. Site directed mutagenesis using the Altered Sites System [Promega Corporation] according to the manufacturer's directions was used to change nucleotide numbers, 622, 625, and 634 (A to C) and 624, 627, and 636 (G to T) of nucleotide sequences [SEQ ID NO:9] encoding the NS1(1-81)H3HA2(1-221) fusion protein of Fig. 2 [SEQ ID NO:10], thereby changing the codons at these regions from AGG to CGT,

both encoding Arg. These changes correspond to nucleotide numbers 367, 370, and 379 (A to C) and 369, 372, and 381 (G to T) of the HA2 fragment of Fig. 2 [SEQ ID NO: 58].

Fig. 2 illustrates the modified nucleotide sequences of the fusion protein [SEQ ID NO: 10] by contrast with the nucleotide sequence [SEQ ID NO: 9] of the "unmodified" fusion gene (nucleotide changes above sequences of unmodified gene). Mutagenesis on this sequence was carried out according to the method provided with the pSelect kit from Promega.

### A. <u>NS1(1-81)H3HA2(1-221) [SEO ID NO: 10]</u>

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Briefly, cloning for the mutagenesis was performed as follows. The pSelect plasmid [Promega] and pMG1H3HA2 (Example 3) were each digested with HindIII. These two plasmids were ligated together and selected on tetracycline plates. The resulting vector is pSelH3HA2. Mutagenesis was performed according to Promega's kit. The following oligonucleotide was used: SEQ ID NO: 43: 5'-AAACTGTTTG AAAAAACACG TCGTCAACTG CGTGAAAATG

5'-AAACTGTTTG AAAAAACACG TCGTCAACTG CGTGAAAATG CTGACGACAT GGGC -3'.

Clones were verified by restriction endonuclease <u>Hinc</u>II. The resulting plasmid, pSelH3HA2mut5585 was digested with <u>Nco</u>I and <u>Xba</u>I, and a 748 bp fragment coding for the H3HA2mut5585 polypeptide was isolated.

pOTS208NS181Nco (Example 9C) was digested with NcoI and XbaI. The ligation of linear pOTS208NS181Nco and the 748 bp fragment resulted in pOTS208NS1H3mut5585 [SEQ ID NO:7]. This vector codes for the polypeptide, NS1(1-81)H3HA2(1-221) [SEQ ID NO:10].

B. Expression of mutated gene encoding H3C13 protein

The plasmid of A was transfected into *E. coli* strain AR58 [SmithKline Beecham]. Cultures are grown at 32°C to mid-log phase at which time cultures are shifted to 39.5°C for two hours. The *E. coli* cell pellets containing the recombinant polypeptide are then stored at -70°C until used. Production of the NS1(1-81)H3HA2(1-221) protein [SEQ ID NO:10] is confirmed by Western blot analysis [Towbin *et al.*, Proc. Natl. Acad. Sci. U.S.A., 76:4350 (1979)] using antisera prepared against A/Udorn virus, as described in Example 4. A major immunoreactive species is expected at a molecular weight of approximately 35,00 daltons.

The expression levels obtained are about 50-100% higher than those obtained by the expression of the unmodified coding sequences in the same expression system.

#### C. Construction of Alternative H3 Mutant

pSelH3HA2mut5585 (part A) was subjected to site-directed mutagenesis, as described above. Oligo SEQ ID NO: 44
TGTGACAATGCTTGCATCGGTTCAATCCGTAATGGTACTTATGACCA

TGATG, was used and clones were verified by restriction endonuclease Rsa1. The resulting plasmid, pSelH3HA2mut2 was digested with Nco1 and Xba1, and an approximately 748 bp fragment encoding for the H3HA2mut2 polypeptide was isolated. pOTS208NS181Nco was digested with Nco1 and Xba1. The ligation of linear pOTS208NS181Nco (Example 9C) and the 748bp fragment resulted in pOTS208NS1H3mut2. This vector codes for the NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> polypeptide [SEQ ID NO: 10].

#### Example 11 - PLASMID pD

Plasmid pAS1\_EH/801 (described above in Example 2) was cut with BglII, end-filled with DNA polymerase I (DNApolI; Klenow), and ligated closed, thus eliminating the BglII site. The resulting plasmid pBgl was digested with Ncol, end-filled with DNApolI (Klenow), and ligated to a BglII linker. The resulting plasmid, pB4, contains a BglII site within the NS1 coding region. Plasmid pB4 was digested with BglII and ligated to a synthetic DNA linker of the sequence:

20 **SEQ ID NO: 45:** 

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5'-GATCCCGGGTGACTGA -3'

**SEQ ID NO: 46:** 

3'- GGCCCACTGACTGACTCTAG-5'.

The resulting plasmid, pB4+, permits insertion of DNA fragments within the linker following the coding region for first 81 amino acids of NS1 followed by termination codons in all three reading frames. Plasmid pB4+ was digested with XmaI (cuts within linker), end-filled (Klenow), and ligated to a 520 base pair PvuII/HindIII, end-filled fragment derived from the HA2 coding region. The resulting plasmid, pD, codes for a protein [SEQ ID NO: 18] comprised of the first 81 amino acids of NS1, three amino acids derived from the synthetic DNA linker (Gln-Ile-Pro), followed by amino acids 65-222 of the HA2.

Expression is obtained by transfecting pD into a desired E. coli strain, preferably LW14, using standard techniques. Purification may be by standard techniques or, preferably, as described in Example 18 below.

# EXAMPLE 12 - H3 SUBTYPE HETEROLOGOUS PROTECTION ELICITED BY VACCINATION WITH NS1(1-81)H3HA2(1-221) [SEO ID NO: 10]

Mice (NIH/Swiss; 15 per group) were vaccinated subcutaneously with 50 or 10 µg NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 9 & 10] in aluminum

hydroxide on days 0 and 21. The mice were boosted intraperitoneally on day 42 with the protein without adjuvant. On day 47, mice were challenged intranasally with 2 - 3 LD<sub>50</sub> doses of either A/PR/8/34 (H1N1) or A/HK/68 (H3N2) virus, and survival was monitored through day 21. This represents a heterologous challenge (A/PR/8/34) and an H3 heterosubtypic challenge, since the NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> construct [SEQ ID NO: 9 & 10] was derived from A/Udorn/72 cDNA. The control group received adjuvant (CFA) only.

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The results in Table 1 below show that survival in mice vaccinated with NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 10] and challenged with A/HK/68 (80-93%) was significantly higher than in control mice which were injected with adjuvant only (26% survival). In contrast, vaccination with NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 10] did not confer protection against challenge with A/PR/8/34, an H1N1 strain (0-26% survival). Thus protection elicited by NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 10] is selective for antigenically diverse virus strains within the H3 subtype.

Likewise, vaccination with the D protein (NS1<sub>(1-81)</sub>HA2<sub>(65-222)</sub> [SEQ ID NO: 18], derived from the H1N1 subtype) elicits protection from heterosubtypic challenge with H1N1, but not the H3N2 subtype [S. Dillon et al, Nature, (1992); Mbawuike et al, Faseb. J., 5:A1362 (abs. 5749 and Table 1)]. These results in outbred mice also suggest that the response to the H1 and H3 proteins will not be restricted to a limited number of individuals with certain major histocompatibility alleles, and therefore the vaccine will be effective in a majority of individuals.

Table 1
Percent Survival After Challenge:

					J
	Immunization	НА	A/PR/8,	 /34 A/HK/	<b>'68</b>
		Si	ubtype	(H1N1)	(H3N2)
30					
	50 μg NS1 <sub>(1-81)</sub> H3F	IA2 <sub>(1-221</sub>	) Н3	26	80 <sup>*</sup>
	10 μg NSΊ <sub>(1-81)</sub> H3F	IA2 <sub>(1-221</sub>	) H3	0	· 93*
35	10 μg NS1 <sub>(1-81)</sub> HA2	<sup>2</sup> (66-222)	H1	67 <sup>*</sup>	13

A/HK/68 virus H3  $60^*$   $100^*$ Control (Al<sup>+3</sup>) - 0 26

p  $\leq$  0.05 vs. control in Fishers exact probability test

Vaccination of mice with live homologous (A/HK/68) virus provided complete or partial protection, reflecting protection mediated by neutralizing antibody (homologous H3N2 challenge) and/or CTL (heterologous H1N1 challenge), respectively.

Duration of protective immunity was tested by immunizing mice subcutaneously with the recombinant influenza protein plus adjuvant on days 0 and 21. Some mice were also given an ip injection of the protein (without adjuvant) on day 42. Mice were challenged with A/HK/68 (H3N2) on day 47, four weeks after the second injection. Control mice were immunized as described above for Table 1, where an ip injection was given at week 6 (5 days prior to challenge). The results in Table 2 show that CB6F<sub>1</sub> mice (15 per group) were significantly protected when challenged with the A/HK/68 heterologous H3 virus strain 5-28 days after the last injection.

			Table 2	
15				
	Dose (µg per injection)	Injection	Percent	
	of NS1 <sub>(1-81)</sub> H3HA2 <sub>(1-2)</sub>	<u> 21) Adjuva</u>	ınt Schedu	le Survival
	50 μg	CFA	0,21	86 <sup>*</sup>
20	50 μg	CFA	0,21,42	100*
	0 μg	CFA-	0,21	6 .
	50 μg	A1 <sup>+3</sup>	0,21	93*
	50 μg	$Al^{+3}$	0,21,42	93*
25	0 μg	A1 <sup>+3</sup>	0,21	0

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## EXAMPLE 13 - TYPE A CROSS-PROTECTION WITH D AND H3C13 30 PROTEIN

Mice (CB6F<sub>1</sub>) were divided randomly into six groups, with fifteen in each group. The mice were injected subcutaneously with proteins in  $Al^{+3}$  (100 µg) on days 0 and 21, and then were challenged with 2-3 LD<sub>50</sub> doses of virus on day 49. Survival was monitored through day 21. The results of this study are illustrated in Table 3 below. For convenience, NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 10] is referred to as H3C13 in the table below.

<sup>\*</sup> $p \le 0.05$  v. control in Fisher's exact probability test

PCT/US94/01149

Table 3

Percent Survival After Challenge with:

		HA	A/PR/8/34	A/HK/68
	Immunization Subtyr		(H1N1)	(H3N2)
1.	50 μg H3C13 H3		73*	73,*
	50 μg D	H1		
2.	10 μg H3C13 H3		67 <sup>*</sup>	100*
	10 μg D	<b>H</b> 1		
3.	1 μg H3C13 H3		86 <sup>*</sup>	73 <sup>*</sup>
	1 μg D H1			
4.	50 μg H3C13 H3		7	73 <sup>*</sup>
5.	50 μg D H1		47* *	7
	.2			
6.	Al <sup>+3</sup> control -		7	0

vs. control group

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This data demonstrates that mice immunized with a mixture of the D protein and H3C13 protein in aluminum adjuvant were protected against challenge with either A/PR/8/34 (H1) or A/HK/68 (H3) virus. In contrast, mice immunized with the D protein were protected against H1 but not H3 challenge. Likewise, mice immunized with the H3C13 protein were protected against the H3 but not the H1 challenge. Therefore, the combination of the D protein and the H3C13 proteins elicited protection against the currently circulating subtypes of influenza A virus. Thus, this combination represents a subtype cross-protective vaccine.

#### EXAMPLE 14 - CREATION OF DEA181KNRBS3 VECTOR

pMG1 (Example 2) and pMG42Kn (Example 5) were both digested with BamHI and NcoI. A 236 BamHI/NcoI fragment containing the coding sequence for amino acid sequence spanning residues 1 to 81 of the NS1 gene was isolated from pMG1. The digested pMG42Kn and the 236 bp fragment were ligated

<sup>\*\*</sup>  $p \le 0.03$  vs. control group

together and transformants were selected on LB and kanamycin agar plates. The resulting vector pMG181Kn(cII) maintains all regulatory elements of pMG42Kn except the NS1 (aa1-42) sequence is replaced with the NS1 (aa1-81) sequence.

pMG181Kn(cII) described above was digested with BstXI and BamHI. The following linker encoding ribosome binding site (RBS3) is cloned in the digested vector, replacing the cII RBS. The linker sequence is:

5' TAAGGAGGATATAACATATG [SEQ ID NO: 47]

3' TGGAATTCCTCCTATATTGTATACCTAG 5' [SEQ ID NO: 48].

BstXI BamHI

10 The resulting vector is pMG181KnRBS3.

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To generate pEA181KnRBS3, a 1.2 kb EcoRI/BgIII fragment from similarly digested pOTSV containing the lambda rexArexB region was cloned into mp18 [Gibco/Bethesda Research Labs] and mutagenized to create silent mutations in the two NdeI sites in this region. The mutations were CATATG to CATGTG in both sites. One site is in the rexA and the other in the rexB. The mutagenized fragment was inserted into both EcoRI/BgIII digested pMG181Kn(cII) and similarly digested pMG181KnRBS3, resulting in the plasmids pEA181Kn(cII) and pEA181KnRBS3, respectively. pEA181KnRBS3 has the useful properties of the pMG vectors, plus the additional attribute of nalidixic acid induction.

An EcoRI/PstI fragment containing the ampR gene of pBR322 was then inserted into EcoRI/PstI digested pEA181Kn(cII) and pEA181KnRBS3 to create pEA181CIIamp and pEA181RBS3amp, respectively. These plasmids are rexB+ and should be nalidixic acid inducible, in contrast to pMG1 and its descendants, which are rexB- and cannot be induced with nalidixic acid. The mutant EcoRI/BglII region was functionally examined by cloning it into a pMG1 vector carrying galK and demonstrating induction of galK with nalidixic acid.

# EXAMPLE 15 - CREATION OF VECTOR FOR PRODUCTION OF NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub>

Plasmid pOTS208BLeeHA2 was created as follows. An EcoRI fragment encoding the B/Lee HA region from plasmid pBHA (Example 5) was cloned into pSelect to generate pSelectPBHAS2. Site-directed mutagenesis inserted an NcoI site at the start of HA2, resulting in an N-terminus: MET GLY PHE PHE, and a C terminus of SER ILE CYS LEU. The resulting construct is called pSelectPBHAS2-B1. This plasmid was cut with NcoI and XbaI (a site in the polylinker of pSelect downstream of the HA gene), and ligated into NcoI/XbaI digested pEA181KnRBS3, described above, to generate pEA181BLeeB1-1. A

BamHI/EcoRI, filled in, fragment was cut out of pEA181BLeeB1-1 and ligated into pOTS208 (Example 9A), that had been digested with XbaI, filled in, and BamHI. The EcoRI and XbaI sites were regenerated by the ligation. This extra cloning step was necessary because there was no convenient cloning site to fuse the gene to NS(1-81) in pOTS208. pEA181KnRBS3 (described above) and pOTS208 have unique BamHI and XbaI sites to facilitate fragment exchanges.

A BamHI/XbaI fragment of about 1011 bp encoding the NS<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub> sequence from plasmid pOTS208BLeeHA2 was isolated and ligated into vector pSelect-1 [Promega], which was also digested with BamHI and XbaI. The resulting construct is called pSelBC13. This vector contains the coding sequence for NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub>, also termed BC13 [SEQ ID NO: 57].

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### **EXAMPLE 16 - CREATION OF VECTOR FOR PRODUCTION OF BC13mut2**

Mutagenesis was carried out on the pSelBC13 using Promega's

protocol and oligonucleotide 5492, SEQ ID NO: 50

GGAGGATGGGAAGGACTCATTGCAGGTTGG. This mutagenesis changed the ATG codon within the HA2 portion of the molecule to CTC (MET to LEU). The resulting plasmid is called pSelBC13mut5492. This plasmid was then digested with NcoI and XbaI, releasing a digestion fragment encoding for HA2, and ligated into pOTS208NS181Nco (Example 9C) that had been digested with NcoI and XbaI. The resulting construct, pOTS208NS1BLHA2mut5492 codes for the same polypeptide of pOTS208BLeeHA2, (i.e. BC13), except the internal translation start is eliminated at amino acid position 98 of the fusion protein. This protein is NS1(1-81)BHA2(1-223)(met-leu) [SEQ ID NO: 55].

A HindIII fragment of approximately 1 kb encoding NS1 (amino acid residues 7-81) and BLee HA2 (amino acid residues 1-223) and which contained the MET to LEU changes from plasmid pOTS208NS1BLHA2mut5492 was isolated. This fragment was ligated into the HindIII site of vector pSelect-1, resulting in pSelBC13mut5492. Mutagenesis was carried out using Promega's protocol and the following oligos 5920, 5921 and 5939, respectively:

SEQ ID NO: 51 CTCTGCTGTAGAAATCGGTAACGGTTGCTTTGAAACCAAAC

SEQ ID NO: 52

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### GGTTTCTTGGAAGGTGGTTGGGAAGGTCTCATTGCAGGTTGGCACGG SEQ ID NO: 53

GCTTTCCAACGAAGGTATCAATCAACAGTGAAGACGAGCATCTCTTGG.

This mutagenesis created the following silent codon changes in the HA2 region:

The codons for GLY at positions 93, 94, 97, 187, 215, and 217 were each mutated from GGG to GGT; the codons for ILE at positions 188, 189, and 214 were each changed from ATA to ATC; the codon for ASP at position 193 was changed from GAT to GAC; and the codon for ASN at position 216 was changed from AAT to AAC.

The resulting plasmid was called pSelBC13mut2. This plasmid was then digested with NcoI and XbaI, releasing a fragment of about 775 bp encoding for HA2. This fragment was ligated into pOTS208NS181Nco (described above), that had been digested with NcoI and XbaI. The resulting construct, pOTS208NS1BLmut2 (see Fig. 5 [SEQ ID NO: 54]), codes for the same polypeptide [SEQ ID NO: 55] as pOTS208NS1BLHA2mut5492, except for the silent codon changes.

#### 20 **EXAMPLE 17 - EXPRESSION OF FUSION PROTEIN**

pOTS208NS1BLmut2 [SEQ ID NO: 54] is transfected into a suitable host cell, preferably an E. coli strain and expressed essentially as described for the , H3 proteins described above. Strain LW14 is a derivative of E. coli K-12 strain W3110 [ATCC E. coli 27325]. The transducing phage P1 [E. coli ATCC 25404-B1] was grown on E. coli K-12 strain AR58, described above, the genotype of which is thr-galE::Tn10 λCI857 bio-uvrB- rpsL. Phenotypically, strain AR58 requires threonine, biotin for growth, is sensitive to UV light and DNA damaging agents, cannot use galactose as a carbon source, and is resistant to streptomycin. Strain W3110, a prototroph, is incubated with the phage and plated onto a medium 30 containing tetracycline to select for the transduction of the Tn10 element. The P1 phage picks up the segment of DNA containing the Tn10 and brings with it the  $\lambda$ CI857 bio- uvrB-. Thus the strain LW14 has the following genotype: galE::Tn10\(\lambda\) CI857 bio- uvrB-. Phenotypically, strain LW14 requires biotin for growth, is sensitive to UV light and DNA damaging agents, and cannot use galactose as a 35 carbon source.

#### **EXAMPLE 18 - PURIFICATION OF BC13mut2**

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E. coli whole cells transformed with the pOTS208NS1BLmut2 plasmid [SEQ ID NO: 54] as described in Example 16 above were recovered after fermentation by centrifugation or tangential flow filtration, washed to remove media, and stored at -70°C until use.

#### A. Step 1: Lysis and centrifugation (Isolation)

E. coli cells, 500 gm wet cell weight (WCW), were thawed and suspended in 4-7 volumes (2L) of buffer containing 0.025 M Tris-HCl, 0.005 M EDTA, pH 8.0. Chicken egg lysozyme (Calbiochem; suspension at 100 mg/mL) was added to a final concentration of 1 g/L and the preparation stirred with a Tekmar mixer at room temperature for 1 hour.

The lysate was centrifuged at 15,000 x g for 1 hour at 4°C and the supernatant discarded. The pellet (P1) was resuspended in 5 mL per gram of original wet cell weight of buffer consisting of 0.025 M Tris-HCl, 0.002 M MgCl<sub>2</sub>, pH 8.0 (about 2.5L).

The yield of this step was 90-100% by SDS-PAGE analysis, and 65-100% by RP-HPLC for product.

#### B. Step 2: Nuclease digestion and extraction

The preparation was treated with benzonase to digest nucleic acids,
then extracted with nonionic detergents to reduce the levels of *E. coli* contaminants in the pellet. Benzon nuclease, 0.2 mL per L of suspension, was added to the suspension, which was then stirred at room temperature for 1 hr. The sample was diluted with one volume of cold water containing 2% w/v Triton X-100 and 0.2% deoxycholate and stirred for 30 min at or below 15°C. Centrifugation was repeated as in step 1 and the supernatant discarded.

#### C. Step 3: Urea extinction

The pellet (P2) was extracted with 5 mL/gm WCW of cold 0.025 M NaH<sub>2</sub>PO<sub>4</sub>, 0.025 M Tris-HCl, pH 6.0, containing 4 M urea and 10 mM dithiothreitol (DTT). The Tekmar was used at a very low speed to mix, and temperature held below 15°C. The sample was stirred at 4°C for 1 hr. then centrifuged as in step 1. The supernatant (S3) was discarded. The pellet (P3) must be stored in the freezer until use.

# D. <u>Step 4: Solubilization, reduction, and DEAE</u> chromatography

The P3 pellet was solubilized and applied to anion exchange chromatography. This step removes remaining nucleic acid and major host cell proteins. P3 was suspended to 5 mL per gm WCW in .01 M Tris base, 8M urea (pH

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not adjusted). DTT was added to 25 mM. The pH was then adjusted to 12.5 using 6N NaOH, stirring for 15 min at room temperature, immediately followed by a 5-fold dilution of the same with 10 mM boric acid containing 25 mM DTT. If needed, the sample may be diluted to keep conductivity below 2mS/cm. The pH was adjusted to 9.0 and the sample stirred for up to 2 hour at room temperature.

The pH 12.5 treatment was necessary to complete solubilization of the B/Lee protein. However since carbamylation may occur under these conditions, the time was controlled very carefully. In addition, the pH 9 adjusted sample was unstable and cannot be held.

The sample (no more than 12 mg total protein per mL of resin) was then loaded onto a 14 x 250 cm (0.75L) DEAE Toyopearl 650M column equilibrated with buffer A. All steps were performed at room temperature at a linear velocity of 100 cm/hr. The column was washed sequentially with 2-3 column volumes of buffers B, C, and D, then eluted with buffer E. When protein began to elute from the column, flow was stopped for 15-20 minutes to improve the efficiency of elution of the B/Lee product; then the peak of product protein was collected. The column was cleaned with buffer D followed by 0.5 N NaOH.

The yield of this step was 85-90% by SDS-PAGE or Western blot analysis, and was estimated at 65-70% by RP-HPLC assay for product.

E. Step 5: Pretreatment and reverse phase chromatography

The buffer E eluate from step 4 was adjusted to no more than 1 g/L protein concentration and made 2% in SDS, 30 mM DTT, 0.1% M Tris, 5 mM EDTA, pH 9, then heated at either: 90°C for 60, 95°C for 30 min, or 100°C for 25 minutes, using a heat exchanger or water bath. This treatment was necessary to break up aggregates and prepare the sample for RP chromatography. The sample was cooled to room temperature and 2-propanol was added to 10% v/v.

The sample was injected on an Amberchrome reverse phase column equilibrated in 10% 2-propanol/0.2% trifluoroacetic acid (TFA)/water. The gradient shown in Table 1 was used to elute the column. Fractions containing product were analyzed by analytical RP-HPLC, pooled, and held at 4°C. The column was 25cm in height and was run at a linear velocity of 75-80 cm/hr at ambient temperature. An Amicon Vantage column, 9 cm in diameter, was used. The loading capacity of the column was 2 g/L.

The reverse phase column step has a yield of 30-80% (60-80% is typical).

#### F. Step 6: Precipitation

The pH of the RP eluate was adjusted to 6.0 +/- 0.5 using 1 N NaOH. After 10-15 min of stirring at room temperature, the precipitate was collected by centrifugation at 16,000 x g for 30 min at 4°C. The precipitate was resuspended to approximately 6-8 mg/mL protein concentration in 25 mM Tris, 8 M urea. DTT was added to 25 mM, and the sample stirred for 30 min at room temperature. The pH was adjusted to 12.5 and stirring repeated for 15 min, immediately followed by pH adjustment to 9.0 using HCl.

Alternately, the precipitate was suspended in buffer containing 0.1 M Tris-HCl, 2% SDS, 0.01 M EDTA, pH 8.0-9.0. DTT was added to 25 mM, and stirred 15-30 min until the solution was clear and all of the precipitate had dissolved. The sample was immediately taken to the next step.

### G. Step 7: Desalting and preparation of final product.

A 7 x 10 cm column was packed with Sephadex G25M (Pharmacia)
at room temperature. It was equilibrated with 3-7 column volumes of 25 mM TrisHCl, pH 9.,0, containing 5% w/v mannitol. Sample, at 6-10 mg/mL protein
concentration, is injected on the column (20-25% of total column volume, i.e. 80100 mL per injection). The column was developed at 150 cm/hr linear velocity and
the product desalted into the column buffer. The final product can be stored at 4°C.

The yield of steps 6 and 7 together was no less than 90%.

The product of the purification process was recovered at an overall yield of about 20-40%, and was over 95% pure by SDS-PAGE and RP-HPLC analysis. The final yield is about 3 g/500 g well cell weight.

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Table 4
Gradient for RP-LC of B/Lee

	Time	Flow	<u></u>	%A_	<u>%B</u>	
	0		80	90		0
	5		80	90	1	0
30	20		80	55	4.	5
•	120		80	35	6:	5
	145	•	80	10	9(	0
	180		80	10	90	0
	181	0	10	90		

35 A: 0.2% TFA in water

B: 99.8% 2-propanol/0.2% TFA

### H. Purification of FluD, NS1<sub>(1-81)</sub>HA2<sub>(65-222)</sub>,

FluD (Example 10) may be purified in much the same manner as the B/Lee with the following parameter alterations. For DEAE chromatography, the FluD column was equilibrated in 8M urea, 50 mM Tris, 25 mM borate at pH 9.0.

5 After the sample is loaded, sequential washes are performed with the following buffers: 4M urea in Tris-borate pH 9.0, 4 M urea and 0.4 M NaCl in Tris-borate pH 9.0, and Tris-borate pH 9.0. The product is eluted with a step elution of 2% SDS, 0.1 to 0.25 M NaCl, in Tris-borate pH 9.0. Prior to RPLC, the protein concentration is adjusted to 1 mg/mL or less, the product is heated at 95°C for 30 minutes, and cooled, and 2-propanol is added to 10% v/v. The column is then loaded. RPLC is then performed on Amberchrome resin, as described above for B/Lee. Up to 2-3 mg of protein are loaded per ml of resin. The final yield is about 4 g/500 g wet cell weight.

#### 15 EXAMPLE 19 - 3-PART INFLUENZA VACCINE

A recombinant vaccine was formulated to contain 1 μg each of the recombinant proteins NS1<sub>(1-81)</sub>HA2<sub>(65-222)</sub> (Example 11), NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub>mut5255 (Example 10), and the BC13mut2 (described in Example 15 above) in Al<sup>+3</sup> (100 μg) plus 3-o-deacylated monophosphoryl-lipid A (3D-MPL) (5 μg) [described in U.S. Patent No. 4 912 093; commercially available from Pibi

[described in U.S. Patent No. 4,912,093; commercially available from Ribi Immunochem Research, Inc., Hamilton, Montana]. Prior to inclusion in the recombinant vaccine, the influenza proteins were purified as described in Example 15 above to remove any contaminating bacterial proteins, DNA, and endotoxin.

Mice (female, CB6F<sub>1</sub>) were divided randomly into groups with 15

mice per group. The mice were injected subcutaneously on days 0 and 21 with the recombinant vaccine. A group of control mice were injected with the same dose of Al/MPL without antigen according to the same schedule. Mice were challenged with 3-5 LD50 doses of virus on day 49 and survival was monitored through day 21 post-challenge. In the following table showing these results, N.D. = not done and under the antigens, HI = NS1(1-81)HA2(65-222), H3 = NS1(1-81)H3HA2(1-221)mut5855 and B = NS1(1-81)BLHA2(1-223)mut2.

Table 5

Type A and B Cross-Protection in Mice Immunized with a Combination of Recombinant HA2 Antigens

Percent Survival	After	Challenge	with:

5	Antigen	A/Pr	·/8/34 (H1)	A/HK	Z/68 (H3) B/Lee/40 (B)	
	#1 H1/H3/	В	73*		80*	73
	H1		60*		N.D.	N.D.
	Н3		N.D.		73*	0
	В		0		7	33 <sup>1</sup>
10	con	itrol .	7		0	0
	#2 H1,	/H3/B	93*		80*	100*
	Hl		86*		N.D.	N.D.
	Н3		N.D.		53**	N.D.
15	В		N.D.		N.D.	<b>*</b> 08
	con	trol 0		7		7
					_	

<sup>\*</sup> p  $\leq$  0.001 vs. control group

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The data in Table 5 above results from two experiments that demonstrate that mice vaccinated with the combination of H1, H3, and Type B HA2 antigens were protected against all three virus challenges (H1, H3 and Type B) (>73-100% survival vs. 0-7% in controls). The H1 and H3 antigens in Al/MPL were subtype protective when administered individually as shown in Table 5. The Type B construct administered without the other antigens was only protective in one study (Exp. 1; 33% survival vs. 0% survival in controls but protected 80% of the mice in a second study, Exp. 2). Thus, preliminary data shows equivocal data on the stability of the Type B construct when formulated in Al/MPL in the absence of the other HA2 antigens. Studies are ongoing to confirm the stability of the construct in other formulations and in NIH/Swiss mice to confirm activity in an outbred system.

Although each antigen contains the NS1<sub>(1-81)</sub> regions from

A/PR/8/34 (H1) virus, protections against H1 challenge was only achieved with the
D protein which contains the H1HA2 region as well. Thus, the H3HA2 and Type B
HA2 portions of each chimeric antigen are responsible for conferring subtype-

<sup>\*\*</sup>  $p \le 0.01$  vs. control group

p > 0.05 (not statistically different than control group)

specific protection. Thus, the combined HA2 constructs provide cross-protections for all currently circulating influenza Type A (H1 and H3 subtypes) and Type B viruses.

Survival of NIH/Swiss outbred mice immunized with the mutant  $NS_{(1-81)}BHA2_{(1-223)}$  (met-leu) (not shown) showed activity at 100 micrograms (73% survival), but reduced activity at lower doses. This confirms earlier studies in outbred mice showing reduced potency relative to H1 or H3 constructs (which are active at  $\geq 1$  microgram per dose). In contrast, in CB6F1 inbred mice, an inverse dose response or no dose response is seen with  $NS_{(1-81)}BHA2_{(1-223)}$  (met-leu).

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#### EXAMPLE 20 - PLASMID pMS3H3HA

Plasmid pFV88 contains the entire 221 amino acid length HA2 from A/Udorn, an H3 subtype virus [C. J. Lai et al., Proc. Natl. Acad. Sci. USA, 77:210-214 (1980)], which HA2 nucleic acid sequence is illustrated in Fig. 7 [SEQ ID NO:

1]. This plasmid was cut with Pst I. The resulting 1900 bp fragment, which contains the entire HA (HA1 and HA2) fragment and some GC tailing, was then inserted into pUC18 [Bethesda Research Laboratories]. The resulting plasmid is termed pMS3 or pMS3H3HA.

#### 20 EXAMPLE 21 - pMG1

Plasmid pAPR801 is a pBR322-derived cloning vector which carries the NS1 coding region (A/PR/8/34). It is described by Young et al., in <u>The Origin of Pandemic Influenza Viruses</u>, ed. by W. G. Laver, Elsevier Science Publishing Co. (1983).

Plasmid pAS1 is a pBR322-derived expression vector which contains the P<sub>L</sub> promoter, an N utilization site (to relieve transcriptional polarity effects in the presence of N protein) and the cII ribosome binding site including the cII translation initiation codon followed immediately by a BamHI site. It is described by Rosenberg et al., in Methods Enzymol., 101:123-138 (1983).

Plasmid pAS1ΔEH was prepared by deleting a non-essential EcoRI-HindIII region of pBR322 origin from pAS1. A 1236 base pair BamHI fragment of pAPR801, containing the NS1 coding region in 861 base pairs of viral origin and 375 base pairs of pBR322 origin, was inserted into the BamHI site of pAS1ΔEH. The resulting plasmid, pAS1ΔEH/801 expresses authentic NS1 (230 amino acids). The plasmid has an NcoI site between the codons for amino acids 81 and 82 and an

The plasmid has an NcoI site between the codons for amino acids 81 and 82 and an NruI site 3' to the NS sequences. The BamHI site between amino acids 1 and 2 is retained.

Plasmid pMG27N, a pAS1 derivative [Mol. Cell. Biol., 5:1015-1024 (1985)], was cut with BamHI and SacI and ligated to a BamHI/NcoI fragment encoding the first 81 amino acids of NS1 from pAS1ΔEH801 and a synthetic DNA NcoI/SacI fragment of the following sequence:

5 SEQ ID NO: 10:

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**SEQ ID NO: 58:** 

#### 3'- CTAGTATACAATTGTCTATAGTTCCGGACTGACTGACTC -5'

The resulting plasmid, pMG1, allows the insertion of DNA fragments after the first 81 amino acids of NS1 in any of the three reading frames within the synthetic linker fragment followed by termination codons in all three reading frames.

#### 15 EXAMPLE 22 - pMG1H3HA

Plasmid pMG1, described above in Example 21, was digested with NcoI and XbaI, releasing a 54 bp fragment, which was discarded. pMS3H3HA, described in Example 1 above, was digested with HhaI and XbaI, and a 701 bp fragment containing the coding sequence for the HA2 subunit of influenza strain A/Udorn (H3N2) was isolated, as illustrated in Fig. 1 [SEQ ID NO: 1].

Synthetic oligonucleotides were annealed to generate an NcoI 5' overhang sequence (at the 5' end) and a HhaI 3' overhang sequence (at the 3' end). The sequence of these oligonucleotides is as follows:

SEQ ID NO: 66: 5'-CATGGGCGCCCATATGGGCATATTCGGCG-3'

SEQ ID NO: 67: 3'- CCGCGGGTATACCCGTATAAGCC -5'

The annealing reaction was performed as follows. The annealing mixture was made up of 2.5μL each of 5' oligo (1.3 μg/μL), the 3' oligo (1.2 μg/μL), and added water (15 μL) to a final volume of 20 μL. The reaction tubes were then placed in 4 mL culture tubes containing water which had been heated to 65°C for 10 minutes and allowed to cool down slowly. The tubes were then put on ice and used immediately for ligation.

This three part ligation generates pMG1H3HA2<sub>(1-221)</sub> [SEQ ID NO: 9] which codes for the first 81 amino acids of NS1 fused to four amino acids donated from the linker and amino acids 1-221 of the HA2 subunit. This sequence is illustrated in Fig. 2 [SEQ ID NO: 9 & 10]. This molecule is also designated NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO: 9 & 10].

#### EXAMPLE 23 - PREPARING SEED VIRUS AND RAISING ANTISERA

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The seed virus, A/Udorn, was prepared according to the procedures described in P. Palese and J. Schulman, <u>Virol.</u>, <u>57</u>:227-237 (1974). Briefly, this technique is as follows.

Influenza virus strain A/Udorn was inoculated in 10-day old embryonated hen's eggs into the allantoic cavity. The eggs were incubated for 24-48 hours at 35°C then chilled at 4°C overnight. A portion of the eggshell over the airsac was removed and the allantoic fluid was aseptically removed using a 10-ml syringe. The fluid was centrifuged at low speed (3,000 x g) to remove particulates. This clarified supernatant was centrifuged at high speed using an SW28 Beckman rotor at 27,000 rpm (4°C for 90 minutes), resulting in the virus pellet. The virus was resuspended in 10 mM Tris (pH 7.5) containing 100 mM NaCl, 1 mM EDTA and repelleted as before. The virus was layered on 30-60% sucrose gradient in 1 mM EDTA (NTE) and spun for 3-5 hours at 25,000 rpm. The band in the middle of the tube was withdrawn, diluted in NTE and centrifuged at 27,000 rpm for 90 minutes. The pellet was suspended in phosphate-buffered saline (PBS). These viral particles were used as immunogens for preparation of antisera.

Antisera was prepared as follows. 100-200 micrograms of purified virus in complete Freund's adjuvant was injected into the subscapula of a New Zealand White rabbit. A second injection in incomplete Freund's adjuvant was done 4 weeks later, and the animals were bled 7-10 days later.

## EXAMPLE 24 - MODIFICATION AND EXPRESSION OF H3HA2 FUSION PROTEINS

The modified nucleotide sequences encoding the H3HA2 proteins were prepared by mutating the nucleotide sequences of the fusion proteins prepared according to Example 22 above. Site directed mutagenesis using the Altered Sites System [Promega Corporation] according to the manufacturer's directions was used to change nucleotide numbers, 622, 625 and 634 (A to C) and 624, 627, and 636 (G to T) of nucleotide sequences [SEQ ID NO:9] encoding the NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> fusion protein of Fig. 3 [SEQ ID NO:10], thereby changing the codons at these regions from AGG to CGT, both encoding Arg. These changes correspond to nucleotide numbers 367, 370 and 379 (A to C) and 369, 372 and 381 (G to T) of the HA2 fragment of Fig. 7 [SEQ ID NO:1].

Fig. 2 illustrates the modified nucleotide sequences of the fusion proteins [SEQ ID NO: 58] by contrast with the nucleotide sequence [SEQ ID NO:9] of the "unmodified" fusion proteins (nucleotide changes below and amino acid

changes in above sequences of unmodified fusion protein). Mutagenesis on this sequence was carried out according to the method provided with the pSelect kit from Promega.

A. NS1(1-81)H3HA2(1-221) [SEQ ID NO: 10]

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Briefly, cloning for the mutagenesis was performed as follows. The pSelect plasmid [Promega] and pMG1H3HA2 (Example 22) were each digested with HindIII. These two plasmids were ligated together and selected on tetracycline plates. The resulting vector is pSelH3HA2. Mutagenesis was performed according to Promega's kit. The following oligonucleotide was used: SEQ ID NO:68: 5'-AAACTGTTTG AAAAAACACG TCGTCAACTG CGTGAAAATG CTGACGACAT GGGC -3'.

Clones were verified by restriction endonuclease HincII. The resulting plasmid, pSelH3HA2mut5585 was digested with NcoI and XbaI, and a 748 bp fragment coding for the H3HA2mut5585 polypeptide was isolated. pOTS208NS181 (Eco-740) was digested with NcoI and XbaI. The ligation of linear pOTS208NS181Nco and the 748 bp fragment resulted in pOTS208NS1H3mut5585 [SEQ ID NO:58]. This vector codes for the polypeptide, NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> [SEQ ID NO:10].

- B. Expression of mutated NS1(1-81)H3HA2 proteins The plasmid of A was transfected into E. coli strain AR58 [SmithKline Beecham]. Cultures are grown at 32°C to mid-log phase at which time cultures are shifted to 39.5°C for two hours. The E. coli cell pellets containing the recombinant polypeptide are then stored at -70°C until used. Production of the NS1(1-81)H3HA2(1-221) protein [SEQ ID NO:10] is confirmed by Western blot analysis [Towbin et al., Proc. Natl. Acad. Sci. U.S.A., 76:4350 (1979)] using antisera prepared against A/Udorn virus, as described in Example 23. A major immunoreactive species is expected at a molecular weight of approximately 35,00 daltons.
- The expression levels obtained are about 50-100% higher than those obtained by the expression of the unmodified coding sequences in the same expression system.

# EXAMPLE 25 - tRNA INSERTION INTO HOST CELLS EXPRESSING H3 PROTEIN

E. coli host cells containing H3N2 fusion protein obtained as described in Example 22 above were transformed using conventional techniques. See, e.g. Sambrook et al, cited above.

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Briefly, a culture of E. coli strain MM294cI<sup>+</sup> containing the plasmid pDC952 was grown overnight in Luria broth with chloramphenicol. The plasmid pDC952 carries the argU gene which encodes the tRNA that recognizes the AGA/AGG codons [P. Saxena and J. Walker, J. Bacteriol., 174(6):1956-1964 (Mar. 1992)]. From this culture the plasmid pDC952 was prepared. A second culture of E. coli, strain AR13 [SmithKline Beecham] carrying the plasmid for the H3 flu antigen, was grown overnight in Luria broth with kanamycin. These cells were made competent for transformation as described below.

The H3/AR13 overnight culture was diluted 1:50 in LB and kanamycin (50 mL total) and incubated at 37°C until it reached an O.D.650 of 0.6. The culture was then transferred to a 50 mL conical tube and chilled at about 4°C. Following this, the tube was centrifuged in a TJ6 centrifuge (10 min; 2000-3000 rev/min), the pellet resuspended in 25 mL 100 mM CaCl<sub>2</sub>, and placed on ice for about 30 minutes. The pellet was then centrifuged as described above and resuspended in about 2.5 mL 100 mM CaCl<sub>2</sub>.

The competent cells were aliquoted (100 µl) into three separate sterile tubes. The first tube was the negative control and did not receive any DNA. The second tube was a positive control and 1 µl of plasmid pT7II was added to the cells. To the third tube was added 3 µl of pDC952. These controls served to ensure that transformation occurred. Each tube of cells was mixed, placed on ice for 60 min., heat shocked at 37°C in a water bath for 2 minutes, and incubated in a 32°C water bath for 60 min. after adding 1 mL LB. The tubes were then microfuged for 1 minute and the supernatants poured off until only about 200 µL were left. The pellets were then resuspended in the remaining supernatant and plated as follows:

(1) on LB and chloramphenicol, (2) on LB and ampicillin, and (3) on LB and

Shake flasks were inoculated with the control strain, H3/AR13, and 4 transformants, pDC952/H3/AR13, and grown at 32°C to an optical density of 0.6 to 0.7 at which point the cultures were shifted to 39.5°C for 3 hours. Samples were taken at induction start (temperature shift to 39.5°C) and 3 hours post-induction. These samples were analyzed by high performance liquid chromatography (HPLC) and Western blotting.

chloramphenicol and kanamycin. The plates were then incubated at 32°C overnight.

The results of these analyses indicated that expression of H3 had increased by as much as 80% and the presence of the argU gene had eliminated the lowest western positive band as compared with the wild-type constructs (H3/AR13). It is believed that these results were obtained by eliminating the frameshifting caused by tandem AGG rare arginine codons. Further, there did not appear to be any difference in product quality between the H3 mutant prepared according to Example 24, and the argU tRNA transformants made according to this Example.

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Numerous modifications and variations of the present invention are included in the above-identified specification and are expected to be obvious to one of skill in the art. Such modifications and alterations to the compositions and processes of the present invention are believed to be encompassed in the scope of the claims appended hereto.

#### SEQUENCE LISTING

5	(1) GENE	RAL INFORMATION:
•	(i)	APPLICANT: Shatzman, Allan Scott, Miller Dillon, Susan B. Kane, James
10	(ii)	TITLE OF INVENTION: Vaccinal Polypeptides
	(iii)	NUMBER OF SEQUENCES: 72
15	(iv)	CORRESPONDENCE ADDRESS:  (A) ADDRESSEE: SmithKline Beecham Corporation - Corporate Patents  (B) STREET: U.S. Mailcode UW2220 - 709 Swedeland Road
20		(C) CITY: King of Prussia (D) STATE: Pennsylvania (E) COUNTRY: USA (F) ZIP: 19406-2799
25	(v)	COMPUTER READABLE FORM:  (A) MEDIUM TYPE: Floppy disk  (B) COMPUTER: IBM PC compatible  (C) OPERATING SYSTEM: PC-DOS/MS-DOS  (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
30	(vi)	CURRENT APPLICATION DATA:  (A) APPLICATION NUMBER:  (B) FILING DATE:  (C) CLASSIFICATION:
35	(vii)	PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 149,150  (B) FILING DATE: 05-NOV-1993
40	(vii)	PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 013,415  (B) FILING DATE: 01-FEB-1993
45		PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 108,914  (B) FILING DATE: 18-AUG-1993
50	(vii)	PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 837,773  (B) FILING DATE: 18-FEB-1992
50	(vii)	PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 751,896  (B) FILING DATE: 30-AUG-1991

5	(vii)	PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 387,200  (B) FILING DATE: 28-JUL-1989	•
J	(vii)	PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 238,801  (B) FILING DATE: 02-NOV-1988	
10	(vii)	PRIOR APPLICATION DATA:  (A) APPLICATION NUMBER: US 645,732  (B) FILING DATE: 30-AUG-1984	
15	(viii)	ATTORNEY/AGENT INFORMATION:  (A) NAME: Baumeister, Kirk  (B) REGISTRATION NUMBER: 33,833  (C) REFERENCE/DOCKET NUMBER: P50134 PCT	
20	(ix)	TELECOMMUNICATION INFORMATION: (A) TELEPHONE: 215-270-5096 (B) TELEFAX: 215-270-5090	
25		RMATION FOR SEQ ID NO:1:  SEQUENCE CHARACTERISTICS:  (A) LENGTH: 666 base pairs	
30		(B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: unknown	
35		MOLECULE TYPE: DNA (genomic)  FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1663	
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:1:	
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45		GAC GGT TGG TAC GGT TTC AGG CAT CAA AAT TCT GAG GGC ACA Asp Gly Trp Tyr Gly Phe Arg His Gln Asn Ser Glu Gly Thr 20 25 30	96
50		GCA GCA GAT CTT AAA AGC ACT CAA GCA GCC ATC GAC CAA ATC Ala Ala Asp Leu Lys Ser Thr Gln Ala Ala Ile Asp Gln Ile 35 40 45	144
55		AAA CTG AAT AGG GTA ATC GAG AAG ACG AAC GAG AAA TTC CAT Lys Leu Asn Arg Val Ile Glu Lys Thr Asn Glu Lys Phe His 55 60	192

			GAA Glu														240
5			TAC Tyr														288
10			CTT Leu														336
15			ATG Met 115												GAA Glu		384
20			GAG Glu														432
			GCT Ala														480
25			AGA Arg												GTT Val		528
30 .			AAG Lys													,	576
35			TGC Cys 195														624
40	Ala		CAG Gln										TGA	•			666
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	Met	Ile	Asp	Gly 20	Trp	Tyr	Gly	Phe	Arg 25	His	Gln	Asn	Ser	Glu 30	Gly	Thr
5	Gly	Gln	Ala 35	Ala	Asp	Leu	Lys	Ser 40	Thr	Gln	Ala	Ala	Ile 45	Asp	Gln	Ile
	Asn	Gly 50	Lys	Leu	Asn	Arg	Val 55	Ile	Glu	Lys	Thr	Asn 60	Glu	Lys	Phe	His
10	Gln 65	Ile	Glu	Lys	Glu	Phe 70	Ser	Glu	Val	Glu	Gly 75	Arg	Ile	Gln	Asp	Leu 80
15	Glu	Lys	Tyr	Val	Glu 85	Asp	Thr	Lys	Ile	Asp 90	Leu	Trp	Ser	Tyr	Asn 95	Ala
	Glu	Leu	Leu	Val 100	Ala	Leu	Glu	Asn	Gln 105	His	Thr	Ile	Asp	Leu 110	Thr	Asp
20	Ser	Glu	Met 115	Asn	Lys	Leu	Phe	Glu 120	Lys	Thr	Arg	Arg	Gln 125	Leu	Arg	Glu
	Asn	Ala 130	Glu	Asp	Met	Gly	<b>As</b> n 135	Gly	Cys	Phe	Lys	Ile 140	Tyr	His	Lys	Cys
25	Asp 145	Asn	Ala	Cys	Ile	Gly 150	Ser	lle	Arg	Asn	Gly 155	Thr	Tyr	Asp	His	Asp 160
30	Val	Tyr	Arg	Asp	Glu 165	Ala	Leu	Asn	Asn	Arg 170	Phe	Gln	Ile	Lys	Gly 175	Val
	Glu	Leu	Lys	Ser 180	Gly	Tyr	Lys	Asp	Trp 185	Ile	Leu	Trp	Ile	Ser 190	Phe	Ala
35	Ile	Ser	Cys 195	Phe	Leu	Leu	Cys	Val 200	Val	Leu	Leu	Gly	Phe 205	Ile	Met	Trp
	Ala	Cys 210	Gln	Lys	Gly	Asn	Ile 215	Arg	Cys	Asn	Ile	Cys 220	Ile			
40	(2)	INFO	ORMA	MOIT	FOR	SEQ	ID N	10:3:	:							
		(i)	(2	A) LE	ENGTI	ARAC i: 66 nucl	66 ba	ise p	airs	3						
45						DEDNE DGY :			ole							
		(ii)	MOI	LECUI	LE TY	PE:	DNA	(ger	omic	:)						
50		(ix)		A) N#	ME/F	CEY:		563								

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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20												AGA Arg						240
25												TGG Trp					,	288
30												ATT Ile						336
												AGG Arg						384
35	Asn	Ala 130	Glu	Asp	Met	Gly	Asn 135	Gly	Cys	Phe	Lys	ATA Ile 140	Tyr	His	Lys	Суѕ		432
40	Asp 145	Asn	Ala	Cys	Ile	Gly 150	Ser	Ile	Arg	Asn	Gly 155	ACT Thr	Tyr	Asp	His	Asp 160		480
45												CAG Gln						528
50												TGG Trp						576
												GGG Gly			-			624

							ATT Ile 215							TGA				666
5	(2)	INFO	ORMA!	rion	FOR	SEQ	ID 1	NO : 4	<b>:</b>									
10	` '			SEQUI	ENCE LEI	CHAI NGTH	RACTI : 22	ERIS	rics: ino a		5							•
10							amino GY:											
		(:	ii) l	MOLE	CULE	TYP	E: p:	rote:	in	٠								
15		. (2	xi) :	SEQUI	ENCE	DES	CRIP'	NOI	: SE(	) ID	NO:	4:						
	Gly 1	Ile	Phe	Gly	Ala 5	Ile	Ala	Gly	Phe	Ile 10	Glu	Asn	Gly	Trp	Glu 15	Gly		
20	Met	Ile	Asp	Gly 20	Trp	Tyr	Gly	Phe	Arg 25	His	Gl'n	Asn	Ser	Glu 30	Gly	Thr		
25	Gly	Gln	Ala 35	Ala	Asp	Leu	Lys	Ser 40	Thr	Gln	Ala	Ala	Ile 45	Asp	Gln	Ile		•
23	Asn	Gly 50	Lys	Leu	Asn	Arg	Val 55	Ile	Glu	Lys	Thr	Asn 60	Glu	Lys	Phe	His		
30	Gln 65	Ile	Glu	Lys	Glu	Phe 70	Ser	Glu	Val	Glu	Gly 75	Arg	Ile	Gln	Asp	Leu 80		
	Glu	Lys	Tyr	Val	Glu 85	Asp	Thr	Lys	Ile	Asp 90	Leu	Trp	Ser	Tyr	Asn 95	Ala		
35	Glu	Leu	Leu	Val. 100	Ala	Leu	Glu	Asn	Gln 105	His	Thr	Ile	Asp	Leu 110	Thr	Asp		
40	Ser	Glu	Met 115	Asn	Lys	Leu	Phe	Glu 120	Lys	Thr	Arg	Arg	Gln 125	Leu	Arg	Glu		
10	Asn	Ala 130	Glu	Asp	Met	Gly	Asn 135	Gly	Cys	Phe	Lys	Ile 140	Tyr	His	Lys	Суз	٠	
45	Asp 145	Asn	Ala	Cys	Ile	Gly 150	Ser	Ile	Arg	Asn	Gly 155	Thr	Tyr	Asp	His	Asp 160		
	Val	Tyr	Arg	Asp	Glu 165	Ala	Leu	Asn	Asn	Arg 170	Phe	Gln	Ile	Lys	Gly 175	Val		•
50	Glu	Leu	Lys	Ser 180	Gly	Tyr,	Lys	Asp	Trp 185	Ile	Leu	Trp	Ile	Ser 190	Phe	Ala		
	Ile	Ser	Cys 195	Phe	Leu	Leu	Cys	Val 200	Val	Leu	Leu	Gly	Phe 205	Ile	Met	Trp		

	Ala	Cys 210	Gln	Lys	Gly	Asn	Ile 215	Arg	Cys	Asn	Ile	Cys 220	Ile				
5	(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO:5	:								
		(i)	() ()	A) LI B) T: C) S:	engti YPE : Irani	HARA( H: 6 nuc DEDNI	70 ba leic ESS:	ase pacio	pair: d	s							
10		42.11	•			OGY:								•			
		(11)	MQ.	LECU.	LE T	YPE:	DNA	(ge	nomi	C)							
15		(ix)	()		AME/I	KEY: ION:		666									
20		(xi)	SE	QUEN	CE DI	ESCR:	IPTI(	ON:	SEQ :	ID N	0:5:						
		CTA Leu														•	4 8
25		ATA Ile													TCA Ser	. 9	96
30	, GGC	TAT														14	4 4
35		AAC Asn 50														19	∋2
40		GTG Val														24	<b>3</b> C
40		AAA Lys														28	3 8
45		TTG Leu														33	3 6
50		AAT Asn														38	} 4
55		GCC Ala 130														43	32

		Asn											TAT Tyr				480
5						•							GTA Val				528
10													ATC Ile				576
15													GCA Ala 205				624
20													TGC Cys				666
	TGA	G															670
25	(2)	INF		rion Seque													
30				(A) (B)	LEN TYN	NGTH: PE: 6	: 222 amino GY: 2	am: ac: linea	ino a id ar		8						
		()	ki) 5	EQUE	ENCE	DESC	CRIPT	ION:	: SE(	ID	NO:	5:					
35	Gly 1	Leu	Phe	Gly	Ala 5	Ile	Ala	Gly	Phe	Ile 10	Glu	Gly	Gly	Trp	Thr 15	Gly	
40	Met	Ile	Asp	Gly 20	Trp	Tyr	Gly	Tyr	His 25	His	Gln	Asn	Glu	Gln 30	С¦ІУ	Ser	
	Gly	Tyr	Ala 35	Ala	Asp	Gln	Lys	Ser 40	Thr	Gl'n	Asn	Ala	Ile 45	Asn	Gly	Ile	
45	Thr	Asn 50	Lys	Val	Asn	Ser	Val 55	Ile	Glu	Lys	Met	Asn 60	Ile	Gln	Phe	Thr	
	Ala 65	Val	Gly	Lys	Glu	Phe 70	Asn	Lys	Leu	Glu	Lys 75	Arg	Met	Glu	Asn	Leu 80	
50	Asn	Lys	Lys	Val	Asp 85	Asp	Gly	Phe	Leu	Asp 90	Ile	Trp	Thr	Tyr	Asn 95	Ala	

	Glu	Leu	Leu	Val 100	Leu	Leu	Glu	Asn	Glu 105	Arg	Thr	Leu	Asp	Phe 110	His	Asp		
5	Ser	Asn	Val 115	Lys	Asn	Leu	Tyr	Ğlu 120	Lys	Val	Lys	Ser	Gln 125	Leu	Lys	Asn		
	Asn	Ala 130	Lys	Glu	Ile	Gly	Asn 135	Gly	Cys	Phe	Glu	Phe 140	Tyr	His	Lys	Cys		
10	Asp 145	Asn	Glu	Cys	Met	Glu 150	Ser	Val	Arg	Asn	Gly 155	Thr	Tyr	Asp	Tyr	Pro 160		
15	Lys	Туг	Ser	Glu	Glu 165	Ser	Lys	Leu	Asn	Arg 170	Glu	Lys	Val	Asp	Gly 175	Val		
13	Lys	Leu	Glu	Ser 180	Met	Gly	Ile	Tyr	Gln 185	Ile	Leu	Ala	Ile	Tyr 190	Ser	Thr		
20	Val	Ala	Ser 195		Leu	Val	Leu	Leu 200	Val	Ser	Leu	Gly	Ala 205	Ile	Ser	Phe		
	Trp	Met 210	-	Ser	Asn	Gly	Ser 215	Leu	Gln	Cys	Arg	Ile 220	Cys	Ile				
25	(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO:7	:									
30		(i	()	QUENCA) LIB) TC) SiD) TC	ENGT: YPE : TRAN	H: 6' nuc	70 ba leic ESS:	ase p acid doul	pair: d	s							•	
		(11	) MO	LECU	LE T	YPE:	DNA	(ge	nomi	c)								
35		(ix	(	ATUR A) N B) L	AME/			670										
40										ID N		ccc	NCCC	አአጥ	C n m n	GACGGT	· 6	
																AAAAGC		
45																ACGAAC		
	GAG	AAAT	TCC	ATCA	AATC	GA A	aagg	AATT	C TC	AGAA	GTAG	AAG	GGAG	AAT	TCAG	GACCTO	24	
	GAG	TAAA	ACG	TTGA	AGAC	AC T	AAAA	TAGA	т ст	CTGG	TCTT	ACA	ATGC	GGA	GCTT	CTTGTC	30	
50	GCI	CTGG	AGA	ACCA	ACAT	AC A	ATTG	ATCT	g ac	TGAC	TCGG	AAA	TGAA	CAA	ACTG	TTTGAA	36	
	AAA	ACAA	GGA	GGCA	ACTG	AG G	GAAA	ATGC	T GA	GGAC	atgg	GCA	atgg	TTG	CTTC	ATAAAA	42	
55	ጥልና	ירארא	ידעע	СТСА	ת ב ב	'GC 파	ጥርር አ	тасс	G TC	ል ልጥር	AGAA	ATC	GGAC	тта	тсас	САТСАТ	r 48	

	GTATACAGAG ACGAAGCATT AAACAACCGG TTTCAGATCA AAGGTGTTGA ACTGAAGTCA	
_	GGATACAAAG ACTGGATCCT GTGGATTTCC TTTGCCATAT CATGCTTTTT GCTTTGTGTT	
5	GTTTTGCTGG GGTTCATCAN NNTGTGGGCC TGCCANAAAG GCAACATTAG GTGCAACATT	
	TGCATTTGAN	
10	(2) INFORMATION FOR SEQ ID NO:8:  (i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 222 amino acids  (B) TYPE: amino acid	
15	(C) STRANDEDNESS: unknown (D) TOPOLOGY: unknown	
	(ii) MOLECULE TYPE: DNA (genomic)	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
20	Gly Ile Phe Gly Ala Ile Ala Gly Phe Ile Glu Asn Gly Trp Glu Gly 1 10 15	
25	Met Ile Asp Gly Trp Tyr Gly Phe Arg His Gln Asn Ser Glu Gly Thr 20 . 25 30 .	
	Gly Gln Ala Ala Asp Leu Lys Ser Thr Gln Ala Ala Ile Asp Gln Ile 35 40 45	
30	Asn Gly Lys Leu Asn Arg Val Ile Glu Lys Thr Asn Glu Lys Phe His 50 55 60	
35	Gln Ile Glu Lys Glu Phe Ser Glu Val Glu Gly Arg Ile Gln Asp Leu 65 70 75 80	
30	Glu Lys Tyr Val Glu Asp Thr Lys Ile Asp Leu Trp Ser Tyr Asn Ala 85 90 95	
40	Glu Leu Leu Val Ala Leu Glu Asn Gln His Thr Ile Asp Leu Thr Asp 100 105 110	
	Ser Glu Met Asn Lys Leu Phe Glu Lys Thr Arg Arg Gln Leu Arg Glu 115 120 125	
45	Asn Ala Glu Asp Met Gly Asn Gly Cys Phe Lys Ile Tyr His Lys Cys 130 135 140	
<b>5</b> 0	Asp Asn Ala Cys Ile Gly Ser Ile Arg Asn Gly Thr Tyr Asp His Asp 145 150 155 160	
50	Val Tyr Arg Asp Glu Ala Leu Asn Asn Arg Phe Gln Lle Lys Gly Val 165 170 175	
55	Glu Leu Lys Ser Xaa Gly Tyr Lys Asp Trp Ile Leu Trp Ile Ser Phe 180 185 190	

Ala Ile Ser Cys Phe Leu Leu Cys Val Val Leu Leu Gly Phe Ile Met

				195	5				200	)				20	5		
5		Trp	Ala 210	_	Glr	ı Lys	s Gly	/ Asi 21:		e Arq	g Cys	s Asr	n Ile 220		s Il	е	
10	(2)	INFO	RMAT	NOI	FOR	SEQ	ID N	10:9	<b>:</b>				,				
	*	(i)	( P	) LE	engti	1: 91	CTERI 18 ba	se p	pairs	<b>s</b> .							
15			(C	) SI	rani	DEDNE	leic ESS: unkr	doub									
		(ii)	MOI	LECUI	LE T	ME:	DNA	(ger	nomi	<b>3</b> ),							
20		(ix)	(P	1) N2	AME/E		CDS								•		
25		(xi)	SEC	UENC	CE DE	ESCR	IPTIC	on: S	SEQ :	ID NO	9:						
23	ATG Met																48
30	CAT																. 96
35	CTT Leu																144
40	ACT Thr		Gly														192
4.5	GTG Val 65				Leu												240
45	ATG Met																288
50	AAT Asn		Trp		Gly	Met		Asp	Gly	Trp	Tyr	Gly	Phe	Arg	His		336

			GAG Glu 115											Thr		GCA Ala	384
5			GAC Asp													ACG Thr	432
10			AAA Lys														480
15			CAG Gln														528
20			TAC Tyr														576
			CTG Leu 195														624
25	AGG Arg	CAA Gln 210	CTG Leu	AGG Arg	GAA Glu	AAT Asn	GCT Ala 215	GAG Glu	GAC Asp	ATG Met	GGC Gly	AAT Asn 220	GGT Gly	TGC Cys	TTC Phe	AAA Lys	672
30	ATA Ile 225	TAC Tyr	CAC His	AAA Lys	TGT Cys	GAC Asp 230	AAT Asn	GCT Ala	TGC Cys	ATA Ile	GGG Gly 235	TCA Ser	ATC Ile	AGA Arg	AAT Asn	GGG Gly 240	720
35			GAC Asp														768
40	CAG Gln	ATC Ile	AAA Lys	GGT Gly 260	GTT Val	GAA Glu	CTG Leu	AAG Lys	TCA Ser 265	GGA Gly	TAC Tyr	AAA Lys	GAC Asp	TGG Trp 270	ATC Ile	CTG Leu	816
	TGG Trp	ATT Ile	TCC Ser 275	TTT Phe	GCC Ala	ATA Ile	TCA Ser	TGC Cys 280	TTT Phe	TTG Leu	CTT Leu	TGT Cys	GTT Val 285	GTT Val	TTG Leu	CTG Leu	864
45	GGG Gly	TTC Phe 290	ATC Ile	ATG Met	TGG Trp	GCC Ala	TGC Cys 295	CAA Gln	AAA Lys	GGC Gly	AAC Asn	ATT Ile 300	AGG Arg	TGC Cys	AAC Asn	ATT Ile	912
50	TGC Cys 305													•			918

#### (2) INFORMATION FOR SEQ ID NO:10:

#### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 306 amino acids

5 (B) TYPE: amino acid

(D) TOPOLOGY: linear

#### (ii) MOLECULE TYPE: protein

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp

1 10 15

His Val Arg Lys Arg Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe 20 25 30

Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu Arg Gly Arg Gly Ser

Thr Leu Gly Leu Asp Ile Glu Thr Ala Thr Arg Ala Gly Lys Gln Ile
50 55 60

Val Glu Arg Ile Leu Lys Glu Glu Ser Asp Glu Ala Leu Lys Met Thr 25 65 70 75 80

Met Gly Ala His Met Gly Ile Phe Gly Ala Ile Ala Gly Phe Ile Glu 85 90 95

30 Asn Gly Trp Glu Gly Met Ile Asp Gly Trp Tyr Gly Phe Arg His Gln
100 105 110

Asn Ser Glu Gly Thr Gly Gln Ala Ala Asp Leu Lys Ser Thr Gln Ala 115 120 125

Ala Ile Asp Gln Ile Asn Gly Lys Leu Asn Arg Val Ile Glu Lys Thr
130 135 140

Asn Glu Lys Phe His Gln Ile Glu Lys Glu Phe Ser Glu Val Glu Gly 40 145 150 155 160

Arg Ile Gln Asp Leu Glu Lys Tyr Val Glu Asp Thr Lys Ile Asp Leu 165 170 175

45 Trp Ser Tyr Asn Ala Glu Leu Leu Val Ala Leu Glu Asn Gln His Thr 180 185 190

50

Ile Asp Leu Thr Asp Ser Glu Met Asn Lys Leu Phe Glu Lys Thr Arg 195 200 205

Arg Gln Leu Arg Glu Asn Ala Glu Asp Met Gly Asn Gly Cys Phe Lys 210 215 220

Ile Tyr His Lys Cys Asp Asn Ala Cys Ile Gly Ser Ile Arg Asn Gly 225 230 235 240

,	Thr	Tyr	Asp	His	Asp 245	Val	Tyr	Arg	Asp	Glu 250	Ala	Leu	Asn	Asn	Arg 255	Phe		
5	Gln	Ile	Lys	Gly 260	Val	Glu	Leu	Lys	Ser 265	Gly	Tyr	Lys	Asp	Trp 270	Ile	Leu		
	Trp	Ile	Ser 275	Phe	Ala	Ile	Ser	Cys 280	Phe	Leu	Leu	Cys	Val 285	Val	Leu	Leu		
10	Gly	Phe 290	Ile	Met	Trp	Ala	Cys 295	Gln	Lys	Gly	Asn	Ile 300	Arg	Cys	Asn	Ile		
15	Cys 305	Ile					•								•			
	(2)	INFO	ORMA!	rion	FOR	SEQ	ID N	NO:1	ι:									
20		(1)	()	A) Li B) T	engti YPE :	HARAC H: 69 nucl	90 ba Leic	ase p	pair:	5								
25	(C) STRANDEDNESS: double (D) TOPOLOGY: unknown  (ii) MOLECULE TYPE: DNA (genomic)  (ix) FEATURE:																	
30		(ix)	(2	A) N	AME/I	KEY: ION:		690										
		(xi	) SE	QUEN	CE DI	ESCR:	IPTIC	ON: S	SEQ :	ID NO	0:11	:						
<b>3</b> 5						GTG Val											,	48
40	_					GTT Val										_		96
4.5						CGA Arg	-										1	144
45						ATC Ile											:	192
50						AAA Lys 70											2	240

						ATT Ile											288
	1160	nsp		nec	85		GIII	nsp	neu	90		ıyı	Val	GIU	95	inr	
5						TCT											336
	Lys	Ile	Asp	Leu 100		Ser	Tyr	Asn	Ala 105	Glu	Leu	Leu	Val	Ala 110		Glu	
10						GAT											384
10	ASN	GID	115		TIE	Asp	Leu	120	Asp	Ser	Glu	Met	Asn 125	Lys	Leu	Phe	
	GAA	AAA	ACA	AGG	AGG	CAA	CTG	AGG	GAA	AAT	GCT	GAG	GAC	ATG	GGC	TĄA	432
15	GIU	130	Inr	Arg	Arg	Gln	135	Arg	GIU	Asn	Ala	G1u 140	Asp	Met	Gly	Asn	
	GGT	TGC	TTC	AAA	ATA	TAC	CAC	AAA	TGT	GAC	AAT	GCT	TGC	ATA	GGG	TCA	480
20	145					Tyr 150		٠			155				_	160	
	ATC	AGA	AAT	GGG	ACT	TAT	GAC	CAT	GAT	GTA.	TAC	AGA	GAC	GAA	GCA	TTA	528
	•				165	Tyr				170					175	•	
25	AAC	AAC	CGG	TTT	CAG	ATC Ile	AAA	GGT	GTT	GAA	CTG	AAG	TCA	GGA	TAC	AAA	576
				180					185				•	190		_	
30	GAC Asp	TGG	ATC	CTG	TGG	ATT Ile	TCC	TTT	GCC	ATA	TCA	TGC	TTT	TTG	CTT	TGT	624
			195					200					205			-	
	Val	Val	Leu	Leu	Gly	TTC Phe	Ile	ATG Met	TGG	GCC	TGC	CAA Gln	AAA Lvs	GGC	AAC	ATT	672
35		210	AAC				215		•			220	-,, 5	or,		116	
			Asn													•	690
40	225					230											
	(2)					SEQ						•					
45		(	(i) S	(A) (B)	LEN TYP	CHAR GTH: E: a OLOG	230 mino	ami aci	no a d								
<b>5</b> 0		<b>(</b> i	.i) M			TYPE											
50		(х	i) S	EQUE	NCE	DESC	RIPT	ION:	SEQ	ID 1	NO:1	2:					
	Met	Asp	Pro	Asn	Thr	Val	Ser	Ser :	Phe		Val .	Asp .	Cys	Phe	Leu '	Trp	
55	1	•			5					10	•				15		

	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe
5	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser
	Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile
10	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Thr 80
15	Met	Asp	His	Met	Leu 85	Ile	Gln	Asp	Leu	Glu 90	Lys	Tyr	Val	Glu	Asp 95	Thr
	Lys	Ile	Asp	Leu 100	Trp	Ser	Tyr	Asn	Ala 105	Glu	Leu	Leu	Val	Ala 110	Leu	Glu
20	Asn	Gln	His 115	Thr	Ile	Asp	Leu	Thr 120	Asp	Ser	Glu	Met	Asn 125	Lys	Leu	Phe
	Glu	Lys 130	Thr	Arg	Arg		Leu 135	Arg	Glu	Asn	Ala	Glu 140	Asp	Met	Gly	Asn
25	145	_		Lys		150		.'	_		155					160
30				Gly	165			•		170		•			175	
				Phe 180	•				185					190		
35	_		195	Leu				200					205			
4.0		210		Leu			11e 215	Met	Trp	Ala	Суѕ	Gln 220	Lys	Gly	Asn	Ile
40	Arg 225	Cys	Asn	Ile	Cys	11e 230			•							
45	(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO:1	3:							
		(i	•	QUEN( A) L						3						

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double

50

- (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)

#### (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..699

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13: ATG GAT CCA AAC ACT GTG TCA AGC TTT CAG GTA GAT TCC TTT CTT TGG Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Ser Phe Leu Trp 5 10 10 CAT GTC CGC AAA CGA GTT GCA GAC CAA GAA CTA GGT GAT GCC CCA TTC 96 His Val Arg Lys Arg Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe 20 15 CTT GAT CGG CTT CGC CGA GAT CAG AAA TCC ATG CAT GGA TCA TAT GTT Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Met His Gly Ser Tyr Val 40 AAC AAG ACA CAA GAA GCT ATA AAC AAG ATA ACA AAA AAT CTC AAC TAT . 192 20 Asn Lys Thr Gln Glu Ala Ile Asn Lys Ile Thr Lys Asn Leu Asn Tyr 50 55 TTA AGT GAG CTA GAA GTA AAA AAC CTT CAA AGA CTA AGC GGA GCA ATG 240 Leu Ser Glu Leu Glu Val Lys Asn Leu Gln Arg Leu Ser Gly Ala Met 25 AAT GAG CTT CAC GAC GAA ATA CTC GAG CTA GAC GAA AAA GTG GAT GAT 288 Asn Glu Leu His Asp Glu Ile Leu Glu Leu Asp Glu Lys Val Asp Asp 85 95 30 CTA AGA GCT GAT ACA ATA AGC TCA CAA ATA GAG CTT GCA GTC TTG CTT 336 Leu Arg Ala Asp Thr Ile Ser Ser Gln Ile Glu Leu Ala Val Leu Leu 100 TCC AAC GAA GGG ATA ATA AAC AGT GAA GAT GAG CAT CTC TTG GCA CTT 35 384 Ser Asn Glu Gly Ile Ile Asn Ser Glu Asp Glu His Leu Leu Ala Leu 115 120 GAA AGA AAA CTG AAG AAA ATG CTT GGC CCC TCT GCT GTA GAA ATA GGG 432 40 Glu Arg Lys Leu Lys Lys Met Leu Gly Pro Ser Ala Val Glu Ile Gly 130 135 AAT GGG TGC TTT GAA ACC AAA CAC AAA TGC AAC CAG ACT TGC CTA GAC 480 Asn Gly Cys Phe Glu Thr Lys His Lys Cys Asn Gln Thr Cys Leu Asp 45 145 150 AGG ATA GCT GCT GGC ACC TTT AAT GCA GGA GAT TTT TCT CTT CCC ACT 528 Arg Ile Ala Ala Gly Thr Phe Asn Ala Gly Asp Phe Ser Leu Pro Thr 165 170 50 TTT GAT TCA TTA AAC ATT ACT GCT GCA TCT TTA AAT GAT GGC TTG 576 Phe Asp Ser Leu Asn Ile Thr Ala Ala Ser Leu Asn Asp Asp Gly Leu 180 185

				Thr										Ser		TTG Leu	624
5																AGA Arg	672
10		Asn				TCC Ser 230											699
15	(2)			SEQUI	ENCE	SEQ CHAI NGTH PE:	RACTI : 23	ERIS:	TICS		s						
20			•	(D)	TO:	POLO: TYP: DES:	SY: :	line:	ar in	Q ID	NO:	14:					
25	Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10	Val	Asp	Ser	Phe	Leu 15	Trp	
30				20					25			Gly	_	30			
			35					40				His	45		_		
35		50					55					Lys 60				•	
• •	65					70					75	Leu				80	
40					85					90		Glu			95	-	
45	,			100					105			Leu		110			
			115	•				120				His	125				•
50	Glu	Arg 130	Lys	Leu	Lys	Lys	Met 135	Leu	Gly	Pro	Ser	Ala 140	Val	Glu	Ile	Gly	
	Asn 145	Gly	Cys	Phe	Glu	Thr 150	Lys	His	Lys	Cys	Asn 155	Gln	Thr	Cys	Leu	Asp 160	

	Arg	, Ile	e Ala	ı Ala	Gly 165		Phe	ASI	n Ala	170		Phe	e Ser	Let	175	Thr	
5	Phe	Asp	Ser	Leu 180		Ile	Thr	Ala	185		. Le	a Asr	a Asp	190	_	/ Leu	
	Asp	Asr	His 195	Thr	Ile	` <b>Le</b> u	Leu	Ту: 200		: Ser	Thi	Ala	Ala 205		: Ser	Leu	
10	Ala	Val 210	Thr	Leu	Met	Ile	Ala 215		Phe	: Ile	e Val	220		Val	. Ser	Arg	
15	Asp 225		val	Ser	Cys	Ser 230		Cys	Leu	1							
	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:1	5:								
20		(i	(	QUENA) L B) T C) S D) T	engt Ype : Tran	H: 9 nuc DEDN	24 b leic ESS:	ase aci dou	pair d ble	s							
25		(ii	) MO	LECU	LE T	YPE:	DNA	(ge	nomi	c)						,	
30			() ()	ATURI A) N. B) Lo QUENO	AME/I	ION:	1		SEO	ID N	0:15	•					
35	ATG Met	GAT	CCA	AAC Asn	ACT	GTG	TCA	AGC	TTT	CAG	GTA	GAT	TGC Cys	TTT Phe	CTT Leu	TGG Trp	48
40	CAT His	GTC Val	CGC Arg	AAA Lys 20	CGA Arg	GȚT Val	GCA Ala	GAC Asp	CAA Gln 25	GAA Glu	CTA Leu	GGT Gly	GAT Asp	GCC Ala 30	CCA Pro	TTC Phe	96
	CTT Leu	GAT Asp	CGG Arg 35	CTT Leu	CGC Arg	CGA Arg	GAT Asp	CAG Gln 40	AAA Lys	TCC Ser	CTA Leu	AGA Arg	GGA Gly 45	AGG Arg	GGC Gly	AGC Ser	144
45	ACT Thr	CTT Leu 50	GGT Gly	CTG Leu	GAC Asp	ATC Ile	GAG Glu 55	ACA <sup>.</sup> Thr	GCC	ACA Thr	CGT Arg	GCT Ala 60	GGA Gly	AAG Lys	CAG Gln	ATA Ile	192
50	GTG Val 65	GAG Glu	CGG Arg	ATT Ile	CTG Leu	AAA Lys 70	GAA Glu	GAA Glu	TCC Ser	GAT Asp	GAG Glu 75	GCA Ala	CTT Leu	AAA Lys	ATG Met	ACC Thr 80	240
55	ATG Met	GAT Asp	CTG Leu	TCC Ser	AGA Arg 85	GGT Gly	CTA Leu	TTT Phe	GGA Gly	GCC Ala 90	ATT Ile	GCC Ala	GGT Gly	TTT Phe	ATT Ile 95	GAA Glu	288

_		GGA Gly														CAG Gln	336
5		GAA Glu															384
10		ATT Ile 130															432
15		ATT															480
20		ATG Met															528
25		ACA Thr															576
	CTG Leu	GAT Asp	TTC Phe 195	CAT His	GAC Asp	TCA Ser	AAT Asn	GTG Val 200	AAG Lys	AAT Asn	CTG Leu	TAT Tyr	GAG Glu 205	AAA Lys	GTA Val	AAA Lys	624
30	AGC Ser	CAA Gln 210	TTA Leu	AAG Lys	AAT Asn	AAT Asn	GCC Ala 215	AAA Lys	GAA Glu	ATC Ile	GGA Gly	AAT Asn 220	GGA Gly	TGT Cys	TTT Phe	GAG Glu	672
35	TTC Phe 225	TAC Tyr	CAC His	AAG Lys	TGT Cys	GAC Asp 230	AAT Asn	GAA Glu	TGC Cys	ATG Met	GAA Glu 235	AGT Ser	GTA Val	AGA Arg	AAT Asn	GGG Gly 240	720
40	ACT Thr	TAT Tyr	GAT Asp	TAT Tyr	CCC Pro 245	AAA Lys	TAT Tyr	TCA Ser	GAA Glu	GAG Glu 250	TCA Ser	AAG Lys	TTG Leu	AAC Asn	AGG Arg 255	GAA Glu	768
45	AAG Lys	GTA Val	GAT Asp	GGA Gly 260	GTG Val	AAA Lys	TTG Leu	GAA Glu	TCA Ser 265	ATG Met	GGG Gly	ATC Ile	TAT Tyr	CAG Gln 270	ATT Ile	CTG Leu	816
	GCG Ala	ATC Ile	TAC Tyr 275	TCA Ser	ACT Thr	GTC Val	GCC Ala	AGT Ser 280	TCA Ser	CTG Leu	GTG Val	CTT Leu	TTG Leu 285	GTC Val	TCC Ser	CTG Leu	864
50	GGG Gly	GCA Ala 290	ATC Ile	AGT Ser	TTC Phe	Trp	ATG Met 295	TGT Cys	TCT Ser	AAT Asn	GGA Gly	TCT Ser 300	TTG Leu	CAG Gln	TGC Cys	AGA Arg	912

924

		e Cy:	C ATO	C TG2	ł.							•					
5												ė.					
	(2)	INE	ORM	40ITA	FOF	R SE(	] ID	NO:1	16:								
10			(i)	( <i>P</i>	L) LE	NGTI PE:	d: 30 amir	TERIS 07 am no ac line	nino cid		is						·
15		(	(ii)	MOLE	CULE	TYF	E: p	prote	in			,					
20		(	(xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	16:					
	Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10		Asp	Cys	Phe	Leu 15	Trp	
25	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe	
	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45		Gly	Ser	
30	Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile	
35	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Thr 80	
	Met	Asp	Leu	Ser	Arg 85	Gly	Leu	Phe	Gly	Ala 90	Ile	Ala	Gly	Phe	Ile 95	Glu	
40	Gly	Gly	Trp	Thr 100	Gly	Met	Ile	Asp	Gly 105	Trp	Tyr	Gly	Tyr	His 110	His	Gln	
	Asn	Glu	Gln 115	Gly	Ser	Gly	Tyr	Ala 120	Ala	Asp	Gln	Lys	Ser 125	Thr	Gln	Asn	,
45	Ala	Ile 130	Asn	Gly	Ile	Thr	Asn 135	Lys	Val	Asn	Ser	Val 140	Ile	Glu	Lys	Met	
50	Asn 145	Ile	Gln	Phe	Thr	Ala 150	Val	Gly	Lys	Glu	Phe 155	Asn	Lys	Leu	Glu	Lys 160	
	Arg	Met	Glu	Asn	Leu 165	Asn	Lys	Lys	Val	Asp 170	Asp	Gly	Phe	Leu	Asp 175	Ile	
55	Trp	Thr	Tyr	Asn 180	Ala	Glu	Leu	Leu	Val 185	Leu	Leu	Glu	Asn	Glu 190	Arg	Thr	

	Leu	Asp	Phe 195	His	Asp	Ser	Asn	Val 200	Lys	Asn	Leu	Tyr	Glu 205	Lys	Val	Lys	
5	Ser	Gln 210	Leu	Lys	Asn	Asn	Ala 215	Lys	Glu	Ile	Gly	Asn 220	Gly	Суз	Phe	Glu	
10	Phe 225	Tyr	His	Lys	Cys	Asp 230	Asn	Glu	Cys	Met	Glu 235	Ser	Val	Arg	Asn	Gly 240	
	Thr	Tyr	Asp	Tyr	Pro 245	Lys	Tyr	Ser	Glu	Glu 250	Ser	Lys	Leu	Asn	Arg 255	Glu	
15	Lys	Val	Asp	Gly 260	Val	Lys	Leu	Glu	Ser 265	Met	Gly	Ile <sub>.</sub>	Tyr	Gln 270	Ile	Leu	
	Ala	Ile	Tyr 275	Ser	Thr	Val	Ala	Ser 280	Ser	Leu	Val	Leu	Leu 285	Val	Ser	Leu	
20	Gly	Ala 290	Ile	Ser	Phe	Trp	Met 295	Cys	Ser	Asn	Gly	Ser 300	Leu	Gln	Cys	Arg	
25	Ile 305	Суs	Ile														
	(2)	INFO	ORMA:	rion	FOR	SEQ	ID 1	10:17	7:		ı						
30		(i)	() ()	A) LE B) T	engti (PE : [rani	nucl	?9 ba Leic ESS:	ISTIC ase p acid doub nown	oair:	s							
35		(ii)	MOI	LECUI	LE T	PE:	DNA	(ger	nomi	c)							
40		(ix)	(2	ATURI A) NI B) L(	AME/I			726									
		(xi)	SE(	QUEN	CE DI	ESCRI	[PTI	ON: S	SEQ :	ID NO	0:17:	•					
45		GAT Asp															. 48
50		GTC Val															96
		GAT Asp															144

													GGA Gly				192
		50	0-,				55				9	60	,	-,0	<b>0</b>		
<b>5</b> ,	GTG	GAG	CGG	ATT	CTG	AAA	GAA	GAA	TCC	GAT	GAG	GCA	CTT	AAA	ATG	ACC	240
		Glu	Arg	Ile	Leu	_	Glu	Glu	Ser	Asp		Ala	Leu	Lys	Met		
	65					70					75					80	
	ATG	CAG	ATC	CCG	GCT	GTG	GGT	AAA	GAA	TTC	AAC	AAA	TTA	GAA	AAA	AGG	288
10	Met	Gln	Ile	Pro	Ala 85	Val	Gly	Lys	Glu	Phe 90	Asn	Lys	Leu	Glu	Lys 95	Arg	
	ATG	GAA	AAT	TTA	AAT	AAA	AAA	GTT	GAT	GAT	GGA	TTT	CTG	GAC	ATT	TGG	336
																Trp	
15				100					105					110		•	
													GAA				384
	Thr	Tyr		Ala	Glu	Leu	Leu		Leu	Leu	Glu	Asn	Glu	Arg	Thr	Leu	
20			115					120					125				
	GAT	TTC	CAT	GAC	TCA	AAT	GTG	AAG	AAT	CTG	TAT	GAG	AAA	GTA	AAA	AGC	432
	Asp		His	Asp	Ser	Asn		Lys	Asn	Leu	Tyr	Glu	Lys	Val	Lys	Ser	
		130					135					140					
25	CAA	TTA	AAG	AAT	AAT	GCC	AAA	GAA	ATC	GGA	AAT	GGA	TGT	TTT	GAG	TTC	480
	Gln												Суз				
	145					150					155					160	
													AGA				528
30	Tyr	His	Lys	Cys		Asn	Glu	Cys	Met		Ser	Val	Arg	Asn			
					165					170					175		
													AAC				576
2.5	Tyr	Asp	Tyr		Lys	Tyr	Ser	Glu		Ser	Lys	Leu	Asn	_	Glu	Lys	
35				180					185					190			
	GTA	GAT	GGA	GTG	AAA	TTG	GAA	TCA	ATG	GGG	ATC	TAT	CAG	ATT	CTG	GCG	624
	Val	Asp		Val	Lys	Leu	Glu		Met	Gly	Ile	Tyr	Gln	Ile	Leu	Ala	
40			195					200					205				
40	ATC	TAC	TCA	ACT	GTC	GCC	AGT	TCA	CTG	GTG	СТТ	TTG	GTC	TCC	СТС	ക്ക	672
													Val				072
		210					215				•	220				-	
45	GCA	ATC	AGT	TTC	TGG	ATG	TGT	TCT	AAT	GGA	TCT	TTG	CAG	TGC	AGA	ATA	720
	Ala					Met							Gln				
	225					230					235					240	
	TGC	ATC	TGA											•			729
50	Cys	Ile															

	(2)	INFC	RMA	LION	FUR	SEQ	IU N	10:10	) <b>.</b>							
5	·	(	(i) S	(A)	LEN TYI	CHAF NGTH: PE: &	242	ami aci	.no a .d		<b>;</b>					
10		·	•			TYPE	-			) ID	NO:1	18:				
15	Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10	Val	Asp	Cys	Phe	Leu 15	Trp
13	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe
20	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser
	Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile
25	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Thr 80
30	Met	Gln	Ile	Pro	Ala 85	Val	Gly	Lys	Glu	Phe 90	Asn	Lys	Leu	Glu	Lys 95	Arg
	Met	Glu	Asn	Leu 100	Asn	Lys	Lys	Val	Asp 105	Asp	Gly	Phe	Leu	Asp 110	Ile	Trp
35	Thr	Tyr	Asn 115	Ala	Glu	Leu	Leu	Val 120	Leu	Leu	Glu	Asn	Glu 125	Arg	Thr	Leu
	Asp	Phe 130	His	Asp	Ser	Asn	Val 135	Lys	Asn	Leu	Tyr <sub>.</sub>	Glu 140	Lys	Val	Lys	Ser
40	Gln 145	Leu	Lys	Asn	Asn	Ala 150	Lys	Glu	Ile	Gly	Asn 155	Gly	Cys	Phe	Glu	Phe 160
45	Tyr	His	Lys	Cys	Asp 165		Glu	Суз	Met	Glu 170	Ser	Val	Arg	Asn	Gly 175	Thr
	Tyr	Asp	Tyr	Pro 180	Lys	Tyr	Ser	Glu	Glu 185	Ser	Lys	Leu	Asn	Arg 190	Glu	Lys
<b>5</b> 0	Val	Asp	Gly 195	Val	Lys	Leu	Glu	Ser 200	Met	Gly	Ile	Tyr	Gln 205	Ile	Leu	Ala
	Ile	Tyr 210		Thr	Val	Ala	Ser 215	Ser	Leu	Val	Leu	Leu 220	Val	Ser	Leu	Gly

	Ala 225		e Ser	Phe	e Trp	230		Se:	Asr	r Gly	235		Glr	ı Cys	s Arg	1le 240		
5	Cys	Ile	:															
	(2)	INF	ORMA	TION	FOR	SEC	ID	NO:1	9:				•					
10		(i	(	A) I B) T C) S	ENGT YPE: TRAN	HARA H: 8 nuc DEDN	10 b leic ESS:	ase aci dou	pair .d .ble	s .								
15		(ii	) мо	LECU	LE T	YPE:	DNA	(ge	nomi	.c)								
20			(	B) L	AME/ OCAT	KEY: ION:	1	807						,				
						ESCR												
25	ATG Met 1	GAT Asp	CCA Pro	AAC Asn	ACT Thr 5	Val	TCA Ser	AGC Ser	TTT	CAG Gln 10	GTA Val	GAT Asp	TGC Cys	TTT	CTT Leu 15	TGG Trp		48
30	CAT His	GTC Val	CGC Arg	AAA Lys 20	Arg	GTT Val	GCA Ala	GAC Asp	CAA Gln 25	Glu	CTA Leu	GGT Gly	GAT Asp	GCC Ala 30	CCA Pro	TTC Phe		96
30	CTT	GAT Asp	CGG Arg 35	CTT	CGC	CGA Arg	GAT Asp	CAG Gln 40	AAA Lys	TCC Ser	ATG Met	GAT Asp	CTG Leu 45	TCC Ser	AGA Arg	GGT Gly	,	144
35	CTA Leu	TTT Phe 50	GGA Gly	GCC Ala	ATT	GCC Ala	GGT Gly 55	TTT Phe	ATT	GAA Glu	GGG Gly	GGA Gly 60	TGG Trp	ACT Thr	GGA Gly	ATG Met		192
40	ATA Ile 65	GAT Asp	GGA Gly	TGG Trp	TAC Tyr	GGT Gly 70	TAT Tyr	CAT His	CAT His	CAG Gln	AAT Asn 75	GAA Glu	CAG Gln	GGA Gly	TCA Ser	GGC Gly 80		240
45	TAT Tyr	GCA Ala	GCG Ala	GAT Asp	CAA Gln 85	AAA Lys	AGC Ser	ACA Thr	CAA Gln	AAT Asn 90	GCC Ala	ATT Ile	AAC Asn	GGG Gly	ATT Ile 95	ACA Thr	•	288
50	AAC Asn	AAG Lys	GTG Val	AAC Asn 100	TCT Ser	GTT Val	ATC Ile	GAG Glu	AAA Lys 105	ATG Met	AAC Asn	ATT Ile	CAA Gln	TTC Phe 110	ACA Thr	GCT Ala		336
	GTG Val	GGT Gly	AAA Lys 115	GAA Glu	TTC Phe	AAC Asn	AAA Lys	TTA Leu 120	GAA Glu	AAA Lys	AGG Arg	ATG Met	GAA Glu 125	AAT Asn	TTA Leu	AAT Asn		384

5		AAA Lys 130															432
J		TTA Leu															480
10		GTG Val															528
15		AAA Lys												_	-		576
20		GAA Glu															624
25		TCA Ser 210															672
		GAA Glu															720
30		AGT Ser															768
35		TGT Cys												TGA			810
40	(2)	INFO		SEQUE	ENCE	CHAF	<b>VACTE</b>	RIST	rics:							٠.	,
45				(B) (D)	TYP	POLOG	mino SY: 1	aci inea	r.	·	<b>;</b>						
				OLEC SEQUE			•		n SEÇ	) ID	NO:2	!O:					
50	1	Asp			5				•	10					15		
55	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe	

		Leu	ASP	35	ьeu	Arg	Arg	ASP	40		Ser	Met	Asp	45		: Arg	Gly
	, 5	Leu	Phe 50	Gly	Ala	Ile	Ala	Gly 55		Ile	Glu	Gly	Gly 60		Thr	Gly	Met
•		Ile 65	Asp	Gly	Trp	Tyr	Gly 70		His	His	Gln	Asn 75		Gln	Gly	Ser	Gly 80
	10	Tyr	Ala	Ala	Asp	Gln 85	Lys	Ser	Thr	Gln	Asn 90	Ala	Ile	Asn	Gly	Ile 95	Thr
	15				Asn 100			•		105					110		
				115	Glu				120					125			
	20	Lys	Lys 130	Val	Asp	Asp	Gly	Phe 135	Leu	Asp	Ile	Trp	Thr 140	Tyr	Asn	Ala	Glu
		Leu 145	Leu	Val	Leu	Leu	Glu 150	Asn	Glu	Arg	Thr	Leu 155	Asp	Phe	His	Asp	Ser 160
	25	Asn	Val	Lys	Asn	Leu 165	Tyr	Glu	Lys	Val	Lys 170	Ser	Gln	Leu	Lys	Asn 175	Asn
•	30				Ile 180					185					190		
		Asn	Glu	Cys 195	Met	Glu	Ser	Val	Arg 200	Asn	Gly	Thr	Tyr	Asp 205	Tyr	Pro	Lys
	35	Tyr	Ser 210	Glu	Glu	Ser	Lys	Leu 215	Așn	Arg	Glu	Lys	Val 220	Asp	Gly	Val	Lys
		Leu 225	Glu	Ser	Met	Gly	Ile 230	Tyr	Gln	Ile	Leu	Ala 235	Ile	Tyr	Ser	Thr	Val 240
	40	Ala	Ser	Ser	Leu	Val 245	Leu	Leu	Val	Ser	Leu 250	Gly	Ala	Ile	Ser	Phe 255	Trp
	45	Met	Cys	Ser	Asn 260	Gly	Ser	Leu	Gln	Cys 265	Arg	Ile	Cys	Ile			•
		(2)	INFO	RMAT	CION	FOR	SEQ	ID N	10:21	:							
	50		(i)	(A (B (C	UENC ) LE ) TY :) ST	NGTH PE: RAND	: 63 nucl EDNE	0 ba eic SS:	se p acid doub	airs							
	55		(ii)	MOL	ECUL	E TY	PE:	DNA	(gen	omic	)		i				

PCT/US94/01149 WO 94/17826

### (ix) FEATURE:

5

(A) NAME/KEY: CDS

(B) LOCATION: 1..627

J		(xi)	SEÇ	UENC	CE DE	ESCRI	PTIC	ON: S	SEQ I	D NO	0:21:	:			
10					ACT Thr										48
15					CGA Arg										96
13					CGC Arg										144
20					GTG Val										192
25					AAA Lys									ACA Thr 80	240
30					TTG Leu 85										288
35					AAT Asn										336
40					GCC Ala										384
••					AAT Asn										432
45	-	Tyr			TAT Tyr										480
50					TTG Leu 165	Glu					Tyr				528
55					GCC Ala										576

-80-

				Trp					Gly					Arg		TGC Cys		624
5	ATC Ile	TGA																630
10	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:2	2:									
15			(i)	(B	) LE ) TY	NGTH PE:	RACT : 20 amin GY:	9 am. o ac:	ino . id		s							
		(.	ii)	MOLE	CULE	TYP	E: p	rote:	in									
20		<b>(</b> :	xi)	SEQU	ENCE	DES	CRIP'	rion	: SE	QI Q	NO:	22:						
	Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10		Asp	Суз	Phe	Leu 15	Trp		
25	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe		
30	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Met	Asp	His 45	Met	Leu	Thr	·	
	Ser	Thr 50	Arg	Ser	Val	Gly	Lys 55	Glu	Phe	Asn	Lys	Leu 60	Glu	Lys	Arg	Met		
<b>3</b> 5	Glu 65	Asn	Leu	Asn	Lys	Lys 70	Val	Asp	Asp	Gly	Phe 75	Leu	Asp	Ile	Trp	Thr 80		
40	Tyr	Asn	Ala	Glu	Leu 85	Leu	Val	Leu	Leu	Glu 90	Asn	Glu	Arg	Thr	Leu 95	Asp		
	Phe	His	Asp	Ser 100	Asn	Val	Lys	Asn	Leu 105	Tyr	Glu	Lys	Val	Lys 110	Ser	Gln		
45	Leu	Lys	Asn 115	Asn	Ala	Lys	Glu	Ile 120	Gly	Asn	Gly	Cys	Phe 125	Glu	Phe	Tyr		
	His	Lys 130	Cys	Asp	Asn	Glu	Cys 135	Met	Glu	Ser	Val	Arg 140	Asn	Gly	Thr	Tyr		
50	Asp 145	Tyr	Pro	Lys	Tyr	Ser 150	Glu	Glu	Ser	Lys	Leu 155	Asn	Arg	Glu	Lys	Val 160		
55	Asp	Gly	Val	Lys	Leu 165	Glu	Ser	Met	Gly	Ile 170	Tyr	Gln	Ile	Leu	Ala 175	Ile		

Tyr Ser Thr Val Ala Ser Ser Leu Val Leu Leu Val Ser Leu Gly Ala 180 185 190

5	Ile	Ser	Phe 195	Trp	Met	, Cys	Ser	Asn 200	Gly	Ser	Leu	Gln	Cys 205	Arg	Ile	Cys	
	Ile							•									
10	(2)	INF	ORMAT	CION	FOR	SEQ	ID 1	NO:23	3:								
15		(i)	(E	A) LE 3) TY C) ST	ENGTI (PE : (RANI		17 ba Leic ESS:	ase p acid doub	pairs i	3							
		(ii)	MOI	LECUI	LE T	YPE:	DNA	(gei	nomic	=)							
20		(ix)	•	A) NI	AME/I	KEY:		714									
25 -		(xi	) SE(	QUENC	CE DI	ESCR:	[PTI	ON:	SEQ :	ED NO	23:	:					,
	-		CCA Pro														48
30			CGC Arg														96
35			CGG Arg 35														144
40			GGT Gly														192
45			CGG Arg														240
- •			ATC Ile														288
50			AAA Lys														336

		TTG														GAC Asp		384
5		AAT Asn 130																432
10		GCC Ala																480
15		AAT Asn																528
20		TAT Tyr																576
25	AAA Lys	TTG Leu	GAA Glu 195	TCA Ser	ATG Met	GGG Gly	ATC Ile	TAT Tyr 200	CAG Gln	ATT Ile	CTG Leu	GCG Ala	ATC Ile 205	TAC Tyr	TCA Ser	ACT Thr		624
20		GCC Ala 210															•	672
30		ATG Met																714
35	TGA	•																717
40	(2)	INFO		EQUE (A) (B)	NCE LEM	SEQ CHAR IGTH: PE: a	ACTE 238 mino	RIST ami	TICS: .no a		3	,						
45						TYPE				) ID	NO:2							
50	1	Asp			5					10					15	-		
		Val		20					25					30				
55	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser		

	Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile
5	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Àla	Leu	Lys	Met	Thr 80
10	Met	Gln	Ile	Pro	Glu 85	Phe	Asn	Lys	Leu	Glu 90	Lys	Arg	Met <sub>.</sub>	Glu	Asn 95	Leu
	Asn	Lys	Lys	Val 100	Asp	Asp	Gly	Phe	Leu 105	Asp	Ile	Trp		Tyr 110	Asn ,	Ala
15	Glu	Leu	Leu 115	Val	Leu	Leu	Glu	Asn 120	Glu	Arg	Thr	Leu	Asp 125	Phe	His	Asp
	Ser	Asn 130	Val	Lys	Asn	Leu	Tyr 135	Glu	Lys	Val	Lys	Ser 140	Gln	Leu	Lys	Asn
20	Asn 145	Ala	Lys	Glu	Ile	Gly 150	Asn	Gly	Cys	Phe	Glu 155	Phe	Tyr	His	Lys	Cys' 160
25	Asp	Asn	Glu	Суѕ	Met 165	Glu	Ser	Val	Arg	Asn 170	Gly	Thr	Tyr	Asp	Tyr 175	Pro
23	(Lys	Tyr	Ser	Glu 180	Glu	Ser	Lys	Leu	Asn 185	Arg	Glu	Lys	Val	Asp 190	Gly	Val
30	Lys	Leu	Glu 195	Ser	Met	Gly	Ile	Tyr 200	Gln	Ile	Leu	Ala	Ile 205	Tyr	Ser	Thr
	Val	Ala 210	Ser	Ser	Leu	Val	Leu 215	Leu	Val	Ser	Leu	Gly 220	Ala	Ile	Ser	Phe
35	Trp 225	Met	Cys	Ser	Asn	Gly 230	Ser	Leu	Gln	Cys	Arg 235	Ile	Суз	Ile		
40	(2)	INF	ORMA'	rion	FOR	SEQ	ID I	NO:2	5 :		•					
		(i	G	QUENC A) Li B) T	ENGT	H: 6	81 ba	ase p	pair	5						
45			((	C) S'	TRAN	DEDNI	ESS:	doul								
٠		. <b>(i</b> i	) MO	LECU:	LE T	YPE:	DNA	(ge	nomi	c)						
50		(ix	΄ (.	ATURI A) N B) L	AME/I			678								

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

																•	
5		Asp				Val					Val					TGG	4.8
10					Arg					Glu					Pro	TTC Phe	96
. <b>- v</b>									Lys					Arg		AGC Ser	144
15			Gly										Gly			ATA	192
20						AAA Lys 70											. 240
25	ATG Met	CAG Gln	ATC Ile	CCG Pro	AAT Asn 85	AAA Lys	AAA Lys	GTT Val	GAT Asp	GAT Asp 90	GGA Gly	TTT Phe	CTG Leu	GAC Asp	ATT Ile 95	TGG Trp	288
30	ACA Thr	TAT Tyr	AAT Asn	GCA Ala 100	GAA Glu	TTG Leu	TTA Leu	GTT Val	CTA Leu 105	CTG Leu	GAA Glu	AAT Asn	GAA Glu	AGG Arg 110	ACT Thr	CTG Leu	336
	GAT Asp	TTC Phe	CAT His 115	GAC Asp	TCA Ser	AAT Asn	GTG Val	AAG Lys 120	AAT Asn	CTG Leu	TAT Tyr	GAG Glu	AAA Lys 125	GTA Val	AAA Lys	AGC Ser	384
35	CAA Gln	TTA Leu 130	AAG Lys	AAT Asn	AAT Asn	GCC Ala	AAA Lys 135	GAA Glu	ATC Ile	GGA Gly	AAT Asn	GGA Gly 140	TGT Cys	TTT Phe	GAG Glu	TTC Phe	432
40	TAC Tyr 145	CAC His	AAG Lys	TGT Cys	GAC Asp	AAT Asn 150	GAA Glu	TGC Cys	ATG Met	GAA Glu	AGT Ser 155	GTA Val	AGA Arg	AAT Asn	GGG Gly	ACT Thr 160	480
45	TAT Tyr	GAT Asp	TAT Tyr	CCC Pro	AAA Lys 165	TAT Tyr	TCA Ser	GAA Glu	GAG Glu	TCA Ser 170	AAG Lys	TTG Leu	AAC Asn	AGG Arg	GAA Glu 175	AAG Lys	528
<b>E</b> 0	GTA Val	GAT Asp	GGA Gly	GTG Val 180	AAA Lys	TTG Leu	GAÁ Glu	TCA Ser	ATG Met 185	GGG Gly	ATC Ile	TAT Tyr	CAG Gln	ATT Ile 190	CTG Leu	GCG Ala	576
50	ATC Ile	TAC Tyr	TCA Ser 195	ACT Thr	GTC Val	GCC Ala	AGT Ser	TCA Ser 200	CTG Leu	GTG Val	CTT Leu	TTG Leu	GTC Val 205	TCC Ser	CTG Leu	GGG Gly	624

												TTG Leu 220						672
5		ATC Ile	TGA															681
10	(2)	INF	ORMA!	rion	FOR	SEQ	ID 1	10:2	6:									
15			(i) S	(B)	LEI TYI	NGTH:	RACTE : 226 amino GY: 1	am:	ino a Id		s							
		(:	ii) 1	MOLE	CULE	TYP	E: p1	ote:	Ln									
20		. (3	κi) S	SEQUI	ENCE	DESC	CRIPT	CION	: SE(	Q ID	NO:	26:						
	Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10	Val	Asp	Суз	Phe	Leu 15	Trp		
25	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe		
	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser		
30	Thr	Leu 50	Gly	Leu	Asp	Ile	G1u 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile		
35	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Thr 80		
	Met	Gln	Ile	Pro	Asn 85	Lys	Lys	Val	Asp	Asp 90	Gly	Phe	Leu	Asp	Ile 95	Trp		
40	Thr	Tyr	Asn	Ala 100	Glu	Leu	Leu	Val	Leu 105	Leu	Glu	Asn	Glu	Arg 110	Thr	Leu	_	
	Asp	Phe	His 115	Asp	Ser	Asn	Val	Lys 120	Asn	Leu	Tyr	Glu	Lys 125	Val	Lys	Ser		
45	Gln	Leu 130	Lys	Asn	Asn	Ala	Lys 135	Glu	Ile	Gly	Asn	Gly 140	Cys	Phe	Glu	Phe		
50	Tyr 145	His	Lys	Cys	Asp	Asn 150	Glu	Cys	Met	Glu	Ser 155	Val	Arg	Asn	Gly	Thr 160	٠	
	Tyr	Asp	Tyr	Pro	Lys 165		Ser	Glu				Leu		-	Glu	-		

	vai	Asp	_	180	гÃа	ьеп	GIU	ser	185	СТУ	TIE	Tyr	GIN	190	ren	ATG	
5	Ile	Tyr	Ser 195	Thr	Val	Ala		Ser 200	Leu	Val	Leu	Leu	Val 205	Ser	Leu	Gly	
	Ala	Ile 210	Ser	Phe	Trp	Met	Cys 215	Ser	Asn	Gly	Ser	Leu 220	Gln	Cys	Arg	Ile	
10	Cys 225	Ile			•												
	(2)	INFO	RMAT	ION	FOR	SEQ	ID N	0:27	:			į.					
15		<b>(</b> 1)	(B	) LE	ngth Pe:	: 15 amin	TERI 8 am o ac unkn	ino id		s							
20		(ii,)	MOL	ECUL	E TY	PE:	prot	ein									
		(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	ĖŌ I	D NO	:27:						
25		Met 1	, Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10	Val	Asp	Cys	Phe	Leu 15	Tr
		His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Ph
30		Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Se
35		Thr	Leu 50	Gly	Leu	Asp	Ile	G1u 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	110
		Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Th:
40	٠	Met	Gln	Ile	Pro	Val 85	Glu	Ser	Val	Arg	Asn 90	Gly	Thr	Tyr	Asp	Tyr 95	Pro
		Lys	Tyr	Ser	Glu 100	Glu	Ser	Lys	Leu	Asn 105	Arg	Glu	Lys	Val	Asp 110	Gly	Va:
45		Lys	Leu	Glu 115	Ser	Met	Gly	Ile	Tyr 120		Ile	Leu	Ala	Ile 125		Ser	Th
50		Val	Ala 130	Ser	Ser	Leu	Val	Leu 135	Leu	Val	Ser	Leu	Gly 140	Ala	Ile	Ser	Phe
		Trp 145	Met	Cys ,	Ser	Asn	Gly 150	Ser	Leu	Gln	Суз	Arg 155	Ile	Cys	Ile		

	(2)	INFO	TAMS	ON E	OR S	SEQ I	ID NO	28	:								
5		(i)	(B)		NGTH:	: 163 amino	am:	lno a id	S: acids	6							
		(ii)	MOLE	CULE	E TYP	PE: p	prote	ein			•						
10		(xi)	SEQU	JENCE	EDES	SCRIE	PTIO	N: SI	II QE	ОИС	:28:						
		Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10	Val	Asp	Cys	Phe	Leu 15	Trp
15		His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe
20		Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser
		Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile
25		Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	<b>Glu</b> 75	Ala	Leu	Lys	Met	Thr 80
30		Met	Asp	Leu	Ser	Arg 85	Gly	Leu	Phe	Gly	Ala 90	Ile	Ala	Gly	Phe	Ile 95	Glu
30		Gly	Gly	Trp	Thr 100	Gly	Met	Ile	Asp	Gly 105	Trp	Tyr	Gly	Tyr	His 110	His	Gln
35		Asn	Glu	Gln 115	Gly	Ser	Gly	Tyr	Ala 120	Ala	Asp	Gln	Lys	Ser 125	Thr	Glņ	Asn
		Ala	Ile 130	Asn	Gly	Ile	Thr	Asn 135	Lys	Val	Asn	Ser	Val 140	Ile	Glu	Lys	Met
40		Asn 145	Ile	Gln	Phe	Thr	Ala 150	Val	Gly	Lys	Glu	Phe 155	Ser	Cys	Leu	Thr	Ala 160
		Tyr	His	Arg													
45	(2)	INFO	RMAT:	ION 1	FOR :	SEQ :	ID N	0:29	:								
50		(i)	(B		NGTH PE: a	: 23	l am	ino d	S: acid	5						·	
		(ii)	MOL	ECUL	E TY	PE: 1	DNA	(gen	omic	)							

	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:29:						
5	Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10	Val	Asp	Cys	Phe	Leu 15	Trp
-	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe
10	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser
	Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile
15	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Thr 80
20	Met	Gln	Ile	Pro	Ala 85	Val	Gly	Lys	Glu	Phe 90	Asn	Lys	Leu	Glu	Lys 95	Arg
	Met	Glu	Asn	Leu 100	Asn	Lys	Lys	Val	Asp 105	Asp	Gly	Phe	Leu	Asp 110	Ile	Trp
25 .	Thr	Tyr	<b>As</b> n 115	Ala	Glu	Leu	Leu	Val 120	Leu	Leu	Glu	Asn	Glu 125	Arg	Thr	Leu
•	Asp	Phe 130	His	Asp	Ser	Asn	Val 135	Lys	Asn	Leu	Tyr	Glu 140	Lys	Val	Lys	Ser
30	Gln 145	Leu	Lys	Asn	Asn	Ala 150	Lys	Glu	Ile	Gly	Asn 155	Gly	Cys	Phe	Glu	Phe 160
35	Tyr	His	Lys	Суз	<b>A</b> sp 165	Asn	Glu	Cys	Met	Glu 170	Ser	Val	Arg	Asn	Gly 175	Thr
	Tyr	Asp	Tyr	Pro 180	Lys	Tyr	Ser	Glu	Glu 185	Ser	Lys	Leu	Asn	Arg 190	Glu	Lys
40	Val	Asp	Gly 195	Val	Lys	Leu	Glu	Ser 200	Met	Gly	Ile	Tyr	Gln 205	Ile	Leu	Ala
	Ile	Tyr 210	Ser	Thr	Val	Ala	Ser 215	Ser	Gly	Gly	Ser	<b>Tyr</b> 220	Ser	Met	Glu	His
45	Phe 225	Arg	Trp	Gly	Lys	Pro 230	Val									

## (2) INFORMATION FOR SEQ ID NO:30: (i) SEQUENCE CHARACTERISTICS: 5 (A) LENGTH: 225 amino acids (B) TYPE: amino acid (D) TOPOLOGY: unknown (ii) MOLECULE TYPE: protein 10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30: Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp 15 His Val Arg Lys Arg Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu Arg Gly Arg Gly Ser 20 Thr Leu Gly Leu Asp Ile Glu Thr Ala Thr Arg Ala Gly Lys Gln Ile 55 25 Val Glu Arg Ile Leu Lys Glu Glu Ser Asp Glu Ala Leu Lys Met Thr 75 Met Gln Ile Pro Ala Val Gly Lys Glu Phe Asn Lys Leu Glu Lys Arg . 85 30 Met Glu Asn Leu Asn Lys Lys Val Asp Asp Gly Phe Leu Asp Ile Trp 105 Thr Tyr Asn Ala Glu Leu Leu Val Leu Leu Glu Asn Glu Arg Thr Leu 35 Asp Phe His Asp Ser Asn Val Lys Asn Leu Tyr Glu Lys Val Lys Ser 40 Gln Leu Lys Asn Asn Ala Lys Glu Ile Gly Asn Gly Cys Phe Glu Phe 145 150 155 Tyr His Lys Cys Asp Asn Glu Cys Met Glu Ser Val Arg Asn Gly Thr 165 170 45 Tyr Asp Tyr Pro Lys Tyr Ser Glu Glu Ser Lys Leu Asn Arg Glu Lys

215

185

Val Asp Gly Val Lys Leu Glu Ser Met Gly Ile Tyr Gln Ile Leu Ala

Ile Tyr Ser Thr Val Ala Ser Ser Gly Gly Ser Tyr Ser Met Leu Val

200

180

50

Asn 225

5																		
	(2)	INF	ORMA	TION	FOR	SEC	ID	NO:3	11:									
10		(i	(	A) L B) T C) S	ENGT YPE : TRAN	H: 9 nuc DEDN	CTER 12 b :leic ESS: unk	ase aci dou	pair d ble	·s								
15			) FE.	ATUR A) N	E: AME/	KEY:	CDS		nomi	c)								
20		(xi)	) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:31	:						
25	ATG Met	GAT Asp	CCA Pro	AAC Asn	ACT Thr 5	GTG Val	TCA Ser	AGC Ser	TTT Phe	CAG Gln 10	GTA Val	GAT Asp	TGC Cys	TTT Phe	CTT Leu 15	TGG Trp	4	8
	CAT His	GTC Val	CGC Arg	AAA Lys 20	CGA Arg	GTT Val	GCA Ala	GAC Asp	CAA Gln 25	GAA Glu	CTA Leu	GGT Gly	GAT Asp	GCC Ala 30	CCA Pro	TTC Phe	9	6
30	CTT (	GAT Asp	CGG Arg 35	CTT Leu	CGC Arg	CGA Arg	GAT Asp	CAG Gln 40	AAA Lys	TCC Ser	CTA Leu	AGA Arg	GGA Gly 45	AGG Arg	GGC	AGC Ser	14	4
35	ACT (	CTT Leu 50	GGT Gly	CTG Leu	GAC Asp	ATC Ile	GAG Glu 55	ACA Thr	GCC Ala	ACA Thr	CGT Arg	GCT Ala 60	GGA Gly	AAG Lys	CAG Gln	ATA Ile	19:	2
40	GTG ( Val ( 65	GAG Glu	CGG Arg	ATT Ile	CTG Leu	AAA Lys 70	GAA Glu	GAA Glu	TCC Ser	GAT Asp	GAG Glu 75	GCA Ala	CTT Leu	AAA Lys	ATG Met	ACC Thr 80	240	0
45	ATG (	CAG Gln	ATC Ile	CCG Pro	GGT Gly 85	CTA Leu	TTT Phe	GGA Gly	GCC Ala	ATT Ile 90	GCC Ala	GGT Gly	TTT Phe	ATT Ile	GAA Glu 95	GGG Gly	288	3
	GGA 1	rgg Frp	ACT Thr	GGA Gly 100	ATG Met	ATA Ile	GAT Asp	GGA Gly	TGG Trp 105	TAC Tyr	GGT Gly	TAT Tyr	CAT	CAT His 110	Gln	AAT Asn	336	5
50	GAA (	31n	GGA Gly 115	TCA Ser	GGC Gly	TAT Tyr	GCA Ala	GCG Ala 120	GAT Asp	CAA Gln	AAA Lys	AGC Ser	ACA Thr 125	CAA Gln	AAT Asn	GCC Ala	384	}

						GTT Val				432
5			Ala			AAC Asn 155				480
10	 					GGA Gly				528
15						GAA Glu				576
20						TAT Tyr				624
						AAT Asn				672
25						AGT Ser 235				720
30						AAG Lys			 ı	768
35						ATC Ile				816
40						CTT Leu				864
						TCT Ser				912
45										

#### (2) INFORMATION FOR SEQ ID NO:32:

50

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 304 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

5	Met 1		Pro	) Asn	Thr 5		Ser	Ser	Phe	Glr 10		L Asp	Су:	s Phe	Let	
	His	Val	. Arg	Lys 20		Val	. Ala	Asp	Gln 25		.Lev	2 G13	/ Asr	Ala 30		) Ph
10	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40		Ser	Leu	Arç	Gl <sub>3</sub>		Gl)	y Se:
	Thr	Leu 50	Gly	Leu	Asp	Ile	G1u 55		Ala	Thr	Arg	Ala 60		Lys	Glr	ılle
15	<b>Val</b> 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75		Leu	Lys	Met	Th:
20	Met	Gln	Ile	Pro	Gly 85	Leu	Phe	Gly	Ala	Ile 90	Ala	Gly	Phe	Ile	Glu 95	
	Gly	Trp	Thr	Gly 100	Met	Ile	Asp	Gly	Trp 105	Tyr	Gly	Tyr	His	His 110	Gln	Asn
25	Glu	Gln	Gly 115	Ser	Gly	Tyr	Ala	Ala 120	Asp	Gln	Lys	Ser	Thr 125	Gln	Asn	Ala
	Ile	Asn 130	Gly	Ile	Thr	Asn	Lys 135	Val	Asn	Ser	Val	Ile 140	Glu	Lys	Met	Asn
30	Ile 145	Gln	Phe	Thr	Ala	Val 150	Gly	Lys	Glu	Phe	Asn 155	Lys	Leu	Glu	Lys	Arg 160
35	Met	Glu	Asn	Leu	Asn 165	Lys	Lys	Val	Asp	Asp 170	Gly	Phe	Leu	Asp	Ile 175	Trp
	Thr	Tyr	Asn	Ala 180	Glu	Leu	Leu	Val	Leu 185	Leu	Glu	Asn	Glu	Arg 190	Thr	Leu
10	Asp	Phe	His 195	Asp	Ser	Asn	Val	Lys 200	Asn	Leu	Tyr	Glu	Lys 205	Val	Lys	Ser
	Gln	Leu 210	Lys	Asn	Asn	Ala	Lys 215	Glu	Ile	Gly	Asn	Gly 220	Cys	Phe	Glu	Phe
15	Tyr 225	His	Lys	Cys	Asp	Asn 230	Glu	Cys	Met	Glu	Ser 235	Val	Arg	Asn	Gly	Thr 240
50	Tyr	Asp	Tyr	Pro	Lys 245	Tyr	Ser	Glu	Glu	Ser 250	Lys	Leu	Asn	Arg	Glu 255	Lys
-	Val	Asp	Gly	Val 260	Lys	Leu	Glu	Ser	Met 265	Gly	Ile	Tyr	Gln	Ile 270	Leu	Ala

55

					,													
	.Ile	Tyr	Ser 275	Thr	Val	Ala	Ser	Ser 280	'Leu	Val	Leu	Leu	Val 285	Ser	Leu	Gly		
5	Ala	Ile 290	Ser	Phe	Trp	Met	Cys 295	Ser	Asn	Gly	Ser	Leu 300	Gln	Cys	Arg	Ile		
10	(2)	INF(	SE	QUEN	CE CI	HARA	CTER:	ISTI	CS:		,							
15			(1	A) L: B) T: C) S: O) T(	YPE : IRANI	nuc: DEDNI	leic ESS:	acidoul	d d	S								
		(ii)	MO	LECU	LE T	YPE:	DNA	(ge	nomi	c)							·	
20		(ix)	(2	ATURI A) Ni B) L	AME/I			471									·	
25		(xi)	SE(	QUENC	CE DI	ESCRI	IPTI(	วพ: :	SEQ :	ID N	D:33	:						
30	GTG Val 1	GGT Gly	AAA Lys	GAA Glu	TTC Phe 5	AAC Asn	AAA Lys	TTA Leu	GAA Glu	AAA Lys 10	AGG Arg	ATG Met	GAA Glu	AAT Asn	TTA Leu 15	AAT Asn	48	
35		AAA Lys															96	
		TTA Leu															144	
40		GTG Val 50															192	
45	GCC Ala 65	AAA Lys	GAA Glu	ATC Ile	GGA Gly	AAT Asn 70	GGA Gly	TGT Cys	TTT Phe	GAG Glu	TTC Phe 75	TAC Tyr	CAC His	AAG Lys	TGT Cys	GAC Asp 80	240	
50	AAT Asn	GAA Glu	TGC Cys	ATG Met	GAA Glu 85	AGT Ser	GTA Val	AGA Arg	AAT Asn	GGG Gly 90	ACT Thr	TAT Tyr	GAT Asp	TAT Tyr	CCC Pro 95	AAA Lys	288	
		TCA Ser															336	

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	Leu	GAA Glu	Y TC	r Met	GGG Gly	ATO	TA:	CAC CGlr 120	ı Ile	CT(	G GCG	ATC Ile	TAC Ty: 125	Se	A ACT	GTC Val
5	GCC Ala	Ser 130	Ser	A CTG	GTC Val	CTI Lev	TTC Lev	val	TCC Ser	CTC	GGG Gly	GCA Ala 140	Ile	AGT Sei	TTC Phe	TGG Trp
1.0	ATG Met 145	Cys	TCT Ser	TAA T	GGA Gly	TCI Ser 150	Leu	G CAG	TGC Cys	AGA	A ATA J Ile 155	Cys	ATC	TG#	1	
15	(2)	INF			ENCE	CHA NGTH	RACT	ERIS	TICS ino		ls					
20			•	(D MOLE	) TO	POLO TYP	GY: E: p		ar in							
25	Val 1										NO:		Glu	Asn	Leu 15	Asn
3Ó	Lys	Lys	Val	Asp 20	Asp	Gly	Phe	Leu	Asp 25	Ile	Trp	Thr	Tyr	Asn 30	Alá	Glu
	Leu	Leu	Val 35	Leu	Leu	Glu	Asn	Glu 40	Arg	Thr	Leu	Asp	Phe 45	His	Asp	Ser
<b>3</b> 5	Asn	Val 50	Lys	Asn	Leu	Tyr	Glu 55	Lys	Val	Lys	Ser	Gln 60	Leu	Lys	Asn	Asn
	Ala 65	Lys	Glu	Ile	Gly	Asn 70	Gly	Cys	Phe	Glu	Phe 75	Tyr	His	Lys	Cys	Asp 80
40	Asn	Glu	Суѕ	Met	Glu 85	Ser	Val	Arg	Asn	Gly 90	Thr	Tyr	Asp	Tyr	Pro 95	Lys
45	Tyr	Ser	Glu	Glu 100	Ser	Lys	Leu	Asn	Arg 105	Glu	Lys	Val	Asp	Gly 110	Val	Lys
	Leu	Glu	Ser 115	Met	Gly	Ile	Tyr	Gln 120	Ile	Leu	Ala		Tyr 125	Ser	Thr	Val
50	Ala	Ser 130	Ser	Leu	Val	Leu	Leu 135	Val	Ser	Leu	Gly	Ala 140	Ile	Ser	Phe	Trp
	Met 145	Cys	Ser	Asn	Gly	Ser 150	Leu	Gln	Суз	Arg	Ile 155	Cys	Ile			

	(2) INFORMATION FOR SEQ ID NO:35:	
5	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 47 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
10	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:	
15	CATGGATCAT ATGTTAACAG ATATCAAGGC CTGACTGACT GAGAGCT	. 4
	(2) INFORMATION FOR SEQ ID NO:36:	
20 25	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 39 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
25	(ii) MOLECULE TYPE: DNA (genomic)	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:  CTCAGTCAGT CAGGCCTTGA TATCTGTTAA CATATGATC	
	(2) INFORMATION FOR SEQ ID NO:37:	
35	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 29 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single	
40	(D) TOPOLOGY: unknown	
	(ii) MOLECULE TYPE: DNA (genomic)	·
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:	
	CATGGGCGCC CATATGGGCA TATTCGGCG	29

	(2) INFORMATION FOR SEQ ID NO:38:	
5	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 23 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
10	(ii) MOLECULE TYPE: DNA (genomic)	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:	
	CCGAATATGC CCATATGGGC GCC	23
15		
	(2) INFORMATION FOR SEQ ID NO:39:	
. 20	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 49 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
25	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:	
30	CATGGATCAT ATGTTAACAA GTACTCGATA TCAATGAGTG ACTGAAGCT	49
	(2) INFORMATION FOR SEQ ID NO:40:	
35	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 41 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
40	(ii) MOLECULE TYPE: DNA (genomic)	:
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	
45	TCAGTCACTC ATTGATATCG AGTACTTGTT AACATATGAT C	41

	(2) INFORMATION FOR SEQ ID NO:41:	
5	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 12 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
10	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	
	AATTCGTACC TA	12
15		
	(2) INFORMATION FOR SEQ ID NO:42:	
·20	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 12 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
25	(ii) MOLECULE TYPE: DNA (genomic)	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:42:	
30	GATCTAGGTA CG	12
	(2) INFORMATION FOR SEQ ID NO:43:	
35	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 54 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
40	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:	
45	ANACTGTTTG AAAAAACACG TCGTCAACTG CGTGAAAATG CTGACGACAT GGGC	54
	(2) INFORMATION FOR SEQ ID NO:44:	
50 55	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 52 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	

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	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:	
5	TGTGACAATG CTTGCATCGG TTCAATCCGT AATGGTACTT ATGACCATGA TG	52
	(2) INFORMATION FOR SEQ ID NO:45:	
10	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 20 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
15	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:	
20	GATCCCGGGT GACTGACTGA	20
25	(2) INFORMATION FOR SEQ ID NO:46:  (i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 20 base pairs	
30	(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown	
	(ii) MOLECULE TYPE: DNA (genomic)	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46: GATCTCAGTC AGTCACCCGG	20
	(2) INFORMATION FOR SEQ ID NO:47:	
40	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 20 base pairs  (B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single (D) TOPOLOGY: unknown	
45	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:	
50	TAAGGAGGAT ATAACATATG	20

ı	(2) INFORMATION FOR SEQ ID NO:48:	
5	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 28 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
10	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:	
	GATCCATATG TTATATCCTC CTTAAGGT	28
15	(2) INFORMATION FOR SEQ ID NO:49:	
20	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 21 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
25	(11) MOLECULE TYPE: DNA (genomic)	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:49:	
	GCATCGCCAT GAGTCACGAC G	21
30	$oldsymbol{\cdot}$	
	(2) INFORMATION FOR SEQ ID NO:50:	
35	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 30 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
40	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:	
4.5	GGAGGATGGG AAGGACTCAT TGCAGGTTGG	30
45		

	(2) INFORMATION FOR SEQ ID NO:51:	
5	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 41 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
	(ii) MOLECULE TYPE: DNA (genomic)	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:	
	CTCTGCTGTA GAAATCGGTA ACGGTTGCTT TGAAACCAAA C	4 ]
15		
	(2) INFORMATION FOR SEQ ID NO:52:	
20	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 47 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
25	(ii) MOLECULE TYPE: DNA (genomic)	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:52:	
30	GGTTTCTTGG AAGGTGGTTG GGAAGGTCTC ATTGCAGGTT GGCACGG	47
	(2) INFORMATION FOR SEQ ID NO:53:	
35	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 48 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
40	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:	
45	GCTTTCCAAC GAAGGTATCA ATCAACAGTG AAGACGAGCA TCTCTTGG	. 48
	(2) INFORMATION FOR SEQ ID NO:54:	
50	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 7616 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: double</li> <li>(D) TOPOLOGY: unknown</li> </ul>	
55	(ii) MOLECULE TYPE: DNA (genomic)	

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#### (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1879..2790

5

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

	AATTCTCATG	TTTGACAGCT	TATCATCGAT	AAGCTTCAGT	TGAAGATATT	AAGAACAGCC	60
10	TCGCAGATGA.	CGAATCATTG	GGATTCCCAT	CTTTTTTGTT	TGTTGAAGGC	GACACCATTG	120
	GTTTTGCCAG	AACTGTTTTC	GGGCCGACCA	CATCCGATCT	GACAGATTTT	TTAATCGGGA	180
15	AAGGAATGTC	ATTAAGCAGT	GGAGAGCGCG	TTCAGATAGA	GCCACTGATG	AGGGGAACCA	240
13	CCAAAGACGA	TGTTATGCAT	ATGCATTTCA	TCGGCCGAAC	AACGGTGAAG	GTAGAAGCCA	300
	AGCTACCTGT	ATTTGGCGAT	ATATTAAAGG	TCTTAGGGGC	AACAGATATT	GAAGGGGAGC	360
20	TTTTTGACTC	ATTGGATATA	GTCATTAAGC	CAAAATTTAA	AAGGGATATA	AAAAAGGTTG	420
	CCAAGGATAT	TATTTTTAAC	CCGTCACCTC	AATTTTCAGA	CATTAGCCTG	CGGGCAAAAG	480
25	ATGAGGCCGG	AGATATTTTA	ACAGAACATT	ATCTATCAGA	AAAAGGCCAT	CTCTCAGCGC	540
23	CTCTGAACAA	GGTCACCAAT	GCTGAGATAG	CTGAAGAGAT	GGCATATTGC	TACGCAAGAA	600
	TGAAAAGTGA	TATACTGGAA	TGTTTTAAAA	GGCAGGTGGG	CAAAGTTAAG	GATTAATTAT	660
30	CAGGAGTAAT	TATGCGGAAC	AGAATCATGC	CTGGTGTTTA	CATAGTAATA	ATTCCTTACG	720
	TTATCGTAAG	CATTTGCTAT	CTCCTTTTCC	GCCACTACAT	TCCTGGTGTT	TCTTTTTCAG	780
35	CTĊATAGAGA	TGGTCTTGGG	GCGACATTGT	CATCATATGC	AGGAACCATG	ATTGCAATCC	840
	TGATTGCTGC	CTTGACGTTT	CTAATCGGAA	GCAGAACGCG	CCGACTGGCC	AAGATTAGAG	900
	AGTATGGGTA	TATGACATCG	GTAGTTATTG	TCTATGCCCT	TAGTTTTGTT	GAGCTTGGAG	960
40	CTTTGTTTTT	CTGCGGGTTA	TTGCTTCTTT	CCAGCATAAG	CGGCTACATG	ATACCCACTA	1020
	TCGCCATCGG	CATTGCCTCT	GCATCGTTCA	TTCATATATG	CATCCTTGTT	TTCCAACTAT	1080
45	ATAATTTGAC	CAGAGAACAA	GAATAACCCG	GCCTCAGCGC	CGGGTTTTCT	TTGCCTCACG	1140
	ATCGCCCCCA	AAACACATAA	CCAATTGTAT	TTATTGAAAA	ATAAATAGAT	ACAACTCACT	1200
	AAACATAGCA	ATTCAGATCT	CTCACCTACC	AAACAATGCC	CCCCTGCAAA	AAATAAATTC	1260
50	АТАТАААААА	CATACAGATA	ACCATCTGCG	GTGATAAATT	ATCTCTGGCG	GTGTTGACAT	1320
	AAATACCACT	GGCGGTGATA	CTGAGCACAT	CAGCAGGACG	CACTGACCAC	CATGAAGGTG	1380
<b>55</b> <sub>.</sub>	ACGCTCTTAA	AAATTAAGCC	CTGAAGAAGG	GCAGCATTCA	AAGCAGAAGG	CTTTGGGGTG	1440

	TGTGATACGA AACGAAGCAT TGGCCGTAAG TGCGATTCCG GATTAGCTGC CAATGTGCCA	1500
	ATCGCGGGG GTTTTCGTTC AGGACTACAA CTGCCACACA CCACCAAAGC TAACTGACAG	1560
5	GAGAATCCAG ATGGATGCAC AAACACGCCG CCGCGAACGT CGCGCAGAGA AACAGGCTCA	1620
	ATGGAAAGCA GCAAATCCCC TGTTGGTTGG GGTAAGCGCA AAACCAGTTC CGAAAGATTT	1680
10	TTTTAACTAT AAACGCTGAT GGAAGCGTTT ATGCGGAAGA GGTAAAGCCC TTCCCGAGTA	1740
10	ACAAAAAAC AACAGCATAA ATAACCCCGC TCTTACACAT TCCAGCCCTG AAAAAAGGGCA	1800
	TCAAATTAAA CCACACCTAT GGTGTATGCA TTTATTTGCA TACATTCAAT CAATTGTTAT	1860
15	CTAAGGAAAT ACTTACAT ATG GAT CCA AAC ACT GTG TCA AGC TTT CAG GTA  Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val  1 5 10	1911
20	GAT TGC TTT CTT TGG CAT GTC CGC AAA CGA GTT GCA GAC CAA GAA CTA Asp Cys Phe Leu Trp His Val Arg Lys Arg Val Ala Asp Gln Glu Leu 15 20 25	1959
25	GGT GAT GCC CCA TTC CTT GAT CGG CTT CGC CGA GAT CAG AAA TCC CTA Gly Asp Ala Pro Phe Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu 30 35 40	2007
30	AGA GGA AGG GGC AGC ACC CTC GGT CTG GAC ATC GAG ACA GCC ACA CGT Arg Gly Arg Gly Ser Thr Leu Gly Leu Asp Ile Glu Thr Ala Thr Arg 45 50 55	2055
,	GCT GGA AAG CAG ATA GTG GAG CGG ATT CTG AAA GAA GAA TCC GAT GAG Ala Gly Lys Gln Ile Val Glu Arg Ile Leu Lys Glu Glu Ser Asp Glu 60 65 70 75	2103
35	GCA CTT AAA ATG ACC ATG GGT TTC TTC GGA GCT ATT GCT GGT TTC TTG Ala Leu Lys Met Thr Met Gly Phe Phe Gly Ala Ile Ala Gly Phe Leu 80 85 90	2151
40	GAA GGT GGT TGG GAA GGT CTC ATT GCA GGT TGG CAC GGA TAC ACA TCT Glu Gly Gly Trp Glu Gly Leu Ile Ala Gly Trp His Gly Tyr Thr Ser 95 100 105	2199
45	CAT GGA GCA CAT GGA GTG GCA GTG GCA GCA GAC CTT AAG AGT ACA CAA His Gly Ala His Gly Val Ala Val Ala Ala Asp Leu Lys Ser Thr Gln 110 115 120	2247
50	GAA GCT ATA AAC AAG ATA ACA AAA AAT CTC AAC TAT TTA AGT GAG CTA Glu Ala Ile Asn Lys Ile Thr Lys Asn Leu Asn Tyr Leu Ser Glu Leu 125 130 135	2295
	GAA GTA AAA AAC CTT CAA AGA CTA AGC GGA GCA ATG AAT GAG CTT CAC Glu Val Lys Asn Leu Gln Arg Leu Ser Gly Ala Met Asn Glu Leu His 140 155 150	2343

55

	GAC Asp	GAA Glu	ATA Ile	CTC Leu	GAG Glu 160	CTA Leu	GAC Asp	GAA Glu	AAA Lys	GTG Val 165	GAT Asp	GAT Asp	CTA Leu	AGA Arg	GCT Ala 170	GAT Asp	2391
5					CAA Gln												2439
10					GAA Glu												2487
15					GGC Gly												2535
20	_				AAA Lys											GCT Ala . 235	2583
	GGC GGC				GCA Ala 240		_										2631
25					GCA Ala											ACT. Thr	2679
30					TAC Tyr												2727
35					TTC Phe												2775
40			ATC Ile		CTG Leu	TGA	GGA(	GAT S	raag(	CCT	ST G1	rttt(	CCTT	r act	rgta(	STGC	2830
10	TCA'	rttg(	CTT (	GTCA	CCAT	TA C	AAAG	AAAC	G TT	ATTG/	AAAA	ATG	CTCT:	rgt :	ract!	ACTGAA	2890
	TTC	TAGA	ATC (	GATA	AGCT:	IC G	ACCG	ATGC	C CT	rgag?	AGCC	TTC	AACC	CAG :	CAG	CTCCTT	2950
45	CCG	GTGG	GCG (	CGGG	GCAT	GA C	PATC	GTCG	C CGC	CACT	ratg	ACT	GTCT:	PCT :	TAT	CATGCA	3010
	ACT	CGTA	GGA (	CAGG'	rgcc	GG C	AGCG	CTCT	G GG	rcat:	TTTC	GGC	GAGG	ACC (	CTT	CGCTG	3070
50	GAG	CGCG	ACG 2	ATGA'	rcgg	CC TO	GTCG	CTTG	C GG	PATTO	CGGA	ATC:	TTGC	ACG (	CCT	CGCTCA	3130
	AGC	CTTC	GTC :	ACTG	GTCC	CG C	CACC	AAAC	G TT	rcgg	CGAG	AAG	CAGG	CCA !	TAT	CGCCGG	3190
	CAT	GGCG	GCC (	GACG	CGCT	GG G(	CTAC	GTCT'	r GC	rĢGC(	STTC	GTC	CAGT	TAP	GACC	CAGAA	3250
55	CTC	CATC'	TGG .	ATTT(	GTTC	AG A	ACGC'	rcgg:	r TG	CCCC	CGGG	CGT	FTTT:	rat :	rggto	GAGAAT	3310

	CGCAGCAACT	TGTCGCGCCA	ATCGAGCCAT	GTCGTCGTCA	ACGACCCCC	ATTCAAGAAC	3370
5	AGCAAGCAGC	ATTGAGAACT	TTGGAATCCA	GTCCCTCTTC	CACCTGCTGA	GACGCGAGGC	3430
	TGGATGGCCT	TCCCCATTAT	GATTCTTCTC	GCTTCCGGCG	GCATCGGGAT	GCCCGCGTTG	3490
	CAGGCCATGC	TGTCCAGGCA	GGTAGATGAC	GACCATCAGG	GACAGCTTCA	AGGATCGCTC	3550
10	GCGGCTCTTA	CCAGCCTAAC	TTCGATCACT	GGACCGCTGA	TCGTCACGGC	GATTTATGCC	3610
•	GCCTCGGCGA	GCACATGGAA	CGGGTTGGCA	TGGATTGTAG	GCGCCGCCCT	ATACCTTGTC	3670
15·	TGCCTCCCCG	CGTTGCGTCG	CGGTGCATGG	AGCCGGGCCA	CCTCGACCTG	AATGGAAGCC	3730
	GGCGGCACCT	CGCTAACGGA	TTCACCACTC	CAAGAATTGG	AGCCAATCAA	TTCTTGCGGA	3790
	GAACTGTGAA	TGCGCAAACC	AACCCTTGGC	AGAACATATC	CATCGCGTCC	GCCATCTCCA	3850
20	GCAGCCGCAC	GCGGCGCATC	TCGGGCAGCG	TTGGGTCCTG	GCCACGGGTG	CGCATGATCG	3910
	TGCTCCTGTC	GTTGAGGACC	CGGCTAGGCT	GGCGGGGTTG	CCTTACTGGT	TAGCAGAATG	3970
25	AATCACCGAT	ACGCGAGCGA	ACGTGAAGCG	ACTGCTGCTG	CAAAACGTCT	GCGACCTGAG	4030
	CAACAACATG	AATGGTCTTC	GGTTTCCGTG	TTTCGTAAAG	TCTGGAAACG	CGGAAGTCAG	4090
	CGCCCTGCAC	CATTATGTTC	CGGATCTGCA	TCGCAGGATG	CTGCTGGCTA	CCCTGTGGAA	4150
30	CACCTACATC	TGTATTAACG	AAGCGCTGGC	ATTGACCCTG	AGTGATTTTT	CTCTGGTCCC	4210
	GCCGCATCCA	TACCGCCAGT	TGTTTACCCT	CACAACGTTC	CAGTAACCGG	GCATGTTCAT	4270
35	CATCAGTAAC	CCGTATCGTG	AGCATCCTCT	CTCGTTTCAT	CGGTATCATT	ACCCCCATGA	4330
	ACAGAAATTC	CCCCTTACAC	GGAGGCATCA	AGTGACCAAA	CAGGAAAAA	CCGCCCTTAA	4390
	CATGGCCCGC	TTTATCAGAA	GCCAGACATT	AACGCTTCTG	GAGAAACTCA	ACGAGCTGGA	4450
40	CGCGGATGAA	CAGGCAGACA	TCTGTGAATC	GCTTCACGAC	CACGCTGATG	AGCTTTACCG	4510
	CAGCTGCCTC	GCGCGTTTCG	GTGATGACGG	TGAAAACCTC	TGACACATGC	AGCTCCCGGA	4570
45	GACGGTCACA	GCTTGTCTGT	AAGCGGATGC	CGGGAGCAGA	CAAGCCCGTC	AGGGCGCGTC	4630
	AGCGGGTGTT	GGCGGGTGTC	GGGGCGCAGC	CATGACCCAG	TCACGTAGCG	ATAGCGGAGT	4690
	GTATACTGGC	TTAACTATGC	GGCATCAGAG	CAGATTGTAC	TGAGAGTGCA	CCATATGCGG	4750
50	TGTGAAATAC	CGCACAGATG	CGTAAGGAGA	AAATACCGCA	TCAGGCGCTC	TTCCGCTTCC	4810
	TCGCTCACTG	ACTCGCTGCG	CTCGGTCGTT	CGGCTGCGGC	GAGCGGTATC	AGCTCACTCA	4870
55	AAGGCGGTAA	TACGGTTATC	CACAGAATCA	GGGGATAACG	CAGGAAAGAA	CATGTGAGCA	4930

	AAAGGCCAGC	AAAAGGCCAG	GAACCGTAAA	AAGGCCGCGT	TGCTGGCGTT	TTTCCATAGG	4990
	CTCCGCCCCC	CTGACGAGCA	TCACAAAAAT	CGACGCTCAA	GTCAGAGGTG	GCGAAACCCG	5050
5	ACAGGACTAT	AAAGATACCA	GGCGTTTCCC	CCTGGAAGCT	CCCTCGTGCG	CTCTCCTGTT	5110
	CCGACCCTGC	CGCTTACCGG	ATACCTGTCC	GCCTTTCTCC	CTTCGGGAAG	CGTGGCGCTT	5170
10	TCTCAATGCT	CACGCTGTAG	GTATCTCAGT	TCGGTGTAGG	TCGTTCGCTC	CAAGCTGGGC	5230
10	TGTGTGCACG	AACCCCCGT	TCAGCCCGAC	CGCTGCGCCT	TATCCGGTAA	CTATCGTCTT	5290
	GAGTCCAACC	CGGTAAGACA	CGACTTATCG	CCACTGGCAG	CAGCCACTGG	TAACAGGATT	5350
15	AGCAGAGCGA	GGTATGTAGG	CGGTGCTACA	GAGTTCTTGA	AGTGGTGGCC	TAACTACGGC	5410
	TACACTAGAA	GGACAGTATT	TGGTATCTGC	GCTCTGCTGA	AGCCAGTTAC	CTTCGGAAAA	5470
20	AGAGTTGGTA	GCTCTTGATC	CGGCAAACAA	ACCACCGCTG	GTAGCGGTGG	TTTTTTTGTT	5530
20	TGCAAGCAGC	AGATTACGCG	CAGAAAAAA	GGATCTCAAG	AAGATCCTTT	GATCTTTTCT	5590
	ACGGGGTCTG	ACGCTCAGTG	GAACGAAAAC	TCACGTTAAG	GGATTTTGGT	CATGAGATTA	5650
25	TCAAAAAGGA	TCTTCACCTA	GATCCTTTTA	TAAAAATTAA	GAAGTTTTAA	ATCAATCTAA	5710
	AGTATATATG	AGTAAACTTG	GTCTGACAGT	TACCAATGCT	TAATCAGTGA	GGCACCTATC	5770
30	TCAGCGATCT	GTCTATTTCG	TTCATCCATA	GTTGCCTGAC	TCCCCGTCGT	GTAGATAACT	5830
	ACGATACGGG	AGGGCTTACC	ATCTGGCCCC	AGTGCTGCAA	TGATACCGCG	AGACCCACGC	5890
	TCACCGGCTC	CAGATTTATC	AGCAATAAAC	CAGCCAGCCG	GAAGGGCCGA	GCGCAGAAGT	5950
35	GGTCCTGCAA	CTTTATCCGC	CTCCATCCAG	TCTATTAATT	GTTGCCGGGA	AGCTAGAGTA	6010
	AGTAGTTCGC	CAGTTAATAG	TTTGCGCAAC	GTTGTTGCCA	TTGCTGCAGG	CATCGTGGTG	6070
40	TCACGCTCGT	CGTTTGGTAT	GGCTTCATTC	AGCTCCGGTT	CCCAACGATC	AAGGCGAGTT	6130
40	ACATGATCCC	CCATGTTGTG	CAAAAAAGCG	GTTAGCTCCT	TCGGTCCTCC	GATCGTTGTC	6190
	AGAAGTAAGT	TGGCCGCAGT	GTTATCACTC	ATGGTTATGG	CAGCACTGCA	TAATTCTCTT	6250
45	ACTGTCATGC	CATCCGTAAG	ATGCTTTTCT	GTGACTGGTG	AGTAGCTTCA	CGCTGCCGCA	6310
	AGCACTCAGG	GCGCAAGGGC	TGCTAAAGGA	AGCGGAACAC	GTAGAAAGCC	AGTCCGCAGA	6370
~ ^	AACGGTGCTG	ACCCCGGATG	AATGTCAGCT	ACTGGGCTAT	CTGGACAAGG	GAAAACGCAA	6430
50	GCGCAAAGAG	AAAGCAGGTA	GCTTGCAGTG	GGCTTACATG	GCGATAGCTA	GACTGGGCGG	6490
•	TTTTATGGAC	AGCAAGCGAA	CCGGAATTGC	CAGCTGGGGC	GCCCTCTGGT	AAGGTTGGGA	6550
55	AGCCCTGCAA	AGTAAACTGG	ATGGCTTTCT	TGCCGCCAAG	GATCTGATGG	CGCAGGGGAT	6610

	CAAGATCTGA	TCAAGAGACA	GGATGAGGAT	CGTTTCGCAT	GATTGAACAA	GATGGATTGC	6670
5 ·	ACGCAGGTTC	TCCGGCCGCT	TGGGTGGAGA	GGCTATTCGG	CTATGACTGG	GCACAACAGA	6730
5	CAATCGGCTG	CTCTGATGCC	GCCGTGTTCC	GGCTGTCAGC	GCAGGGGCGC	CCGGTTCTTT	6790
	TTGTCAAGAC	CGACCTGTCC	GGTGCCCTGA	ATGAACTGCA	GGACGAGGCA	GCGCGGCTAT	6850
10	CGTGGCTGGC	CACGACGGGC	GTTCCTTGCG	CAGCTGTGCT	CGACGTTGTC	ACTGAAGCGG	6910
	GAAGGGACTG	GCTGCTATTG	GGCGAAGTGC	CGGGGCAGGA	TCTCCTGTCA	TCTCACCTTG	6970
15	CTCCTGCCGA	GAAAGTATCC	ATCATGGCTG	ATGCAATGCG	GCGGCTGCAT	ACGCTTGATC	7030
13	CGGCTACCTG	CCCATTCGAC	CACCAAGCGA	AACATCGCAT	CGAGCGAGCA	CGTACTCGGA	7090
	TGGAAGCCGG	TCTTGTCGAT	CAGGATGATC	TGGACGAAGA	GCATCAGGG	CTCGCGCCAG	7150
20	CCGAACTGTT	CGCCAGGCTC	AAGGCGCGCA	TGCCCGACGG	CGAGGATCTC	GTCGTGACTC	7210
	ATGGCGATGC	CTGCTTGCCG	AATATCATGG	TGGAAAATGG	CCGCTTTTCT	GGATTCATCG	7270
25	ACTGTGGCCG	GCTGGGTGTG	GCGGACCGCT	ATCAGGACAT	AGCGTTGGCT	ACCCGTGATA	7330
2.0	TTGCTGAAGA	GCTTGGCGGC	GAATGGGCTG	ACCGCTTCCT	CGTGCTTTAC	GGTATCGCCG	7390
	CTCCCGATTC	GCAGCGCATC	GCCTTCTATC	GCCTTCTTGA	CGAGTTCTTC	TGAGCGGGAC	7450
30	TCTGGGGTTC	GAAATGACCG	ACCAAGCGAC	GCCCAACCTG	CCATCACGAG	ATTTCGATTC	7510
	CACCGCCGCC	TTCTATGAAA	GGTTGGGCTT	CGGAATCGTT	TTCCGGGACG	CCGGCTGGAT	7570
35	GATCCTCCAG	CGCGGGGATC	TCATGCTGGA	GTTCTTCGCC	CACCCC		7616

#### (2) INFORMATION FOR SEQ ID NO:55:

40 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 304 amino acids

(B) TYPE: amino acid(D) TOPOLOGY: linear

45 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp 50 1 5 10 15

His Val Arg Lys Arg Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe 20 25 30

	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser
5	Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile
•	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Thr 80
10	Met	Gly	Phe	Phe	Gly 85	Ala	Ile	Ala	Gly	Phe 90	Leu	Glu	Gly	Gly	Trp 95	Glu
15	Gly	Leu	Ile	Ala 100	Gly	Trp	His	Gly	Tyr 105	Thr	Ser	His	Gly	Ala 110	His	Gly
	Val	Ala	Val 115	Ala	Ala	Asp	Leu	Lys 120	Ser	Thr	Gln	Glu	Ala 125	Ile	Asn	Lys
20	Ile	Thr 130	Lys	Asn	Leu	Asn	Tyr 135	Leu	Ser	Glu	Leu	Glu 140	Val	Lys	Asn	Leu
	Gln 145	Arg	Leu	Ser	Gly	Ala 150	Met	Asn	Glu	Leu	His 155	Asp	Glu	Ile	Leu	Glu 160
25	Leu	Asp	Glu	Lys	Val 165	Asp	Asp	Leu	Arg	Ala 170	Asp	Thr	Ile	Ser	Ser 175	Gln
30	Ile	Glu	Leu	Ala 180	Val	Leu	Leu	Ser	Asn 185	Glu	Gly	Ile	Ile	Asn 190	Ser	Glu
٠	,		195					200					205	Met		
35		210					215					220		Lys		
	225	٠.			_	230					235			Phe		240
40	_				245					250				Thr	255	
45	Ser	Leu	Asn	Asp 260	_	Gly	Leu	Asp	Asn 265		Thr	Ile	Leu	Leu 270	Tyr	Tyr
	Ser	Thr	Ala 275		Ser	Ser	Leu	Ala 280		Thr	Leu	Met	Ile 285	Ala	Ile	Phe
50	Ile	Val	_	Met	Val	Ser	Arg		Asn	Val	Ser	Cys 300		Ile	Cys	Leu

(2) INFORMATION FOR SEQ ID NO:56:

#### (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 915 base pairs 5 (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: unknown (ii) MOLECULE TYPE: DNA (genomic) 10 (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1..912 15 (x1) SEQUENCE DESCRIPTION: SEQ ID NO:56: ATG GAT CCA AAC ACT GTG TCA AGC TTT CAG GTA GAT TGC TTT CTT TGG 48 Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp 20 CAT GTC CGC AAA CGA GTT GCA GAC CAA GAA CTA GGT GAT GCC CCA TTC 96 His Val Arg Lys Arg Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe 25 25 CTT GAT CGG CTT CGC CGA GAT CAG AAA TCC CTA AGA GGA AGG GGC AGC 144 Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu Arg Gly Arg Gly Ser 35 40 ACC CTC GGT CTG GAC ATC GAG ACA GCC ACA CGT GCT GGA AAG CAG ATA 192 Thr Leu Gly Leu Asp Ile Glu Thr Ala Thr Arg Ala Gly Lys Gln Ile 30 50 55 GTG GAG CGG ATT CTG AAA GAA GAA TCC GAT GAG GCA CTT AAA ATG ACC 240 Val Glu Arg Ile Leu Lys Glu Glu Ser Asp Glu Ala Leu Lys Met Thr 35 65 70 ATG GGT TTC TTC GGA GCT ATT GCT GGT TTC TTG GAA GGA GGA TGG GAA 288 Met Gly Phe Phe Gly Ala Ile Ala Gly Phe Leu Glu Gly Gly Trp Glu 85 . 90 95 40 GGA ATG ATT GCA GGT TGG CAC GGA TAC ACA TCT CAT GGA GCA CAT GGA 336 Gly Met Ile Ala Gly Trp His Gly Tyr Thr Ser His Gly Ala His Gly 100 45 GTG GCA GTG GCA GCA GAC CTT AAG AGT ACA CAA GAA GCT ATA AAC AAG 384 Val Ala Val Ala Ala Asp Leu Lys Ser Thr Gln Glu Ala Ile Asn Lys 115 120 ATA ACA AAA AAT CTC AAC TAT TTA AGT GAG CTA GAA GTA AAA AAC CTT 432 50 Ile Thr Lys Asn Leu Asn Tyr Leu Ser Glu Leu Glu Val Lys Asn Leu 130

	CAA Gln 145	AGA Arg	CTA Leu	AGC Ser	GGA Gly	GCA Ala 150	ATG Met	AAT Asn	GAG Glu	CTT Leu	CAC His 155	GAC Asp	GAA Glu	ATA Ile	CTC Leu	GAG Glu 160	480
5			GAA Glu														528
10			CTT Leu													GAA Glu	576
15			CAT His 195														624
20			GCT Ala													AAA Lys	672
. – -			CAG Gln														720
25			TTT Phe													GCA . Ala	768
30			AAT Asn														816
35			GCT Ala 275														864
40			TAC														912
	TGA															•	915

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(2) INFORMATION FOR SEQ ID NO:57:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 304 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

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# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

5	Met 1	Asp	Pro	Asn	Thr 5	Val	Ser	Ser	Phe	Gln 10	Val	Asp	Cys	Phe	Leu 15	Trp
J	His	Val	Arg	Lys 20	Arg	Val	Ala	Asp	Gln 25	Glu	Leu	Gly	Asp	Ala 30	Pro	Phe
10	Leu	Asp	Arg 35	Leu	Arg	Arg	Asp	Gln 40	Lys	Ser	Leu	Arg	Gly 45	Arg	Gly	Ser
	Thr	Leu 50	Gly	Leu	Asp	Ile	Glu 55	Thr	Ala	Thr	Arg	Ala 60	Gly	Lys	Gln	Ile
15	Val 65	Glu	Arg	Ile	Leu	Lys 70	Glu	Glu	Ser	Asp	Glu 75	Ala	Leu	Lys	Met	Thr 80
20	Met	Gly	Phe	Phe	Gly 85	Ala	Ile	Ala	Gly	Phe 90	Leu	Glu	Gly	Gly	Trp 95	Glu
	Gly	Met	Ile	Ala 100	Gly	Trp	His	Gly	Tyr 105	Thr	Ser	His	Gly	Ala 110	His	Gly
25	Val	Ala	Val 115	Ala	Ala	Asp	Leu	Lys 120	Ser	Thr	Gln	Glu	Ala 125	Ile	Asn	Lys
	Ile	Thr 130	Lys	Asn	Leu	Asn	Tyr 135	Leu	Ser	Glu	Leu	Glu 140	Val	Lys	Asn	Leu
30	Gln 145	Arg	Leu	Ser	Gly	Ala 150	Met	Asn	Glu	Leu	His 155	Asp	Glu	Ile	Leu	Glu 160
35	Leu	Asp	Glu	Lys	Val 165	Asp	Asp	Leu	Arg	Ala 170	Asp	Thr	Ile	Ser	Ser 175	Gln
	Ile	Glu	Leu	Ala 180	Val	Leu	Leu	Ser	Asn 185	Glu	Gly	Ile	Ile	Asn 190	Ser	Glu
40	Asp	Glu	His 195	Leu	Leu	Ala	Leu	Glu 200	Arg	Lys	Leu	Lys	Lys 205	Met	Leu	Gly
	Pro	Ser 210	Ala	Val	Glu	Ile		Asn			Phe	Glu 220	Thr	Lys	His	Lys
45	Cys 225	Asn	Gln	Thr	Суз	Leu 230	Asp	Arg	Ile	Ala	Ala 235	Gly	Thr	Phe	Asn	Ala 240
50	Glý	Asp	Phe	Ser	Leu 245	Pro	Thr	Phe	Asp	Ser 250	Leu	Asn	Ile	Thr	Ala 255	Ala
	Ser	Leu	Asn	Asp 260	Asp	Gly	Leu	Asp	Asn 265	His	Thr	Ile	Leu	Leu 270	Tyr	Tyr
55	Ser	Thr	Ala 275	Ala	Ser	Ser	Leu	Ala 280	Val	Thr	Leu	Met	Ile 285	Ala	Ile	Phe

Ile Val Tyr Met Val Ser Arg Asp Asn Val Ser Cys Ser Ile Cys Leu 290 295 300

(2) INFORMATION FOR SEQ ID NO:58:

10 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 918 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: unknown

15

5

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

20 ATGGATCCAA ACACTGTGTC AAGCTTTCAG GTAGATTGCT TTCTTTGGCA TGTCCGCAAA 60 CGAGTTGCAG ACCAAGAACT AGGTGATGCC CCATTCCTTG ATCGGCTTCG CCGAGATCAG 120 ANATCCCTAN GAGGANGGG CAGCACTCTT GGTCTGGACA TCGAGACAGC CACACGTGCT 180 25 GGAAAGCAGA TAGTGGAGCG GATTCTGAAA GAAGAATCCG ATGAGGCACT TAAAATGACC 240 ATGGGCGCCC ATATGGGCAT ATTCGGCGCA ATAGCAGGTT TCATAGAAAA TGGTTGGGAG 300 30 GGAATGATAG ACGGTTGGTA CGGTTTCAGG CATCAAAATT CTGAGGGCAC AGGACAAGCA 360 GCAGATCTTA AAAGCACTCA AGCAGCCATC GACCAAATCA ATGGGAAACT GAATAGGGTA 420 ATCGAGAAGA CGAACGAGAA ATTCCATCAA ATCGAAAAGG AATTCTCAGA AGTAGAAGGG 480 35 AGAATTCAGG ACCTCGAGAA ATACGTTGAA GACACTAAAA TAGATCTCTG GTCTTACAAT 540 GCGGAGCTTC TTGTCGCTCT GGAGAACCAA CATACAATTG ATCTGACTGA CTCGGAAATG 600 40 AACAAACTGT TTGAAAAAAC ACGTCGTCAA CTGCGTGAAA ATGCTGAGGA CATGGGCAAT 660 GGTTGCTTCA AAATATACCA CAAATGTGAC AATGCTTGCA TAGGGTCAAT CAGAAATGGG 720 ACTTATGACC ATGATGTATA CAGAGACGAA GCATTAAACA ACCGGTTTCA GATCAAAGGT 780 45 GTTGAACTGA AGTCAGGATA CAAAGACTGG ATCCTGTGGA TTTCCTTTGC CATATCATGC 840 TTTTTGCTTT GTGTTGTTTT GCTGGGGTTC ATCATGTGGG CCTGCCAAAA AGGCAACATT 900 50 AGGTGCAACA TTTGCATT 918

	(2)		0.0.2			. 520	, 10	.,	, , ,							
5			(i)	(A	JENCE () LE () TY () TO	NGTH PE:	l: 22 amin	1 am	ino id		ls					
10					CULE											
		(	Xì)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	59:				
15	Gly 1		Phe	Gly	Ala 5		Ala	Gly	Phe	Ile 10		Asn	Gly	Trp	Glu 15	
	Met	Ile	Asp	Gly 20	Trp	Tyr	Gly	Phe	Arg 25		Gln	Asn	Ser	Glu 30		Th
20	Gly	Gln	Ala 35	Ala	Asp	Leu	Lys	Ser 40	Thr	Gln	Ala	Ala	Ile 45		Gln	Il
25	Asn	Gly 50	Lys	Leu	Asn	Arg	Val 55	Ile	Glu	Lys	Thr	Asn 60		Lys	Phe	Hi.
	Gln 65	Ile	Glu	Lys	Glu	Phe 70	Ser	Glu	Val	Glu	Gly 75		Ile	Gln	Asp	Let
30	Glu	Lys	Tyr	Val	G1ນ 85	Asp	Thr	Lys	Ile	Asp 90	Leu	Trp	Ser	Tyr	Asn 95	Ala
	Glu	Leu	Leu	Val 100	Ala	Leu	Glu	Asn	Gln 105	His	Thr	Ile	Asp	Leu 110	Thr	Asp
35	Ser	Glu	Met 115	Asn	Lys	Leu	Phe	Glu 120	Lys	Thr	Arg	Arg	Gln 125	Leu	Arg	Glu
40	Asn	Ala 130	Glu	Asp	Met	Gly	Asn 135	Gly	Суз	Phe	Lys	Ile 140	Tyr	His	Lys	Cys
	Asp 145	Asn	Ala	Cys	Ile	Gly 150	Ser	Ile	Arg	Asn	Gly 155	Thr	Tyr	Asp	His	Asp
45	Val	Tyr	Arg	Asp	Glu 165	Ala	Leu	Asn	Asn	Arg 170	Phe	Gln	Ile	Lys	Gly 175	Val
	Glu	Leu	Lys	Ser 180	Gly	Tyr	Lys	Asp	Trp 185	Ile	Leu	Trp	Ile	Ser 190		Ala
50	Ile	Ser	Cys 195	Phe	Leu	Leu	Cys	Val 200	Val	Leu	Leu	Gly	Phe 205	Ile	Met	Trp
	Ala	Cys 210	Gln	Lys	Gly	Asn	Ile 215	Arg	Cys	Asņ	Ile	Cys 220	Ile			

5	(2)	INF	ORMA	TION	FOR	SEQ	ID :	NO:6	0:							
			(i)	(A)	) LEI	NGTH PE:	: 22 amin	ERIS 1 am o ac line	ino : id		S					
10		€.	ii) I	MOLE	CULE	TYP	E: p	rote	in							
		(:	xi)	SEQUI	ENCE	DES	CRIP'	TION	: SE	Q ID	NO:	60:				
15	Gly 1	Ile	Phe	Gly	Ala 5	Ile	Ala	Gly	Phe	Ile 10	Glu	Asn	Gly	Trp	Glu 15	Gly
20	Met	Ile	Asp	Gly 20	Trp	Tyr	Gly	Phe	Arg 25	His	Gln	Asn	Ser	Glu 30	Gly	Thi
20	Gly	Gln	Ala 35	Ala	Asp	Leu	Lys	Ser 40	Thr	Gln	Ala	Ala	Ile 45	Asp	Gln	Ile
25	Asn	Gly 50	Lys	Leu	Asn	Arg	Val 55	Ile	Glu	Lys	Thr	Asn 60	Glu	Lys	Phe	His
	Gln 65	Ile	Glu	Lys	Glu	Phe 70	Ser	Glu	Val	Glu	Gly 75	Arg	Ile	Gln	Asp	Leu 80
30	Glu	Lys	Tyr	Val	Glu 85	Asp	Thr	Lys	Ile	Asp 90	Leu	Trp	Ser	Tyr	Asn 95	Ala
35	Glu	Leu	Leu	Val 100	Ala	Leu	Glu	Asn	Gln 105	His	Thr	Ile	Asp	Leu 110	Thr	Asp
<i>J J</i>	Ser	Glu	Met 115	Asn	Lys	Leu	Phe	Glu 120	Lys	Thr	Arg	Arg	Gln 125	Leu	Arg	Glu
40	Asn	Ala 130	Glu	Asp	Met	Gly	Asn 135	Gly	Cys	Phe	Lys	Ile 140	Tyr	His	Lys	Cys
	Asp 145	Asn	Ala	Cys	Ile	Gly 150	Ser	Ile	Arg	Asn	Gly 155	Thr	Tyr	Asp	His	Asp 160
45	Val	Tyr	Arg	Asp	Glu 165	Ala	Leu	Asn	Asn	Arg 170	Phe	Gln	Ile	Lys	Gly 175	Val
50	Glu	Leu	Lys	Sér 180	Gly	Tyr	Lys	Asp	Trp 185	Ile	Leu	Trp	Ile	Ser. 190	Phe	Ala
J 0	Ile	Ser	Cys 195	Phe	Leu	Leu	Cys	Val 200	Val	Lėu	Leu	Gly	Phe 205	Ile	Met	Trp

```
Ala Cys Gln Lys Gly Asn Ile Arg Cys Asn Ile Cys Ile
                                215
   5
       (2) INFORMATION FOR SEQ ID NO:61:
             (i) SEQUENCE CHARACTERISTICS:
  10
                  (A) LENGTH: 5 amino acids
                  (B) TYPE: amino acid
                  (C) STRANDEDNESS: single
                  (D) TOPOLOGY: unknown
. 15
            (ii) MOLECULE TYPE: protein
            (x1) SEQUENCE DESCRIPTION: SEQ ID NO:61:
            Arg Arg Xaa Xaa Arg
  20
 25
       (2) INFORMATION FOR SEQ ID NO:62:
            (i) SEQUENCE CHARACTERISTICS:
                 (A) LENGTH: 15 base pairs
                 (B) TYPE: nucleic acid
 30
                 (C) STRANDEDNESS: double
                 (D) TOPOLOGY: unknown
           (ii) MOLECULE TYPE: DNA (genomic)
 35
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:
       CGNCGNNNNN NNCGN
                                                                               15
 40
       (2) INFORMATION FOR SEQ ID NO:63:
            (i) SEQUENCE CHARACTERISTICS:
 45
                 (A) LENGTH: 15 base pairs
                 (B) TYPE: nucleic acid
                 (C) STRANDEDNESS: double
                 (D) TOPOLOGY: unknown
 50
           (ii) MOLECULE TYPE: protein
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:
 55
      AGRAGRNNNN NNAGR
                                                                               15
```

5	(2) INFORMATION FOR SEQ ID NO:64:	
10	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 47 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
	(ii) MOLECULE TYPE: DNA (genomic)	
15	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:	
	CATGGATCAT ATGTTAACAG ATATCAAGGC CTGACTGACT GAGAGCT	47
20		
	(2) INFORMATION FOR SEQ ID NO:65:	
25	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 39 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
30	(ii) MOLECULE TYPE: DNA (genomic)	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:65:	
35	CTCAGTCAGT CAGGCCTTGA TATCTGTTAA CATATGATC	39
40	(2) INFORMATION FOR SEQ ID NO:66:	
45	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 29 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single	
40	(D) TOPOLOGY: unknown	
	(11) MOLECULE TYPE: DNA (genomic)	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:	
	CATGGGCGCC CATATGGGCA TATTCGGCG	29

	(2) INFORMATION FOR SEQ ID NO:67:	
5	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 23 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
10	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:	
15	CCGAATATGC CCATATGGGC GCC	23
20	(2) INFORMATION FOR SEQ ID NO:68:  (i) SEQUENCE CHARACTERISTICS:	
25	(A) LENGTH: 54 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown	
	(ii) MOLECULE TYPE: DNA (genomic)	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:  ARACTGTTTG ARABACACG TCGTCAACTG CGTGAAAATG CTGACGACAT GGGC	
35	(2) INFORMATION FOR SEQ ID NO:69:	54
40	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 145 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: unknown	
	(ii) MOLECULE TYPE: protein	
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:	
	Ile Gln Asp Leu Glu Lys Tyr Val Glu Asp Thr Lys Ile Asp Leu Trp 1 5 10 15	
50	Ser Tyr Asn Ala Glu Leu Leu Val Ala Leu Glu Asn Gln His Thr Ile 20 25 30	
55	Asp Leu Thr Asp Ser Glu Met Asn Lys Leu Phe Glu Lys Thr Arg Arg 35 40 45	

		Gln	Leu 50	Arg	Glu	Asn	Ala	Glu 55	Asp	Met	Gly	Asn	Gly 60	Cys	Phe	Lys	Ile
5		Tyr 65	His	Lys	Суѕ	Asp	Asn 70	Ala	Cys	Ile	Gly	Ser 75	Ile	Arg	Asn	Gly	Thr 80
		Tyr	Asp	His	Asp	Val 85	Tyr	Arg	Asp	Glu	Ala 90	Leu	Asn	Asn	Arg	Phe 95	Gln
10		Ile	Lys	Gly	Val 100	Glu	Leu	Lys		Gly 105		Lys	Asp	Trp	Ile 110	Leu	Trp
15		Ile	Ser	Phe 115	Ala	Ile	Ser	Cys	Phe 120	Leu	Leu	Cys	Val	Val 125	Leu	Leu	Gly
13		Phe	Ile 130	Met	Trp	Ala	Cys	Gln 135	Lys	Gly	Asn	Ile	Arg 140	Cys	Asn	Ile	Суs
20		Ile 145															
25	(2)	INFO	RMAT:	ION I	FOR :	SEQ :	ID N	D: <b>7</b> 0	<b>:</b>							,	
30		(i)	(A) (B) (C)	LEI TYI	E CHANGTH PE: 6 RANDI POLO6	: 149 amino EDNES	5 am: o ac: SS:	ino a id sing:	acid	5							
		(ii)	MOL	ECULI	E TY	PE: 1	prot	ein					•				
35	•	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: Si	EQ II	ои с	:70:						
<i>:</i>		Ile 1	Gln	Asp	Leu	Glu 5	Lys	Tyr	Val	Glu	Asp 10	Thr	Lys	Ile	Asp	Leu 15	Trp
40		Ser	Tyr	Asn	Ala 20	Glu	Leu	Leu	Val	Ala 25	Leu	Ġlu	Asn	Gln	His 30	Thr	Ile
45		Asp	Leu	Thr 35	Asp	Ser	Glu	Met	Asn 40	Lys	Leu	Phe	Glu	Lys 45	Thr	Arg	Arg
43		Gln	Leu 50	Arg	Glu	Asn	Ala	Glu 55	Asp	Met	Gly	Asn	Gly 60	Cys	Phe	Lys	Ile
50		Tyr 65	His	Lys	Cys	asp	Asn 70	Ala	Cys	Ile	Gly	Ser 75	Ile	Arg	Asn	Gly	Thr 80
		Tyr	Asp	His	Asp	Val	Tyr	Arg	Asp	Glu	Ala	Leu	Asn	Asn	Arg	Phe	Gln

		Ile	Lys	G1	y Va 10		u Le	u Ly	s Se	r Gl 10		r Ly	s As	o Tr	0 Il 11	e Leu O	Trp	
5		Ile	Ser	Phe 115		a Il	e Se:	r Cy	s Ph		u Le	u Cy	s Vai	1 Va:		u Leu	Gly	
		Phe	Ile 130		Tr	p Al	а Су	s Gl:		s Gl	y As	n Il	e Arg	-	s As	n Ile	Cys	
10		Ile 145																
15	(2)	INFO	RMAT	ION	FOR	SEQ	ID 1	NO:7	1:									
20		(i)	(A (B (C	) LE ) TY ) ST	ENGT: (PE : [RAN]	H: 6 nuc DEDNI	CTER: 90 ba leic ESS: unkr	ase p acid doub	pair:	5								
25		(ii)	MOL	ECUI	LE T	YPE:	DNA	(gei	nomi	C)								
30		(ix)		) NA	ME/I	KEY: ION:	CDS 1	590				,						
		(xi)	SEQ	ÚENC	E DE	ESCR	IPTIC	on: s	SEQ I	ID NO	):71	:						
35	ATG Met 1	GAT (	CCA /	AAC Asn	ACT Thr 5	GTG Val	TCA Ser	AGC Ser	TTT Phe	CAG Gln 10	GTA Val	GAT Asp	TGC Cys	TTT Phe	CTT Leu 15	TGG Trp		48
40	CAT His	GTC (	CGC Arg :	AAA Lys 20	CGA Arg	GTT Val	GCA Ala	GAC Asp	CAA Gln 25	GAA Glu	CTA Leu	GGT Gly	GAT Asp	GCC Ala 30	CCA Pro	TTC Phe		96
45	CTT Leu	GAT (	CGG ( Arg 1	CTT Leu	CGC Arg	CGA Arg	GAT Asp	CAG Gln 40	AAA Lys	TCC Ser	CTA Leu	AGA Arg	GGA Gly 45	AGG Arg	GGC GGC	AGC Ser		144
70	ACT Thr	CTT ( Leu ( 50	GGT (	CTG Leu	GAC Asp	ATC Ile	GAG Glu 55	ACA Thr	GCC Ala	ACA Thr	CGT Arg	GCT Ala 60	GGA Gly	AAG Lys	CAG Gln	ATA Ile		192
50	GTG Val 65	GAG ( Glu /	CGG 1 Arg :	ATT Ile	CTG Leu	AAA Lys 70	GAA Glu	GAA Glu	TCC Ser	GAT Asp	GAG Glu 75	GCA Ala	CTT Leu	AAA Lys	ATG Met	ACC Thr 80		240

											Lys	TAC Tyr					288
5												CTT Leu					336
10												ATG Met					384
15												GAG Glu 140					432
20	GGT Gly 145	TGC Cys	TTC Phe	AAA Lys	ATA Ile	TAC Tyr 150	CAC His	AAA Lys	TGT Cys	GAC Asp	AAT Asn 155	GCT Ala	TGC Cys	ATA Ile	GGG Gly	TCA Ser 160	480
25	ATC Ile	AGA Arg	AAT Asn	GGG Gly	ACT Thr 165	TAT Tyr	GAC Asp	CAT His	GAT Asp	GTA Val 170	TAC Tyr	AGA Arg	GAC Asp	GAA Glu	GCA Ala 175	TTA Leu	528
30	AAC Asn	AAC Asn	CGG Arg	TTT Phe 180	CAG Gln	ATC Ile	AAA Lys	GGT Gly	GTT Val 185	GAA Glu	CTG Leu	AAG Lys	TCA Ser	GGA Gly 190	TAC Tyr	AAA Lys	576
35	GAC Asp	TGG Trp	ATC Ile 195	CTG Leu	TGG Trp	ATT Ile	TCC Ser	TTT Phe 200	GCC Ala	ATA Ile	TCA Ser	TGC Cys	TTT Phe 205	TTG Leu	CTT Leu	TGT Cys	624
40	GTT Val	GTT Val 210	TTG Leu	CTG Leu	GGG Gly	TTC Phe	ATC Ile 215	ATG Met	TGG Trp	GCC Ala	TGC Cys	CAA Gln 220	AAA Lys	GGC Gly	AAC Asn	ATT Ile	672
45				ATT Ile													690

2) INFORMATION	FOR	SEQ	ID	NO:72:
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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 230 amino acids
- 5 (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

#### (ii) MOLECULE TYPE: protein

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:

Met Asp Pro Asn Thr Val Ser Ser Phe Gln Val Asp Cys Phe Leu Trp  $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$ 

His Val Arg Lys Arg Val Ala Asp Gln Glu Leu Gly Asp Ala Pro Phe 20 25 30

Leu Asp Arg Leu Arg Arg Asp Gln Lys Ser Leu Arg Gly Arg Gly Ser
35 40 45

Thr Leu Gly Leu Asp Ile Glu Thr Ala Thr Arg Ala Gly Lys Gln Ile
50 55 60

Val Glu Arg Ile Leu Lys Glu Glu Ser Asp Glu Ala Leu Lys Met Thr 25 65 70 75 80

Met Asp His Met Leu Ile Gln Asp Leu Glu Lys Tyr Val Glu Asp Thr 85 90 95

30 Lys Ile Asp Leu Trp Ser Tyr Asn Ala Glu Leu Leu Val Ala Leu Glu 100 105 110

Asn Gln His Thr Ile Asp Leu Thr Asp Ser Glu Met Asn Lys Leu Phe
115 120 125

35
Glu Lys Thr Arg Arg Gln Leu Arg Glu Asn Ala Glu Asp Met Gly Asn
130
135
140

Gly Cys Phe Lys Ile Tyr His Lys Cys Asp Asn Ala Cys Ile Gly Ser 40 145 150 155 160

Ile Arg Asn Gly Thr Tyr Asp His Asp Val Tyr Arg Asp Glu Ala Leu 165 170 175

45 Asn Asn Arg Phe Gln Ile Lys Gly Val Glu Leu Lys Ser Gly Tyr Lys 180 185 190

Asp Trp Ile Leu Trp Ile Ser Phe Ala Ile Ser Cys Phe Leu Leu Cys 195 200 205

Val Val Leu Leu Gly Phe Ile Met Trp Ala Cys Gln Lys Gly Asn Ile
210 215 220

#### WHAT IS CLAIMED IS:

1. A vaccine for stimulating protection in animals against infection by influenza virus which comprises an effective amount of an immunogenic fragment of the HA2 subunit of an HA protein selected from the group consisting of a Type A subtype influenza virus and a Type B influenza virus.

2. The vaccine according to claim 1 wherein said Type A subunit is H3N2.

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- 3. The vaccine according to claim 1 wherein the polypeptide is fused to a second polypeptide.
- 4. The vaccine according to claim 3 wherein the second polypeptide comprises the N terminal amino acids of influenza NS1 protein.
- 5. The vaccine according to claim 1 wherein the immunogenic fragment of the HA2 subunit is selected from the group consisting of a peptide comprising amino acids 1 to 221 of the H3HA2 subtype, a peptide comprising amino acids 77 to 221 of the H3HA2 subtype, a peptide comprising amino acids 1 to 223 of the BHA2 Type, and a peptide comprising amino acids 41 to 223 of the BHA2 type.
- 6. The vaccine according to claim 5 comprising NS1<sub>(1-25)</sub> 81)H3HA2<sub>(1-221)</sub> SEQ ID NO: 10.
  - 7. The vaccine according to claim 5 comprising NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> SEQ ID NO: 12.
- 30 8. The vaccine according to claim 5 comprising NS1<sub>(1-42)</sub>BLHA2<sub>(41-223)</sub> SEQ ID NO: 14.
  - 9. The vaccine according to claim 5 comprising NS1<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub> SEQ ID NO: 57.

10. The vaccine according to claim 5 comprising NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> SEQ ID NO:10 and NS1<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub>(met-leu) SEQ ID NO: 55.

- 5 11. A protein comprising an immunogenic fragment of the HA2 subunit of an HA protein selected from the group consisting of Type A subtype or Type B influenza virus.
- 12. The protein according to claim 11 wherein said Type A subtype is H3N2.
  - 13. The protein according to claim 11 wherein the peptide containing the immunogenic fragment is fused to a second peptide or protein.
- 15 14. The protein according to claim 13 wherein the second peptide comprises the N terminal amino acids of a NS1 protein.
- 15. The protein according to claim 11 wherein the immunogenic fragment of the HA2 subunit is selected from the group consisting of a peptide comprising amino acids 1 to 221 of the H3HA2 subunit, a peptide comprising amino acids 77 to 221 of the H3HA2 subunit, a peptide comprising amino acids 1-223 of the BHA2 subunit, and a peptide comprising amino acids 41-223 of the BHA2 subunit.
- 25 16. A polypeptide NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> SEQ ID NO: 10.
  - 17. A polypeptide NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> SEQ ID NO: 12.
  - 18. A polypeptide NS1<sub>(1-41)</sub>BLHA2<sub>(41-223)</sub> SEQ ID NO: 14.
  - 19. A polypeptide NS1<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub> SEQ ID NO: 57.
  - 20. A polypeptide  $NS1_{(1-81)}BLHA2_{(1-223)}$ (met-leu) SEQ ID NO: 55.

35

21. A DNA molecule comprising a coding sequence for an immunogenic fragment of the HA2 subunit of an HA protein selected from the group consisting of a Type A subtype or Type B influenza virus.

- 5 22. The DNA molecule according to claim 21 wherein said Type A subunit is H3N2.
  - 23. The DNA molecule according to claim 22 comprising a coding sequence for the polypeptide NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> SEQ ID NO: 10.
  - 24. The DNA molecule according to claim 21 comprising a coding sequence for the polypeptide NS1<sub>(1-42)</sub>BLHA2<sub>(41-223)</sub> SEQ ID NO: 14.
- 25. The DNA molecule according to claim 21 comprising a coding sequence for the polypeptide NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> SEQ ID NO: 12.
  - 26. The DNA molecule according to claim 21 comprising a coding sequence for the polypeptide NS1<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub> SEQ ID NO: 57.
- 20 27. A vector pOTS208NS1BLmut2 SEQ ID NO: 54.

- 28. A microorganism transformed with a DNA molecule comprising a coding sequence for an immunogenic fragment of the HA2 subunit of an HA protein selected from the group consisting of a Type A subtype or Type B influenza virus.
  - 29. The microorganism according to claim 28 wherein said Type A subunit is H3N2.
- 30. The microorganism according to claim 28 wherein said DNA molecule comprises a coding sequence for the polypeptide NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> SEQ ID NO: 10.
- 31. The microorganism according to claim 28 wherein said DNA molecule comprises a coding sequence for the polypeptide NS1<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub> SEQ ID NO: 57.

32. The microorganism according to claim 28 wherein said DNA molecule comprises a coding sequence for the polypeptide NS1<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub>(met-leu) SEQ ID NO: 55.

- 33. A combination vaccine for stimulating protection in animals against infection by influenza virus which comprises a first polypeptide having an immunogenic fragment of the HA2 subunit of an influenza H3 subtype virus and a second polypeptide selected from the group consisting of a polypeptide having an immunogenic fragment of the HA2 subunit of a Type B influenza virus, and a polypeptide having an immunogenic fragment of the HA2 subunit of an H1 subtype influenza virus, and a polypeptide having an immunogenic fragment of the HA2 subunit of an H2 subtype influenza virus.
- 34. The combination vaccine according to claim 33 wherein the first polypeptide is selected from the group consisting of NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> SEQ ID NO: 10 and NS1<sub>(1-81)</sub>H3HA2<sub>(77-221)</sub> SEQ ID NO: 12.
  - 35. The combination vaccine according to claim 33 wherein the second polypeptide is a polypeptide having an immunogenic fragment of the HA2 subunit of an H1 subtype influenza virus.

- 36. The combination vaccine according to claim 33 wherein said second polypeptide is selected from the group consisting of C13 SEQ ID NO: 16, D SEQ ID NO: 18, C13 short SEQ ID NO: 20, D short SEQ ID NO: 22, A SEQ ID NO: 24, C SEQ ID NO: 26, ΔD SEQ ID NO: 27, Δ13 SEQ ID NO: 28, M SEQ ID NO: 29, ΔM SEQ ID NO: 30, ΔM+ SEQ ID NO: 32, and H1HA266-222 SEQ ID NO: 34.
- 37. The combination vaccine according to claim 33 wherein said second polypeptide is NS1(1-42)BLHA2(41-223) SEQ ID NO: 14.
  - 38. The combination vaccine according to claim 33 wherein said second polypeptide is NS1<sub>(1-81)</sub>BLHA2<sub>(1-223)</sub> SEQ ID NO: 57.
- 35 39. A combination vaccine for stimulating protection in animals against infection by influenza virus which comprises a first polypeptide having an immunogenic fragment of the HA2 subunit of an influenza H3 subtype virus, a

second polypeptide having an immunogenic fragment of the HA2 subunit of an influenza B Type virus, and a third polypeptide selected from the group consisting of a polypeptide having an immunogenic fragment of the HA2 subunit of an H1 subtype influenza virus and a polypeptide having an immunogenic fragment of the HA2 subunit of an H2 subtype influenza virus.

40. The combination vaccine according to claim 39 wherein the first polypeptides is NS1<sub>(1-81)</sub>H3HA2<sub>(1-221)</sub> SEQ ID NO: 10, the second polypeptide is NS1<sub>(1-81)</sub>BHA2<sub>(1-223)</sub>(met-leu) SEQ ID NO: 57, and the third polypeptide is NS1<sub>(1-81)</sub>HA2<sub>(65-222)</sub> SEQ ID NO: 18.

# FIGURE 1

1-1						
(a)						
(b)						
(c)				ggga-		
(d)	GGCATATTCG	GCGCAATAGC	AGGTTTCATA	GAAAATGGTT	GGGAGGGAAT	50
(a)				t		
(b)				c		
(c)	ta		atcat	ggaac	atct	
(d)	GATAGACGGT	TGGTACGGTT	TCAGGCATCA	AAATTC-GAG	GGCACAGGAC	100
(a)					~~~~~	
(p)						
(c)	-tg	aa	aaat-	tagg	gt-caaac	
(d)				CCATCGACCA		150
(a)			*********			
(b)						
(c)	ggct	ctt	a-t	attca	cagctg-g-g	
(d)	AAACTGAATA	GGGTAATCGA	GAAGACGAAC	GAGAAATTCC	ATCAAATCGA	200
•						200
(a)						
(p)						
(c)	ta	aacat	aaag	gg-aa-tt-a	a-ta-a-	
(d)	AAAGGAATTC	TCAGAAGTAG	AAGGGAGAAT	TCAGGACCTC	GAGAAATACG	250
(a)						
(b)						
(c)				-ta		
(d)	TTGAAGACAC	TAAAATAGAT	CTCTGGTCTT	ACAATGCGGA	GCTTCTTGTC	300
(a)						
(b)						
(c)	ctaa-	-tgagg	tc-at-c	caaa	-taa	
(d)	GCTCTGGAGA	ACCAACATAC	AATTGATCTG	ACTGACTCGG	AAATGAACAA	350
						330
(a)						
(b)						
(c)	tg	gtaa-	-ct-a-a	-a-tc	a-aac-	
(d)	ACTGTTTGAA	AAAACAAGGA	GGCAACTGAG	GGAAAATGCT	GAGGACATGG	400
(a)						
(b)						
(C)	-aa	ttg-gt-c	g-	a	ag-aa	
(d)	GCAATGGTTG	CTTCAAAATA	TACCACAAAT	GTGACAATGC	TTGCATAGGG	450
(a)						
(b)						
(c)	agtg-a			aattc		
(d)	TCAATCAGAA	ATGGGACTTA	TGACCATGAT	GTATACAGAG	ACGAAGCAMM	E00
					ormidchil	500
(a)						
(b)					~	
(C)	gttga	gaaag-ag	-taga-	-tg-a	atgggg-tct	
(d)	AAACAACCGG	TTTCAGATC?	AAGGTGTTGA	ACTGAAGTCA	GGATACAAAG	550

# FIGURE 1 (cont'd)

(a)						
(b)		~~				
(c)	-tcat	-gcc-a-	-caa-tg-cg	-ca-t-cac-	-g-gct-t-g	
(d)	ACTGGATCCT	GTGGATTTCC	TTTGCCATAT	CATGCTTTTT	GCTTTGTGTT	600
		•,				
(a)				g		
(b)				a		
(c)	c-cc	gcag	tt-catg	ttctt-	-atctt-gca	
(d)	GTTTTGCTGG	GGTTCATCA-	TGTGGGCC	TGCCA-AAAG	GCAACATTAG	650
		. *				•
(a)						
(b)						
(c)	gaa	g				
(d)	GTGCAACATT	TGCATTTGA-				670

## FIGURE 2

Met					GTG Val				Gln					42
					AAA									84
15	_				20				-	25			Gly	
					GAT Asp									12
					AGC Ser									168
					AAG Lys									210
					GCA Ala									252
					GCA Ala 90									294
					GAC Asp									336
AAT Asn	TCT Ser	GAG Glu 115	GGC Gly	ACA Thr	GGA Gly	CAA Gln	GCA Ala 120	GCA Ala	GAT Asp	CTT Leu	AAA Lys	AGC Ser 125	ACT Thr	378
					CAA Gln									420
					GAG Glu									462
TTC Phe 155	TCA Ser	GAA Glu	GTA Val	GAA Glu	GGG Gly 160	AGA Arg	ATT Ile	CAG Gln	GAC Asp	CTC Leu 165	GAG Glu	AAA Lys	TAC Tyr	504
GTT Val	GAA Glu 170	GAC Asp	ACT Thr	AAA Lys	ATA Ile	GAT Asp 175	CTC Leu	TGG Trp	TCT Ser	TAC Tyr	AAT Asn 180	Ala	GAG Glu	546

# FIGURE 2 (cont'd)

											GAT Asp			588
		185	•				190				•	195		
GAC	TCG	GAA	ATG	AAC	AAA	CTG	ጥጥጥ	GAA	ΔΔΔ	מים	C T	C T	CNA	630
Asp	Ser	Glu	Met 200	Asn	Lys	Leu	Phe	Glu	Lys	Thr	Arg	Arg	Gln	630
	C T							205					210	
CTG	AGG	GAA	AAT	GCT	GAG	GAC	ATG	GGC	AAT	GGT	TGC Cys	TTC	AAA	672
200		O1u	non	215	GIU	иор	Met	GIY	220	GIĀ	Cys	Pne	rys	
ATA	TAC	CAC	AAA	TGT	GAC	AAT	GCT	TGC	ATA	GGG	TCA	ATC	AGA	714
225	туr	His	Lys	Cys	230	Asn	Ala	Cys	Ile	Gly 235	Ser	Ile	Arg	
AAT	GGG	ACT	TAT	GAC	CAT	GAT	GTA	TAC	AGA	GAC	GAA	GCA	TTA	756
Asn	G1y 240	Thr	Tyr	Asp	His	Asp 245	Val	Tyr	Arg	Asp	Glu 250	Ala	Leu	
AAC	AAC	CGG	TTT	CAG	ATC	AAA	GGT	GTT	GAA	CTG	AAG	TCA	GGA	798
Asn	Asn	Arg 255	Phe	Gln	Ile	Lys	Gly 260	Val	Glu	Leu	Lys	Ser 265	Ģly	
TAC	AAA	GAC	TGG	ATC	CTG	TGG	ATT	TCC	TTT	GCC	ATA	TCA	TGC	840
Tyr	rys	Asp	Trp 270	Ile	Leu	Trp	Ile	Ser 275	Phe	Ala	Ile	Ser	Cys 280	F
TTT	TTG	CTT	TGT	GTT	GTT	TTG	CTG	GGG	TTC	ATC	ATG	TGG	GCC	882
Phe	Leu	Leu	Суз	Val 285	Val	Leu	Leu	Gly	Phe 290	Ile	Met	Trp	Ala	
TGC	CAA	AAA	GGC	AAC	ATT	AGG	TGC	AAC	ATT	TGC	ATT			918
Cys 295	Gln	Lys	Gly	Asn	Ile 300	Arg	Суз	Asn	Ile	Cys 305	Ile			

## FIGURE 3

														CTT Leu 15	45
		GTC Val												GCC Ala 30	90
														GGA Gly 45	135
		AGC Ser													100
		CAG Gln													225
GCA Ala	CTT Leu	AAA Lys	ATG Met	ACC Thr 80	ATG Met	GAT Asp	CAT His	ATG Met	TTA Leu 85	ATT Ile	CAG Gln	GAC Asp	CTC Leu	GAG Glu 90	270
AAA Lys	TAC Tyr	GTT Val	GAA Glu	GAC Asp 95	ACT Thr	AAA Lys	ATĀ Ile	GAT Asp	CTC Leu 100	TGG Trp	TCT Ser	TAC Tyr	AAT Asn	GCG Ala 105	315
GAG Glu	CTT Leu	CTT Leu	GTC Val	GCT Ala 110	CTG Leu	GAG Glu	AAC Asn	CAA Gln	CAT His 115	ACA Thr	ATT Ile	GAT Asp	CTG Leu	ACT Thr 120	360
GAC Asp	TCG Ser	GAA Glu	ATG Met	AAC Asn 125	AAA Lys	CTG Leu	TTT Phe	GAA Glu	AAA Lys 130	ACA Thr	AGG Arg	AGG Arg	CAA Gln	CTG Leu 135	405
AGG Arg	GAA Glu	AAT Asn	GCT Ala	GAG Glu 140	GAC Asp	ATG Met	GGC Gly	AAT Asn	GGT Gly 145	TGC Cys	TTC Phe	AAA Lys	ATA Ile	TAC Tyr 150	450
CAC His	AAA Lys	TGT Cys	GAC Asp	AAT Asn 155	GCT Ala	TGC Cys	ATA Ile	GGG Gly	TCA Ser 160	ATC Ile	AGA Arg	AAT Asn	GGG Gly	ACT Thr 165	495
TAT Tyr	GAC Asp	CAT His	GAT Asp	GTA Val 170	TAC Tyr	AGA Arg	GAC Asp	GAA Glu	GCA Ala 175	TTA Leu	AAC Asn	AAC Asn	CGG <sup>.</sup>	TTT Phe 180	540
CAG Gln	ATC Ile	AAA Lys	GGT Gly	GTT Val 185	GAA Glu	CTG Leu	AAG Lys	TCA Ser	GGA Gly 190	TAC Tyr	AAA Lys	GAC Asp	TGG Trp	ATC Ile 195	585

# FIGURE 3 (cont'd)

													GTT Val		6	30
				200					205					210		
													ATT		6	75
Leu	Leu	Gly	Phe	Ile	Met	Trp	Ala	Суз	Gln	Lys	Gly	Asn	Ile	Arg		
				215					220	٠				225		
TGC	AAC	TTA	TGC	ATT		•									6	90
Cys	Asn	Ile	Cys	Ile 230												

## FIGURE 4

										Val				CTT Leu 15	45
														GCC Ala 30	90
														GGA Gly 45	135
TCA Ser	TAT Tyr	GTT Val	AAC Asn	AAG Lys 50	ACA Thr	CAA Gln	GAA Glu	GCT Ala	ATA Ile 55	AAC Asn	AAG Lys	ATA Ile	ACA Thr	AAA Lys 60	180
AAT Asn	CTC Leu	AAC Asn	TAT Tyr	TTA Leu 65	AGT Ser	GAG Glu	CTA Leu	GAA Glu	GTA Val 70	AAA Lys	AAC Asn	CTT Leu	CAA Gln	AGA Arg 75	225
CTA Leu	AGC Ser	GGA Gly	GCA Ala	ATG Met 80	AAT. Asn	GAG Glü	CTT Leu	CAC His	GAC Asp 85	GAA Glu	ATA Ile	CTC Leu	GAG Glu	CTA Leu 90	. 270
GAC Asp	GAA Glu	AAA Lys	GTG Val	GAT Asp 95	GAT Asp	CTA Leu	AGA Arg	GCT Ala	GAT Asp 100	ACA Thr	ATA Ile	AGC Ser	TCA Ser	CAA Gln 105	315
ATA Ile	GAG Glu	CTT Leu	GCA Ala	GTC Val 110	TTG Leu	CTT Leu	TCC Ser	AAC Asn	GAA Glu 115	GGG Gly	ATA Ile	ATA Ile	AAC Asn	AGT Ser 120	360
GAA Glu	GAT Asp	GAG Glu	CAT His	CTC Leu 125	TTG Leu	GCA Ala	CTT Leu	GAA Glu	AGA Arg 130	AAA Lys	CTG Leu	AAG Lys	AAA Lys	ATG Met 135	405
CTT Leu	GGC Gly	CCC Pro	TCT Ser	GCT Ala 140	GTA Val	GAA Glu	ATA Ile	GGG# Gly	AT G Asn 145	GG 1	rgc 1 Cys	TTT (	GAA A	Thr 150	450
AAA Lys	CAC His	AAA Lys	TGC Cys	AAC Asn 155	CAG Gln	ACT Thr	TGC Cys	CTA Leu	GAC Asp 160	AGG Arg	ATA Ile	GCT Ala	GCT Ala	GGC Gly 165	495
ACC Thr	TTT Phe	AAT Asn	GCA Ala	GGA Gly 170	GAT Asp	TTT Phe	TCT Ser	CŢT Leu	CCC Pro 175	ACT Thr	TTT Phe	GAT Asp	TCA Ser	TTA Leu 180	540
AAC Asn	ATT Ile	ACT Thr	GCT Ala	GCA Ala 185	TCT Ser	TTA Leu	AAT Asn	Asp	GAT Asp 190	GGC Gly	TTG Leu	GAT Asp	AAT Asn	CAT His 195	585

# FIGURE 4 (cont'd)

Thr	Ile	Leu	CTC	TAC Tyr 200	TAC Tyr	TCA Ser	ACT	GCT Ala	GCT Ala 205	TCT Ser	AGC Ser	TTG Leu	GCT Ala	GTA Val 210	630
ACA Thr	TTA Leu	ATG Met	ATA Ile	GCT Ala 215	ATC Ile	TTC Phe	ATT Ile	GTC Val	TAC Tyr 220	ATG Met	GTC Val	TCC Ser	AGA Arg	GAC Asp 225	675
AAT Asn	GTT Val	TCT Ser	TGT Cys	TCC Ser 230	ATC Ile	TGT Cys	CTG Leu							•	699

#### FIGURE 5

AATTCTCATG TTTGACAGCT TATCATCGAT AAGCTTCAGT TGAAGATATT AAGAACAGCC 60 TCGCAGATGA CGAATCATTG GGATTCCCAT CTTTTTTGTT TGTTGAAGGC GACACCATTG 120 GTTTTGCCAG AACTGTTTTC GGGCCGACCA CATCCGATCT GACAGATTTT TTAATCGGGA 180 AAGGAATGTC ATTAAGCAGT GGAGAGCGCG TTCAGATAGA GCCACTGATG AGGGGAACCA 240 CCAAAGACGA TGTTATGCAT ATGCATTTCA TCGGCCGAAC AACGGTGAAG GTAGAAGCCA 300 AGCTACCTGT ATTTGGCGAT ATATTAAAGG TCTTAGGGGC AACAGATATT GAAGGGGAGC 360 TTTTTGACTC ATTGGATATA GTCATTAAGC CAAAATTTAA AAGGGATATA AAAAAGGTTG 420 CCAAGGATAT TATTTTTAAC CCGTCACCTC AATTTTCAGA CATTAGCCTG CGGGCAAAAG 480 ATGAGGCCGG AGATATTTTA ACAGAACATT ATCTATCAGA AAAAGGCCAT CTCTCAGCGC 540 CTCTGAACAA GGTCACCAAT GCTGAGATAG CTGAAGAGAT GGCATATTGC TACGCAAGAA 600 TGAAAAGTGA TATACTGGAA TGTTTTAAAA GGCAGGTGGG CAAAGTTAAG GATTAATTAT 660 CAGGAGTAAT TATGCGGAAC AGAATCATGC CTGGTGTTTA CATAGTAATA ATTCCTTACG 720 TTATCGTAAG CATTTGCTAT CTCCTTTTCC GCCACTACAT TCCTGGTGTT TCTTTTCAG 780 CTCATAGAGA TGGTCTTGGG GCGACATTGT CATCATATGC AGGAACCATG ATTGCAATCC 840 TGATTGCTGC CTTGACGTTT CTAATCGGAA GCAGAACGCG CCGACTGGCC AAGATTAGAG 900 AGTATGGGTA TATGACATCG GTAGTTATTG TCTATGCCCT TAGTTTTGTT GAGCTTGGAG 960 CTTTGTTTTT CTGCGGGTTA TTGCTTCTTT CCAGCATAAG CGGCTACATG ATACCCACTA 1020 TCGCCATCGG CATTGCCTCT GCATCGTTCA TTCATATATG CATCCTTGTT TTCCAACTAT 1080 ATAATTTGAC CAGAGAACAA GAATAACCCG GCCTCAGCGC CGGGTTTTCT TTGCCTCACG 1140 ATCGCCCCCA AAACACATAA CCAATTGTAT TTATTGAAAA ATAAATAGAT ACAACTCACT 1200 AAACATAGCA ATTCAGATCT CTCACCTACC AAACAATGCC CCCCTGCAAA AAATAAATTC 1260 ATATAAAAAA CATACAGATA ACCATCTGCG GTGATAAATT ATCTCTGGCG GTGTTGACAT 1320 AAATACCACT GGCGGTGATA CTGAGCACAT CAGCAGGACG CACTGACCAC CATGAAGGTG 1380 ACGCTCTTAA AAATTAAGCC CTGAAGAAGG GCAGCATTCA AAGCAGAAGG CTTTGGGGTG 1440 TGTGATACGA AACGAAGCAT TGGCCGTAAG TGCGATTCCG GATTAGCTGC CAATGTGCCA 1500 ATCGCGGGGG GTTTTCGTTC AGGACTACAA CTGCCACACA CCACCAAAGC TAACTGACAG 1560 GAGAATCCAG ATGGATGCAC AAACACGCCG CCGCGAACGT CGCGCAGAGA AACAGGCTCA 1620

# FIGURE 5 (cont'd)

AT	GGAA	AGCA	GCA	AATC	acc '	тстт	GGTT	ഭദേദ	ממים	cccc	מ מ מי	ארכא	CMMC	CCA	AAGATT	m 160
															CCGAGT	
															AAGGGC.	
TC	TAAA	AAA	CCA	CACC	TAT (	GTG:	ratg(	CA T	TAT	TTGC.	A TA	CATT	CAAT	CAA	TTGTTA	T 186
CT	AAGG2	TAA	ACT:	TACA	Met	G GA: L Asp	CC Pro	AA A ISA C	1 Th	T GT r Va:	G TC. 1 Se	A AG	C TT	T CA	G GTA n Val	191:
GAT Asp	r TGC Cys	TTT Phe	CT:	ı Trp	G CAT	r GTC B Veil	CGC	C AAA J Lys 20	Arg	A GT	r GC	A GAG A Asp	C CAM O Glr 25	Gl	A CTA 1 Leu	1959
GG1 G1y	GAT Asp	GCC Ala 30	Pro	A TTC	CTI Leu	CAI Asp	CGC Arc	, Leu	CGC Arc	C CGA	A GAT	CAC Glr 40	Lys	TCC Ser	CTA Leu	2007
AGA Arg	GGA Gly 45	Arg	GLy	AGC Ser	ACC Thr	CTC Leu 50	Gly	CTG Leu	GAC Asp	ATC Ile	GAC Glu	Thr	GCC Ala	ACA Thr	CGT Arg	2055
GCT Ala 60	GTA	AAG Lys	Gln	ATA Ile	GTG Val 65	Glu	CGG Arg	ATT Ile	CTG Leu	AAA Lys 70	Glu	GAA Glu	TCC Ser	GAT Asp	GAG Glu 75	2103
GCA Ala	CTT Leu	AAA Lys	ATG Met	ACC Thr 80	ATG Met	GGT Gly	TTC Phe	TTC Phe	GGA Gly 85	Ala	ATT	GCT Ala	GGT Gly	TTC Phe 90	TTG Leu	2151
GAA Glu	A GGT Gly	GGT	TGG Trp 95	GAA Glu	GGT	ATG CTC Leu Met	ATT Ile	GCA Ala 100	GGT Gly	TGG Trp	CAC His	GGA Gly	TAC Tyr 105	ACA Thr	TCT Ser	2199
nis	GIÀ	110	HIS	Gly	Val	Ala	Val 115	Ala	Ala	Asp	Leu	Lys 120	Ser	Thr		2247
31u	125	TTE	Asn	AAG Lys	Ile	130	Lys	Asn	Leu	Asn	Tyr 135	Leu	Ser	Glu	Leu	2295
140	val	ъÃ2	ASII	CTT Leu	145	Arg	Leu	Ser	Gly	Ala 150	Met	Asn	Glu	Leu	His 155	2343
SAC Asp	GAA Glu	ATA Ile	CTC Leu	GAG Glu 160	CTA Leu	GAC Asp	GAA Glu	AAA Lys	GTG Val	GAT Asp	GAT Asp	CTA Leu	AGA Arg	GCT Ala	GAT Asp	2391

#### FIGURE 5 (cont'd)

																•	
															GAA Glu		243
	A ATC Ile	A ATC Ile	AAC Asn 190	AGT Ser	GAA Glu	T GAC Asp	GAG Glu	CAT His 195	CTC Leu	TTG Leu	GCA Ala	CTT Leu	GAA Glu 200	AGA Arg	AAA Lys	CTG Leu	248
	AAG Lys	AAA Lys 205	ATG Met	CTT Leu	GGC Gly	CCC Pro	TCT Ser 210	GCT Ala	GTA Val	GAA Glu	C ATA Ile	GGG Gly 215	T AAC Asn	G GGT Gly	TGC	TTT Phe	2535
															GCT Ala		2583
	GGC Gly	ACC Thr	TTT Phe	AAT Asn	GCA Ala 240	GGA Gly	GAT Asp	TTT Phe	TCT Ser	CTT Leu 245	CCC Pro	ACT Thr	TTT Phe	GAT Asp	TCA Ser 250	TTA Leu	2631
	AAC Asn	ATT Ile	ACT Thr	GCT Ala 255	GCA Ala	TCT Ser	TTA Leu	AAT Asn	GAT Asp 260	GAT Asp	Gly	TTG Leu	GAT Asp	AAT Asn 265	CAT His	ACT Thr	2679
	ATA Ile	CTG Leu	CTC Leu 270	TAC Tyr	TAC Tyr	TCA Ser	ACT Thr	GCT Ala 275	GCT Ala	TCT Ser	AGC Ser	TTG Leu	GCT Ala 280	GTA Val	ACA Thr	TTA . Leu	2727
	ATG Met	ATA Ile 285	GCT Ala	ATC Ile	TTC Phe	ATT	GTC Val 290	TAC Tyr	ATG Met	GTC Val	TCC Ser	AGA Arg 295	GAC Asp	AAT Asn	GTT Val	TCT Ser	2775
				TGT Cys		TGAG	GGAG	AT I	TAAGO	CCTG	FT GI	TTTC	CTTI	' ACI	rgtag	TGC	2830
	TCAT	TTGC	TT G	TCAC	CATI	'A CA	AAGA	AACG	TTA	TTGA	AAA	ATGO	TCTT	GT 1	'ACTA	.CTGAA	2890
	TTCT	'AGAA	TC G	ATAA	GCTI	'C GA	CCGA	TGCC	CTI	GAGA	GCC	TTCA	ACCC	AG I	CAGC	TCCTT	2950
																ATGCA	
																CGCTG	
																GCTCA	
																GCCGG	
ł	CATG	GCGG	CC G	ACGC	:GCTG	G GC	TACG	ጥጥጋጥ	CCT	GGCC	ግጥጥ:	CTCC	አርጥን	ልጥ ሶ	ነ አ ር ር ጦ	C2C22	2254

## FIGURE 5 (cont'd)

CTCCATCTGG ATTTGTTCAG AACGCTCGGT TGCCGCCGGG CGTTTTTTAT TGGTGAGAAT 3310 CGCAGCAACT TGTCGCGCCA ATCGAGCCAT GTCGTCGTCA ACGACCCCCC ATTCAAGAAC 3370 AGCAAGCAGC ATTGAGAACT TTGGAATCCA GTCCCTCTTC CACCTGCTGA GACGCGAGGC 3430 TGGATGGCCT TCCCCATTAT GATTCTTCTC GCTTCCGGCG GCATCGGGAT GCCCGCGTTG 3490 CAGGCCATGC TGTCCAGGCA GGTAGATGAC GACCATCAGG GACAGCTTCA AGGATCGCTC 3550 GCGGCTCTTA CCAGCCTAAC TTCGATCACT GGACCGCTGA TCGTCACGGC GATTTATGCC 3610 GCCTCGGCGA GCACATGGAA CGGGTTGGCA TGGATTGTAG GCGCCGCCCT ATACCTTGTC 3670 TGCCTCCCCG CGTTGCGTCG CGGTGCATGG AGCCGGGCCA CCTCGACCTG AATGGAAGCC 3730 GGCGGCACCT CGCTAACGGA TTCACCACTC CAAGAATTGG AGCCAATCAA TTCTTGCGGA 3790 GAACTGTGAA TGCGCAAACC AACCCTTGGC AGAACATATC CATCGCGTCC GCCATCTCCA 3850 GCAGCCGCAC GCGGCGCATC TCGGGCAGCG TTGGGTCCTG GCCACGGGTG CGCATGATCG 3910 TGCTCCTGTC GTTGAGGACC CGGCTAGGCT GGCGGGGTTG CCTTACTGGT TAGCAGAATG 3970 AATCACCGAT ACGCGAGCGA ACGTGAAGCG ACTGCTGCTG CAAAACGTCT GCGACCTGAG 4030 CAACAACATG AATGGTCTTC GGTTTCCGTG TTTCGTAAAG TCTGGAAACG CGGAAGTCAG 4090 CGCCCTGCAC CATTATGTTC CGGATCTGCA TCGCAGGATG CTGCTGGCTA CCCTGTGGAA 4150 CACCTACATC TGTATTAACG AAGCGCTGGC ATTGACCCTG AGTGATTTTT CTCTGGTCCC 4210 GCCGCATCCA TACCGCCAGT TGTTTACCCT CACAACGTTC CAGTAACCGG GCATGTTCAT 4270 CATCAGTAAC CCGTATCGTG AGCATCCTCT CTCGTTTCAT CGGTATCATT ACCCCCATGA 4330 ACAGAAATTC CCCCTTACAC GGAGGCATCA AGTGACCAAA CAGGAAAAAA CCGCCCTTAA 4390 CATGGCCCGC TTTATCAGAA GCCAGACATT AACGCTTCTG GAGAAACTCA ACGAGCTGGA 4450 CGCGGATGAA CAGGCAGACA TCTGTGAATC GCTTCACGAC CACGCTGATG AGCTTTACCG 4510 CAGCTGCCTC GCGCGTTTCG GTGATGACGG TGAAAACCTC TGACACATGC AGCTCCCGGA 4570 GACGGTCACA GCTTGTCTGT AAGCGGATGC CGGGAGCAGA CAAGCCCGTC AGGGCGCGTC 4630 AGCGGGTGTT GGCGGGTGTC GGGGCGCAGC CATGACCCAG TCACGTAGCG ATAGCGGAGT 4690 GTATACTGGC TTAACTATGC GGCATCAGAG CAGATTGTAC TGAGAGTGCA CCATATGCGG 4750 TGTGAAATAC CGCACAGATG CGTAAGGAGA AAATACCGCA TCAGGCGCTC TTCCGCTTCC 4810 TCGCTCACTG ACTCGCTGCG CTCGGTCGTT CGGCTGCGGC GAGCGGTATC AGCTCACTCA 4870

#### FIGURE 5 (cont'd)

AAGGCGGTAA TACGGTTATC CACAGAATCA GGGGATAACG CAGGAAAGAA CATGTGAGCA 4930 AAAGGCCAGC AAAAGGCCAG GAACCGTAAA AAGGCCGCGT TGCTGGCGTT TTTCCATAGG 4990 CTCCGCCCC CTGACGAGCA TCACAAAAAT CGACGCTCAA GTCAGAGGTG GCGAAACCCG 5050 ACAGGACTAT AAAGATACCA GGCGTTTCCC CCTGGAAGCT CCCTCGTGCG CTCTCCTGTT 5110 CCGACCCTGC CGCTTACCGG ATACCTGTCC GCCTTTCTCC CTTCGGGAAG CGTGGCGCTT 5170 TCTCAATGCT CACGCTGTAG GTATCTCAGT TCGGTGTAGG TCGTTCGCTC CAAGCTGGGC 5230 TGTGTGCACG AACCCCCCGT TCAGCCCGAC CGCTGCGCCT TATCCGGTAA CTATCGTCTT 5290 GAGTCCAACC CGGTAAGACA CGACTTATCG CCACTGGCAG CAGCCACTGG TAACAGGATT 5350 AGCAGAGCGA GGTATGTAGG CGGTGCTACA GAGTTCTTGA AGTGGTGGCC TAACTACGGC 5410 TACACTAGAA GGACAGTATT TGGTATCTGC GCTCTGCTGA AGCCAGTTAC CTTCGGAAAA 5470 AGAGTTGGTA GCTCTTGATC CGGCAAACAA ACCACCGCTG GTAGCGGTGG TTTTTTTGTT 5530 TGCAAGCAGC AGATTACGCG CAGAAAAAAA GGATCTCAAG AAGATCCTTT GATCTTTTCT 5590 ACGGGGTCTG ACGCTCAGTG GAACGAAAAC TCACGTTAAG GGATTTTGGT CATGAGATTA 5650 TCAAAAAGGA TCTTCACCTA GATCCTTTTA AATTAAAAAT GAAGTTTTAA ATCAATCTAA 5710 AGTATATATG AGTAAACTTG GTCTGACAGT TACCAATGCT TAATCAGTGA GGCACCTATC 5770 TCAGCGATCT GTCTATTTCG TTCATCCATA GTTGCCTGAC TCCCCGTCGT GTAGATAACT 5830 ACGATACGGG AGGGCTTACC ATCTGGCCCC AGTGCTGCAA TGATACCGCG AGACCCACGC 5890 TCACCGGCTC CAGATTTATC AGCAATAAAC CAGCCAGCCG GAAGGGCCGA GCGCAGAAGT 5950 GGTCCTGCAA CTTTATCCGC CTCCATCCAG TCTATTAATT GTTGCCGGGA AGCTAGAGTA 6010 AGTAGTTCGC CAGTTAATAG TTTGCGCAAC GTTGTTGCCA TTGCTGCAGG CATCGTGGTG 6070 TCACGCTCGT CGTTTGGTAT GGCTTCATTC AGCTCCGGTT CCCAACGATC AAGGCGAGTT 6130 ACATGATCCC CCATGTTGTG CAAAAAAGCG GTTAGCTCCT TCGGTCCTCC GATCGTTGTC 6190 AGAAGTAAGT TGGCCGCAGT GTTATCACTC ATGGTTATGG CAGCACTGCA TAATTCTCTT 6250 ACTGTCATGC CATCCGTAAG ATGCTTTTCT GTGACTGGTG AGTAGCTTCA CGCTGCCGCA 6310 AGCACTCAGG GCGCAAGGGC TGCTAAAGGA AGCGGAACAC GTAGAAAGCC AGTCCGCAGA 6370 AACGGTGCTG ACCCCGGATG AATGTCAGCT ACTGGGCTAT CTGGACAAGG GAAAACGCAA 6430 GCGCAAAGAG AAAGCAGGTA GCTTGCAGTG GGCTTACATG GCGATAGCTA GACTGGGCGG 6490

# FIGURE 5 (cont'd)

TTTTATGGAC	AGCAAGCGAA	CCGGAATTGC	CAGCTGGGGC	GCCCTCTGGT	AAGGTTGGGA	6550
AGCCCTGCAA	AGTAAACTGG	ATGGCTTTCT	TGCCGCCAAG	GATCTGATGG	CGCAGGGGAT	6610
CAAGATCTGA	TCAAGAGACA	GGATGAGGAT	CGTTTCGCAT	GATTGAACAA	GATGGATTGC	6670
ACGCAGGTTC	TCCGGCCGCT	TGGGTGGAGA	GGCTATTCGG	CTATGACTGG	GCACAACAGA	6730
CAATCGGCTG	CTCTGATGCC	GCCGTGTTCC	GGCTGTCAGC	GCAGGGGGCGC	CCGGTTCTTT	6790
TTGTCAAGAC	CGACCTGTCC	GGTGCCCTGA	ATGAACTGCA	GGACGAGGCA	GCGCGGCTAT	6850
CGTGGCTGGC	CACGACGGGC	GTTCCTTGCG	CAGCTGTGCT	CGACGTTGTC	ACTGAAGCGG	6910
GAAGGGACTG	GCTGCTATTG	GGCGAAGTGC	CGGGGCAGGA	TCTCCTGTCA	TCTCACCTTG	6970
CTCCTGCCGA	GAAAGTATCC	ATCATGGCTG	ATGCAATGCG	GCGGCTGCAT	ACGCTTGATC	7030
CGGCTACCTG	CCCATTCGAC	CACCAAGCGA	AACATCGCAT	CGAGCGAGCA	CGTACTCGGA	7090
TGGAAGCCGG	TCTTGTCGAT	CAGGATGATC	TGGACGAAGA	GCATCAGGGG	CTCGCGCCAG	7150
CCGAACTGTT	CGCCAGGCTC	AAGGCGCGCA	TGCCCGACGG	CGAGGATCTC	GTCGTGACTC	7210
ATGGCGATGC	CTGCTTGCCG	AATATCATGG	TGGAAAATGG	CCGCTTTTCT	GGATTCATCG	7270
ACTGTGGCCG	GCTGGGTGTG	GCGGACCGCT	ATCAGGACAT	AGCGTTGGCT	ACCCGTGATA	7330
TTGCTGAAGA	GCTTGGCGGC	GAATGGGCTG	ACCGCTTCCT	CGTGCTTTAC	GGTATCGCCG	7390
CTCCCGATTC	GCAGCGCATC	GCCTTCTATC	GCCTTCTTGA	CGAGTTCTTC	TGAGCGGGAC	7450
TCTGGGGTTC	GAAATGACCG	ACCAAGCGAC	GCCCAACCTG	CCATCACGAG	ATTTCGATTC	7510
CACCGCCGCC	TTCTATGAAA	GGTTGGGCTT	CGGAATCGTT	TTCCGGGACG	CCGGCTGGAT	7570
GATCCTCCAG	CGCGGGGATC	TCATGCTGGA	GTTCTTCGCC	CACCCC		7616

# Figure 6

NS1 ATGGATCCAAACACTGTGTCAAGCTTTCAGGTAGATTGCTTTCTTT
TACCTAGGTTTGTGACACAGTTCGAAAGTCCATCTAACGAAAGAAA
1 MetAspProAsnThrValSerSerPheGlnValAspCysPheLeuTrpHisValArgLys
CGAGTTGCAGACCAAGAACTAGGTGATGCCCCATTCCTTGATCGGCTTCGCCGAGATCAG
GCTCAACGTCTGGTTCTTGATCCACTACGGGGTAAGGAACTAGCCGAAGCGGCTCTAGTC
ArgValAlaAspGlnGluLeuGlyAspAlaProPheLeuAspArgLeuArgArgAspGlr
AAATCCCTAAGAGGAAGGGGCAGCACTCTTGGTCTGGACATCGAGACAGCCACACGTGCT
TTTAGGGATTCTCCTTCCCCGTCGTGAGAACCAGACCTGTAGCTCTGTCGGTGTGCACGA
LysSerLeuArgGlyArgGlySerThrLeuGlyLeuAspIleGluThrAlaThrArgAla
GGAAAGCAGATAGTGGAGCGGATTCTGAAAGAAGAATCCGATGAGGCACTTAAAATGACC
CCTTTCGTCTATCACCTCGCCTAAGACTTTCTTCTTAGGCTACTCCGTGAATTTTACTGG
GlyLysGlnIleValGluArgIleLeuLysGluGluSerAspGluAlaLeuLysMetThr
HA2
ATGCAGATCCCGGCTGTGGGTAAAGAATTCAACAAATTAGAAAAAAGGATGGAAAATTA
ATGCAGATCCCGGCTGTGGGTAAAGAATTCAACAAATTAGAAAAAAGGATGGAAAATTTA
ATGCAGATCCCGGCTGTGGGTAAAGAATTCAACAAATTAGAAAAAAGGATGGAAAATTTA TACGTCTAGGGCCGACACCCATTTCTTAAGTTGTTTAATCTTTTTTCCTACCTTTTAAAT  81 linker 65 69 MetGlnlleProAlaValGlyLysGluPheAsnLysLeuGluLysArgMetGluAsnLeu AATAAAAAAGTTGATGATGATGTTTCTGGACATTTGGACATATAATGCAGAATTGTTAGTT
ATGCAGATCCCGGCTGTGGGTAAAGAATTCAACAAATTAGAAAAAAGGATGGAAAATTTA TACGTCTAGGGCCGACACCCATTTCTTAAGTTGTTTAATCTTTTTTCCTACCTTTTAAAT  81 linker 65 69 MetGlnlleProAlaValGlyLysGluPheAsnLysLeuGluLysArgMetGluAsnLeu
ATGCAGATCCCGGCTGTGGGTAAAGAATTCAACAAATTAGAAAAAAGGATGGAAAATTTA TACGTCTAGGGCCGACACCCATTTCTTAAGTTGTTTAATCTTTTTTCCTACCTTTTAAAT  81 linker 65 69 MetGlnlleProAlaValGlyLysGluPheAsnLysLeuGluLysArgMetGluAsnLeu AATAAAAAAGTTGATGATGGATTTCTGGACATTTGGACATATAATGCAGAATTGTTAGTT
ATGCAGATCCCGGCTGTGGGTAAAGALTTCAACAAATTAGAAAAAAGGATGGAAAATTTA++++++
ATGCAGATCCCGGCTGTGGGTAAAGALTTCAACAAATTAGAAAAAAGGATGGAAAATTTA+++++

#### Figure 6 (Cont'd.)

AAAGTAAAAAGCCAATTAAAGAATAATGCCAAAGAAATCGGAAATGGATGTTTTGAGTTC \_\_\_\_\_\_ TTTCATTTTTCGGTTAATTTCTTATTACGGTTTCTTTAGCCTTTACCTACAAAACTCAAG LysValLysSerGlnLeuLysAsnAsnAlaLysGluIleGlyAsnGlyCysPheGluPhe TACCACAGTGTGACAATGAATGCATGGAAAGTGTAAGAAATGGGACTTATGATTATCCC ------ATGGTGTTCACACTGTTACTTACGTACCTTTCACATTCTTTACCCTGAATACTAATAGGG 150 TyrHisLysCysAspAsnGluCysMetGluSerValArgAsnGlyThrTyrAspTyrPro AAATATTCAGAAGAGTCAAAGTTGAACAGGGAAAAGGTAGATGGAGTGAAATTGGAATCA \_\_\_\_\_\_ TTTATAAGTCTTCTCAGTTTCAACTTGTCCCTTTTCCATCTACCTCACTTTAACCTTAGT LysTyrSerGluGluSerLysLeuAsnArgGluLysValAspGlyValLysLeuGluSer ATGGGGATCTATCAGATTCTGGCGATCTACTCAACTGTCGCCAGTTCACTGGTGCTTTTG TACCCCTAGATAGTCTAAGACCGCTAGATGAGTTGACAGCGGTCAAGTGACCACGAAAAC MetGlyIleTyrGlnIleLeuAlaIleTyrSerThrValAlaSerSerLeuValLeuLeu GTCTCCCTGGGGCAATCAGTTTCTGGATGTGTTCTAATGGATCTTTGCAGTGCAGAATA CAGAGGGACCCCGTTAGTCAAAGACCTACACAAGATTACCTAGAAACGTCACGTCTTAT ValSerLeuGlyAlaIleSerPheTrpMetCysSerAsnGlySerLeuGlnCysArgIle **TGCATCTGA** ----+--ACGTAGACT 222 CysIle

## FIGURE 7

		GGC Gly						39
		ATG Met						78
		GAG Glu 30						117
_		GCA Ala						156
	_	ATC Ile	-					195
		GAA Glu						234
		 AAA Lys						273
		AAT Asn 95						312
		ATT Ile						351
		AAA Lys						390
		GGC Gly						429
		 GCT Ala	-					468
		GAT Asp 160						507
	Phe	ATC Ile						546

# FIGURE 7 (cont'd)

											TCA			585
Lys	Asp	Trp	Ile	Leu	Trp	Ile	Ser	Phe	Ala	Ile	Ser	Cys		
		185					190					195		
											ATG		•	624
Phe	Leu	Leu	Cys	Val	Val	Leu	Leu	Gly	Phe	Ile	Met	Trp		
				200					205			-		
GCC	TGC	CAG	AAA	GGC	AAC	ATT	AGG	TGC	AAC	ATT	TGC	ATT		663
Ala	Cys	Gln	Lys	Gly	Asn	Ile	Arg	Cys	Asn	Ile	Cys	Ile		
	210					215					220			
TGA														666

## FIGURE 8

GGC	ATA	TTC	GGC	GCA	ATA	GCA	GGT	TTC	ATA	GAA	AAT	GGT		39
Gly	Ile	Phe	Gly	Ala	Ile	Ala	Gly	Phe	Ile	Glu	Asn	Gly		
1				5					10					
TCC	GAG	CCA	ATG	ልጥል	GAC	CCT	тес	ጥልር	ССТ	ጥጥር	ACC.	`. Сът		78
			Met											70
	15	1				20		-1	,		25			
									•					
			GAG									•		117
Gln	Asn	Ser	Glu	Gly	Thr	Gly	Gln		Ala	Asp	Leu	Lys		
			30					35						
AGC	ACT	CAA	GCA	GCC	ATC	GAC	CAA	ATC	AAT	GGG	AAA	CTG		156
Ser	Thr	Gln	Ala	Ala	Ile	Asp	Gln	Ile	Asn	Gly	Lys	Leu		
40					45					50				
		Om>		~>~							~~~	~		
			ATC Ile										•	195
ASII	nry	55	116	GIU	ָבעַם	1111	60	Gru	туз	FIIE	птэ	65		
							-							
			GAA											234
Ile	Glu	Lys	Glu		Ser	Glu	Val	Glu		Arg	Ile	Gln		
				70					75					
GAC	CTC	GAG	AAA	TAC	СТТ	GAA	GAC	ACT	AAA	АТА	CAT	CTC		273
			Lys											2.0
	80					85	-		_		90			
			AAT											312
ILD	Ser	ıyı	Asn 95	MIG	GIU	теп	ren	100	AIG	ren	GIU	Asn		
											•			
			ATT											351
	His	Thr	Ile	Asp		Thr	Asp	Ser	Glu		Asn	Lys		
105					110					115				
CTG	<b>T</b> TT	GAA	AAA	ACA	AGG	AGG	CAA	CTG	AGG	GAA	таа	ССТ		390
			Lys											330
		120	_		_	_	125					130		
			GGC											429
GIU	Asp	met	Gly	135	GTĀ	Cys	Pne	гÀ2	11e	Tyr	HIS	rys		
				133					140					
TGT	GAC	AAT	GĊT	TGC	ATA	GGG	TCA	ATC	AGA	AAT	GGG	ACT		468
Cys	Asp	Asn	Ala	Cys	Ile	Gly	Ser	Ile	Arg	Asn	Gly	Thr	•	
	145					150					155			
ጥልጥ	CAC	ጥልን	GAT	ርሞል	ጥልሶ	מממ	GAC	CAA	GC A	ጥጥአ	220	220		507
			Asp											507
- 4 -	<b>.</b> .		160	- <b></b>	- 4 -	y		165						

## FIGURE 8 (cont'd)

											GGA Gly		546
											TCA Ser		585
											ATG Met		624
GCC Ala	TGC Cys 210	CAA Gln	AAA Lys	GJA	AAC Asn	ATT Ile 215	AGG Arg	TGC Cys	AAC Asn	ATT Ile	TGC Cys 220	ATT Ile	663
TGA													666

In ational application No. PCT/US94/01149

1	SSIFICATION OF SUBJECT MATTER	12/00 15/00 17/00							
IPC(5) :A61K39/12; C12P 21/06, C12N 15/00; C07K 3/00, 13/00, 15/00, 17/00 US CL : 424/89; 530/350; 435/69.1, 172.1, 320.1									
	According to International Patent Classification (IPC) or to both national classification and IPC								
B. FIEI	LDS SEARCHED								
Minimum d	ocumentation searched (classification system followe	d by classification symbols)							
U.S. :	424/89; 530/350; 435/69.1, 172.1, 320.1								
Documental	Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched								
	lata base consulted during the international search (nalog, search terms: influenza, NS1, HA, vaccin	•	, search terms used)						
C. DOC	UMENTS CONSIDERED TO BE RELEVANT								
Category*	Citation of document, with indication, where a	ppropriate, of the relevant passages	Relevant to claim No.						
x	EPA 0,366,238 (YOUNG, et al), 02 May 1990, pages 1-25, see pages 3 and 7.								
x	The Journal of Immunology, Volume 140, Number 4, issued 15 February 1988, Kuwano et al, "HA2 Subunit of Influenza A H1 and H2 Subtype Viruses Induces A Protective Cross-Reactive Cytotoxic T Lymphocyte Response", pages 1264-1268, see entire document.								
X	Virology, Volume 178, issued 19 Reactive Protection Against Influe an NS1-Specific CTL Clone*, pa document.	enza A Virus Infections by	1-3, 11-13, 21, 22, 28, 29						
Furth	er documents are listed in the continuation of Box C	See patent family annex.							
*A* doc	ocial categories of cited documents:  cument defining the general state of the art which is not considered be part of particular relevance	"T" later document published after the inte date and not in conflict with the applies principle or theory underlying the inve	ation but cited to understand the						
°L° doc cite spe °O° doc	tier document published on or after the international filing date sument which may throw doubts on priority claim(s) or which is do to establish the publication date of another citation or other scial reason (as specified)	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone  "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art							
	rument published prior to the internstional filing date but later than priority date claimed	*&* document member of the same patent							
	Date of the actual completion of the international search  19 MAY 1994  Date of mailing of the international search report  JUN 01 1994								
Commission Box PCT Washington	nailing address of the ISA/US nor of Patents and Trademarks a, D.C. 20231 c. (703) 305-3230	Authorized officer  LYNETTE F. SMITH  Telephone No. (703) 308-0196							

Form PCT/ISA/210 (second sheet)(July 1992)\*

Int tional application No. PCT/US94/01149

Box I Ob	servations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This internal	tional report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
	Claims Nos.: eccause they relate to subject matter not required to be searched by this Authority, namely:
	Claims Nos.:
	ecause they relate to parts of the international application that do not comply with the prescribed requirements to such in extent that no meaningful international search can be carried out, specifically:
	Claims Nos.:
	ecause they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
lox II Ob	servations where unity of invention is lacking (Continuation of item 2 of first sheet)
his Intern	ational Searching Authority found multiple inventions in this international application, as follows:
Pleas	se See Extra Sheet.
	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite paymen of any additional fee.
	As only some of the required additional search fees were timely paid by the applicant, this international search report cover only those claims for which fees were paid, specifically claims Nos.:
، س	No required additional search fees were timely paid by the applicant. Consequently, this international search report i restricted to the invention first mentioned in the claims; it is covered by claims Nos.:  , 11-16,21-23, 27-30
Remark o	n Protest The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the payment of additional search fees

In ational application No. PCT/US94/01149

# BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

- I. Claims 1-6, 11-16, 21-23 and 27-30, drawn to a vaccine containing an immunogenic fragment of Type A influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- II. Claims 1-6, 11-16, 21-23 and 27-30, drawn to a vaccine containing an immunogenic fragment of Type B influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- III. Claims 1-5, 7, 11-15, 17, 21, 22, 25 and 27-29, drawn to a vaccine containing an immunogenic fragment of Type A influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- IV. Claims 1-5, 7, 11-15, 17, 21, 22, 25 and 27-29, drawn to a vaccine containing an immunogenic fragment of Type B influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- V. Claims 1-5, 8, 11-15, 18, 21, 22, 25 and 27-29, drawn to a vaccine containing an immunogenic fragment of Type A influenza virus and including onepolypeptide, protein, microorganism and vector, classes 424, 530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- VI. Claims 1-5, 8, 11-15, 18, 21, 22, 24 and 27-29 drawn to a vaccine containing an immunogenic fragment of Type B influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, aubclasses, 89, 350 and 69.1, 172.1, 320.1.
- VII. Claims 1-5, 9, 11-15, 19, 21, 22, 26-29 and 31, drawn to a vaccine containing an immunogenic fragment of Type A influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- VIII. Claims 1-5, 9, 11-15, 19, 21, 22, 26-29 and 31 drawn to a vaccine containing an immunogenic fragment of Type B influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- IX. Claims 1-5, 11-15, 20-22, 27-29 and 32, drawn to a vaccine containing an immunogenic fragment of Type A influenza virus and including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- X. Claims 1-5, 11-15, 20-22, 27-29 and 32, drawn to a vaccine containing an immunogenic fragment of TypeBinfluenzavirusand including one polypeptide, protein, microorganism and vector, classes 424,530 and 435, subclasses, 89, 350 and 69.1, 172.1, 320.1.
- XI. Claims 10, 33, 34 and 35, drawn to a vaccine containing two polypeptides, wherein the first polypeptide has SEQ ID No. 10.
- XII. Claims 33, 34 and 35, drawn to a vaccine containing two polypeptides, wherein the first polypeptide has SEQ ID No. 12.
- XIII. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 16.
- XIV. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 18.
- XV. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 20.
- XVI. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 22.

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- XVII. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 24.
- XVIII. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No.26.
- XIX. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 27.
- XX. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEO ID No. 28.
- XXI. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 29.
- XXII. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEO ID No. 30.
- XXIII. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 32.
- XXIV. Claims 33, 35 and 36, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 34.
- XXV. Claims 33, 35 and 37, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 14.
- XXVI. Claims 33, 35 and 38, drawn to a vaccine containing two polypeptides, wherein the second polypeptide has SEQ ID No. 57.
- XXVII. Claims 39 and 40 drawn to a vaccine containing three polypeptides.

and it considered that the International Application did not comply with the requirements of unity of invention (Rules 13.1, 13.2 and 13.3) for the reasons indicated below:

The claims of groups I-XXVII are drawn to multiple products which are not linked by a special technical feature so as to form a single inventive concept. PCT Rule 13.1 and Rule 13.2 do not provide for multiple products and methods within a single general inventive concept.